

NIH Public Access

Author Manuscript

Eur J Integr Med. Author manuscript; available in PMC 2014 April 01.

Published in final edited form as:

Eur J Integr Med. 2013 April 1; 5(2): 126–140. doi:10.1016/j.eujim.2012.11.002.

Advances in Integrative Nanomedicine for Improving Infectious Disease Treatment in Public Health

Iris R. Bell, MD PhD^{a,b,c,d,e,f,*}, Gary E. Schwartz, PhD^{b,c,e}, Nancy N. Boyer, RN NP PA-C⁹, Mary Koithan, RN PhD^{a,d,e}, and Audrey J. Brooks, PhD^{c,e}

^aDepartment of Family and Community Medicine, the University of Arizona College of Medicine, Tucson, AZ, USA

^bDepartment of Psychiatry, the University of Arizona College of Medicine, Tucson, AZ, USA

^cDepartment of Psychology, the University of Arizona, Tucson, AZ, USA

^dCollege of Nursing, the University of Arizona, Tucson, AZ, USA

^eDepartment of Medicine (Integrative Medicine), the University of Arizona College of Medicine, Tucson, AZ, USA

^fMel and Enid Zuckerman College of Public Health, the University of Arizona, Tucson, AZ USA

^gPrivate Practice, Rochester, NY USA

Abstract

Introduction—Infectious diseases present public health challenges worldwide. An emerging integrative approach to treating infectious diseases is using nanoparticle (NP) forms of traditional and alternative medicines. Advantages of nanomedicine delivery methods include better disease targeting, especially for intracellular pathogens, ability to cross membranes and enter cells, longer duration drug action, reduced side effects, and cost savings from lower doses.

Methods—We searched Pubmed articles in English with keywords related to nanoparticles and nanomedicine. Nanotechnology terms were also combined with keywords for drug delivery, infectious diseases, herbs, antioxidants, homeopathy, and adaptation.

Results—NPs are very small forms of material substances, measuring 1–100 nanometers along at least one dimension. Compared with bulk forms, NPs' large ratio of surface-area-to-volume confers increased reactivity and adsorptive capacity, with unique electromagnetic, chemical, biological, and quantum properties. Nanotechnology uses natural botanical agents for green manufacturing of less toxic NPs.

Discussion—Nanoparticle herbs and nutriceuticals can treat infections via improved bioavailability and antiinflammatory, antioxidant, and immunomodulatory effects. Recent studies

Conflict of Interest

^{*}Corresponding Author: Iris R. Bell, MD PhD, Department of Family and Community Medicine, The University of Arizona College of Medicine, 1450 North Cherry Avenue, MS 245052, Tucson, AZ 85719, Ph. 520-626-4188 (Cell 520-906-6767), FAX 520-749-4509, ibell@email.arizona.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Dr. Bell, Ms. Boyer, and Dr. Brooks serve as consultants to Standard Homeopathic Co./Hyland's Inc., a homeopathic manufacturer whose products were not used in the cited studies. Standard Homeopathic Co./Hyland's Inc. did not provide any funding for the current paper or publication costs.

demonstrate that homeopathic medicines may contain source and/or silica nanoparticles because of their traditional manufacturing processes. Homeopathy, as a form of nanomedicine, has a promising history of treating epidemic infectious diseases, including malaria, leptospirosis and HIV/AIDS, in addition to acute upper respiratory infections. Adaptive changes in the host's complex networks underlie effects.

Conclusions—Nanomedicine is integrative, blending modern technology with natural products to reduce toxicity and support immune function. Nanomedicine using traditional agents from alternative systems of medicine can facilitate progress in integrative public health approaches to infectious diseases.

Keywords

Nanomedicine; Drug delivery systems; Medicinal plants; Herbal medicine; Antioxidants; Homeopathy; Nanoparticles; Silica; Infectious disease treatment; Adaptation; Network medicine

Introduction

Infectious diseases continue to present a major public health challenge in both developed and developing countries. Even when modern conventional treatments such as antibiotics or antiretroviral drugs are available for a given type of infection, limitations of their usefulness can arise. Such limitations include emergence of antibiotic resistant bacteria, prohibitive costs and limited accessibility; and poor patient adherence issues. Conventional medical drugs have specific targets, but typically also cause significant side effects. Safety and riskbenefit analyses take on major relevance in mainstream health care decision making.

Whenever possible, integrative practitioners seek to find safer treatments to accomplish their clinical treatment objectives [1]. They often turn to alternative health care approaches for additional options. For example, the literature offers preclinical and clinical evidence in support of the effectiveness of traditional herbs and homeopathic remedies in treating individuals with various acute and chronic infectious and inflammatory conditions. In particular, alternative medical systems such as Ayurveda, Chinese herbalism, homeopathy, and naturopathy can show historical and observational trial evidence of good outcomes, greater safety, patient acceptance, accessibility and cost-savings over conventional drug treatments. Many alternative treatment modalities have multi-target effects [2], strengthening host defenses more than directly destroying infectious agents. Many herbs and homeopathic medicines act as adaptogens, i.e., nontoxic agents that increase the body's global ability to adapt to stress or environmental change without relying on specific local receptor targeting.

However, pragmatic, as well as sociopolitical, issues often confront integrative practitioners seeking to utilize these alternative types of treatments. Various barriers limit widespread inclusion of alternative therapies in public health programs for preventing and treating infections, including only a small number of pharmaceutical clinical trial studies [3, 4], with difficulty in setting reliable standards for purity and activity in the agents under investigation. Although certain herbs and nutriceuticals demonstrate a striking ability to stimulate immune system defenses in vitro or with certain forms of administration in animals, solubility and/or absorption from the gut *in vivo* can be poor. As a result, a practical challenge with some traditional botanical and nutriceutical agents is that the poor solubility and/or absorption prevent clinical effects of the desired magnitude for treating an infectious process as definitively as a conventional pharmacological agent may accomplish (but with even more side effects). Widespread adoption of herbs and other natural products for public health applications then is seen as impractical for a variety of reasons.

In addition, skeptics of homeopathy have historically raised doubts regarding its fundamental scientific plausibility because of the manner of preparing its medicines with a process of serial dilutions and succussions (vigorous shaking). Such critics assert that the regulated medicines could not be biologically medicine the effects that have been

resultant medicines could not be biologically producing the effects that have been extensively documented in case studies because of their bulk form dilution, sometimes seemingly past Avogadro's number of molecules. The skeptics may be both partially correct – and incorrect.

New empirical discoveries suggest that even though the *bulk form* source materials might not persist into the more "dilute" remedy potencies, *observable nanoparticle forms* of the source material and silica do persist across all homeopathic potencies, from "lower" to "higher" [5, 6]. The unique properties of nanoparticles would help account for many perplexing findings on homeopathic medicines and reposition homeopathy as a traditional form of adaptive nanomedicine for the organism as a whole network [7–9]. Understanding nanoparticles and their characteristics [10–12] may finally put to rest the skeptics' wellintentioned attacks, which are based on bulk form scientific assumptions, on the preparation methods for homeopathic medicines.

Surprisingly, advances in conventional drug targeting and safety [12], herbal product delivery into the body [13, 14] and the nature of homeopathic remedies [5, 6] all converge in the fields of nanotechnology and nanomedicine [15]. The purpose of this paper is to provide a brief introduction and overview of cutting-edge research on nanoparticles (NPs) and nanomedicine, with implications for improving safe and effective integrative treatments for infectious diseases using conventional drugs, botanical agents, nutriceuticals, and homeopathics. The discussion will include emerging research on nanoparticles and homeopathy as an exemplar traditional whole medical system. The data suggest potential physiological adaptive mechanisms in intracellular and systemic networks of the host, more than direct local pharmacological effects. Advances in cutting-edge research on nanotechnology-based drug delivery approaches, of which most integrative providers are not as yet aware, could facilitate major improvements in the capacity to deliver better and safer care for infectious diseases to larger populations.

Methods

The authors searched Pubmed articles in English with keywords related to nanoparticles, nanotechnology, and nanomedicine. The nanomaterial search terms were also combined with keywords for drug delivery, infectious diseases, specific herbs, herbal medicine, antioxidant, and homeopathy. The search revealed 55,697 articles on nanoparticles, 3,143 articles on nanomedicine, 57 articles on nanoparticles and herbs or herbal topics, 1,358 articles on nanoparticles and antioxidants, and 9 articles on nanoparticles, nanobubbles, and homeopathy or homeopathic. The term "low dose nanoparticle" yielded 507 citations. On mechanisms relevant to nanoparticle actions in infectious disease, the search showed 852 articles on hormesis, 201 articles on cross-adaptation, 2,889 articles on heat shock proteins and infection, and 1,020 articles on inflammasome. We focused primarily on papers published within the last 5 years, but also included highly relevant peer-reviewed research articles and reviews from any date identified in the literature search or as key citations in bibliographies of more recent papers. A core set of 79 papers provided the essential information to inform the development of the current paper.

Results

Nanomedicines in Drug and Herb Delivery

Nanoparticles are very small forms of material substances, measuring from 1 to 100 nanometers in diameter along at least one dimension [11]. Nanoparticles can form via physical, chemical, or biological methods, from either a top-down (e.g., milling or otherwise disrupting solid bulk forms of an insoluble substance to generate nanoparticles) [16–18] or bottom-up (e.g., molecular self-assembly of nanoaggregate conformations) approach [19, 20]. Both natural and synthetic NPs occur in the modern world [21, 22].

Nanotechnology is the applied engineering term for manipulating nanomaterials at the atomic and molecular scale [23]. The field encompasses nanomedicine, which strives to utilize nanotechnology to improve health care [15]. Various NPs have applications for diagnosis and treatment [24], but some can cause toxic effects on living systems [10, 22, 25]. Toxic NPs from the environment, for instance, are under study as possible etiological agents in autoimmune diseases, neurodegenerative disorders, and Crohn's disease [10, 21, 22]. Toxic NPs arise from natural and anthropogenic sources of occupational exposures, medicinal applications, and ambient air pollution, including volcanic activities, dusts or petroleum-based fuel combustion products [21, 22, 26]. Several recent reviews discuss the extensive evidence on the toxicity of certain nanoparticles, including adverse cardiopulmonary effects [21, 22, 26].

Generally, smaller NPs are more toxic to cells than larger NPs of the same source material [27, 28]. The state of the recipient living system (e.g., cancerous versus healthy cells) and higher versus lower dose levels [29, 30] also can affect the degree of nanotoxicity. For instance, one study demonstrated greater apoptotic effects of larger versus smaller calcium phosphate nanoparticles in osteosarcoma cells [31]. Several other studies have shown that certain nanoparticles exert selective apoptotic effects on specific breast, lung, liver, or brain glioma cancer cells but not on healthy cells [32–35]. Moreover, other NPs are benign and even activate antiinflammatory, antioxidant and immunostimulatory effects [36–43].

Because of their large surface to volume area, NPs possess unique, sometimes atom-like properties not seen in bulk form versions of the "same" material, e.g., a drug, metal, herb or environmental chemical pollutant [11, 44]. For example, nanoparticles readily cross cell membranes, including the blood-brain barrier [45], and gain access to cells, as well as translocate around the body via blood and lymph [10]. Their access to the inside of cells makes them an attractive tool for delivering treatment with drugs, herbs, and/or antioxidant nutriceuticals to intracellular pathogens [12, 46].

Armstead and Li [12] recently summarized the range of intracellular infectious diseases that nanomedicines may be more effective in treating than conventional bulk form drugs. Such diseases include tuberculosis, HIV (see also [47]), hepatitis C, salmonella [48], typhoid fever, candidiasis, leismaniasis (see also [49]), and malaria (see also [50–52]). For various reasons related to their small size, e.g., better solubility, absorption and uptake, nanoparticle-based medicines can get across cell membranes and reach specific targets more easily than bulk form agents. It is also possible to formulate certain nanoparticle drugs to release their active agent mainly when they are inside the cell, instead of dispersing drug prematurely or indiscriminately, thereby reducing side effect risks while optimizing dose [53].

The size and morphology of nanoparticles and the nanoclusters that are formed lead to different physico-chemical, biological, electromagnetic [54], magnetic [11, 55, 56], optical [57], thermal [58], and quantum [59] properties from those of the bulk forms of a given substance [10, 11]. Atoms and electrons lie closer to the surfaces of nanoparticles than in

bulk form materials and lead to markedly increased ability to adsorb DNA, proteins, drugs, herbs, lactose, and other nanoparticles onto their surfaces [18, 60–65]. Catalytic capacities and often magnetic properties, are acquired or markedly increased in NPs [11].

Specific cells can be targeted [66–69], effects of a given dose last longer [70], and therapeutic effects are magnified [32]. The dose levels of an agent needed to produce a given response can down shift as much as 1000-fold for the nanoparticle form versus the commercial bulk form, e.g., amount of an antigen needed in a vaccine to stimulate an immune response [71] – see also [72]. One study showed that the nano-form of an anti-tuberculosis drug mixture required only 3 doses, versus 45 doses of the "same" bulk form drugs, to produce a comparable elimination of tubercle bacilli in mice [70].

Table 1 lists exemplar studies of herbalfacilitated nanoparticle medicines drawn from Ayurveda, Chinese medicine, Western herbalism, homeopathic and nutriceutical systems. Preclinical studies indicate that the NP forms of antioxidant and anti-inflammatory herbs can exert heightened ability to stimulate innate immunity, mobilize cytokines, alter generation of reactive oxygen species (free radicals), and exert antibacterial, antiviral, and/or antifungal effects [64, 73–77]. For nutriceuticals with antioxidant, anti-inflammatory, and antimicrobial effects but poor gastrointestinal absorption or bioavailability, e.g., coenzyme Q10 [78–80], quercetin [81], curcumin [39, 82], resveratrol [42], or alpha-lipoic acid [83], nano forms can dramatically improve clinical utility.

For example, the antioxidant quercetin can exert beneficial effects against infections ranging from leishmania [84] and dengue virus type-2 [85] to rhinovirus [86] and influenza virus [87], but bulk forms are poorly absorbed from the gut. Making a lipid nanoparticle form of quercetin improves relative bioavailability 571.4% after oral administration in animals [88]. Another example is silymarin, an herb often used in integrative medicine to support liver function. Attaching silymarin to porous silica nanoparticles generates an orally-administered, sustained release product that maintains higher plasma concentrations of silymarin in animals, compared with bulk form commercial herbal tablets [89]. NP forms make it possible to take advantage of the significantly improved absorption across the gut and entry into target cells for better bioavailability at lower doses.

The ability to amplify these beneficial effects of herbs or nutriceuticals for treatment of infectious diseases by attaching them to nanoparticle carriers [64, 90] and delivering them directly into infected cells throughout the body is a significant advantage. NPs also have other potential public health applications. For instance, compared with crude extract controls, silver nanoparticles synthesized using *Euphorbia hirta* plant leaf extract exert significantly greater larvicidal and pupicidal effects in mosquitoes that transmit malaria [91]. The nanotechnology approach even has potential application to reduce spoilage in antioxidant food packaging films. For instance, the nanoparticle forms of curcumin and ascorbyl dipalmitate exhibit higher antioxidant activities on four different assays than their respective usual bulk forms in water [92].

As a result, nanomedicine researchers have discovered that therapeutic NPs can serve as more effective drug delivery vehicles than conventional bulk form drugs [93, 94]. Empirical studies demonstrate that using NPs for delivery of drugs [95, 96] and/or herbs [14] can (a) better reach their intended targets in the cells of the body; and (b) reduce the total amount of an agent needed to accomplish a therapeutic effect [74]. NP herb or drug forms can not only bypass or dramatically improve upon the poor gastrointestinal absorption and/or solubility problems that limit the usefulness of many different bulk form agents [15, 97], but they can also cross the blood-brain barrier for enhanced delivery into the central nervous system, e.g., for antiretroviral drugs in HIV/AIDS [98]. Inhalation administration routes for certain

nanomedicines and nanoherbs may also be more acceptable and accessible for the general population [99] than injections or pills [100]. In turn, these properties of nanodrugs and nanoherbs can translate into (a) reduced side effect risks; (b) greater effectiveness; and (c) lower costs. These advantages would have a favorable impact on public health.

The Role of Natural Substances in Nanoparticle Manufacturing: Reducing Toxic Risks

Nanoparticles for nanomedicine drug delivery applications include NPs from gold and silver as well as silica [101], calcium and magnesium phosphate [102], carbon [103], chitosan (a linear polysaccharide from the exoskeletons of crustraceans or cell walls of fungi) [82], alginate (a natural polymer) [104], liposomes or lipid nanocapsules [13, 105]. A major limitation in translating advances in medical nanotechnology to the clinical and public health arenas has been uncertainties about the potential cumulative toxicity of the manufacturing processes and/or the drug delivery NPs themselves, especially the metalbased NPs. For example, silver or copper NPs can release toxic ionic forms that can accumulate in cells [106, 107]. Researchers are concerned about the potential toxic accumulation of therapeutic gold or silver NPs inside cells, such as liver and spleen, given their lack of biodegradability [108].

Identifying less toxic and/or biodegradable types of nano-forms, e.g., using lipid-based [43] or calcium phosphate NPs [102], and using lower NP doses [71, 109, 110] are among the current strategies for reducing toxicity in medicinal applications [70, 104]. Silica NPs [24, 26, 111, 112] may offer a potentially safer option in infectious disease treatment [12], as evidence suggests that they can enhance immune reactivity but biodegrade without accumulation [113]. Preliminary data also indicate a similarly lower cytotoxicity for some types of carbon nanotubes [114] and perhaps calcium carbonate [62] or calcium or magnesium phosphate [60, 61]. Chitosan NPs are considered benign [50, 82], and lipid-based NPs are one of the preferred ways to deliver antimalarial agents to infected cells [51].

In addition, chemicals used in the manufacturing process also adsorb, along with the intended drug or herb, onto the surface of the NPs [115]. Consequently, nanotechnology engineers are increasingly seeking more ecofriendly ways to manufacture nanoparticles that avoid or limit reliance on toxic chemical methods [116]. The adsorbed materials can modify the properties, effects, and/or toxicity of the NPs.

For instance, nanotechnology engineers have developed novel biological methods for making NPs. They are turning to aqueous plant extracts and natural phytochemicals rather than synthetic chemicals to generate the gold or silver nanoparticles for nanomedicine applications [117]. In the latter context, investigators have successfully used *Zingiber officinale, Mirubilis jalapa* flower, *Stevia rebaudiana, Cinnamon camphora* leaf, *Cassia fistula* bark [116], *Hibiscus rosa sinensis* [118], tea [119], *Rhizophora mucronata* mangrove leaf [52], *Gnidia glauca* flower [115], *Phytolacca decandra* [120], *Thuja occidental, Hydrastis canadensis, and Gelsemium sempervirens* [117]. Relying on botanical agents to manufacture state-of-the-science NPs makes this aspect of medical nanotechnology truly "integrative."

Homeopathy: 200 Years of Low-Dose Natural Nanomedicines for Infectious Diseases?

Homeopathic Medicines as Nanoparticles—Many therapeutic applications of NPs used as conventional drugs are in preclinical testing phases. Ironically, one of the most controversial systems of alternative medicine, homeopathy, could turn out to be one of the oldest and demonstrably safest [121–124] forms of nanoparticle-based [5, 6] treatment already used worldwide for infectious diseases. That is, one "top-down" way in which modern nanotechnology makes nanoparticles from bulk materials is simply by grinding or

milling insoluble substances for long periods of time [16–18, 125]. This type of milling procedure is a sophisticated mechanized approach [16, 18, 19] to what homeopathic manufacturers have done by hand or simple machines for over 200 years, i.e., trituration of bulk form plant, mineral, or animal source materials in lactose, with mortar and pestle, to make "dry" dilutions of particles [126, 127]. Consequently, the lactose trituration method of manufacturing homeopathic remedies, especially for the lowest potencies of source agents, begins to blur the line between herbal nanomedicines and homeopathic medicines. Nanomedicines include gold or silver [74, 128], silica, calcium or magnesium phosphates as drug or gene delivery vehicles [60, 61, 129], and nanovaccines [71, 130], all of which overlap with the source materials for common low potency homeopathic medicines [131].

To make liquid homeopathic medicine potencies, the milled or ground materials for insoluble sources or ionic salts for soluble sources are serially diluted and succussed in ethanol-water solutions. During liquid dilution manufacturing, the succussions after each dilution step cause intense fluid turbulence from vigorous shaking, agitation, or physical pounding of the glass container against a hard surface [132]. In modern nanotechnology, this type of fluid turbulence produces particle collisions and shearing forces that break off increasingly smaller particles [133].

Both homeopathic [6, 134–136] and non-homeopathic [137] pharmacy research studies indicate that glass can release measurable but variable amounts of silica precursors and silica nanoparticles into solution. Agitation causes glass-derived silica nanoparticles to accelerate aggregration of protein molecules in solution with them [137]. Also, once in aqueous solution at room temperature, silica precursors can form self assembled crystalline silica structures in the presence of plant extracts [138].

Even at liquid dilutions seemingly past Avogadro's number (e.g., in homeopathic potencies of 12C or 24X, beyond Avogadro's number of 6×10^{23} molecules, where C=bulk dilution factor of 1/100, done 30 times for 30C or 200 times for 200C and then succussed 10 or more times after each "dilution" step), modern electron microscopy and other laboratory analytic methods now show that homeopathic metal-derived medicines still contain source nanoparticles, transferred from dilution to dilution [5]. Using scanning and transmission electron microscopy, a different research group subsequently demonstrated nanostructure forms in three different homeopathic plant medicines at potencies from 1C to 15C [6]. A third research group recently reported the biosynthesis of silver nanoparticles had different sizes and biological effects as a function of the specific plant tincture with which they were manufactured [117].

Homeopathic medicines likely contain source particles adsorbed onto lactose particles in lower potencies [18, 63] or silica nanoparticles at higher potencies [73, 134–136, 139]. Variations in dilution and succussion procedures may contribute to the variability in manufacturing results [5, 73, 132]. The nanoparticles would include any metal or mineral-derived source materials [5] or organic plant [64] or animal source materials, as well as lactose [18], silica [6, 134–136, 139] or polypropylene [73] nanocrystals with any plant or animal source proteins and/or nucleic acids adsorbed. Lactose can even adsorb biologically-active animal protein nanoparticles created by electrospraying [63].

Alternatively, modern homeopathic manufacturing methods sometimes rely on vortexing or sonication rather than manual shaking to accomplish the mixing and succussion, as well using polypropylene tubes rather than the traditional glass vials [73, 135, 136]. The type of succussion and the type of material in the walls of the vial in which the remedy is succussed would measurably affect the properties of the resultant homeopathic medicine [73, 136]. In

addition, nanoparticles undergo spontaneous aggregration into larger particles (which would alter their properties) if left undisturbed for periods of time, a thermodynamic process termed Ostwald ripening [140]. Sonication disperses aggregates of nanostructures that can form after wet-grinding procedures alone [141], thereby likely changing the sizes and properties of the final product [58, 142].

Such findings suggest the possibility that not only the glassware, but also the type, number or duration and force, as well as recency of the succussion procedures could result in biologically meaningful variations in the properties of homeopathic medicines in a specific batch or dose. Differences in ethanol concentration, pH, and/or temperature can also contribute to significant variations in the sizes, shapes and, thus, properties of nanostructures [20, 142]. The well-known reproducibility problems in homeopathic research could relate in part to these multiple variables that can affect nanoparticle formation in complex, nonlinear ways.

Furthermore, the glass-derived silica may be much more than an "artifact" in homeopathic medicines [9]. A body of recent research suggests that nanosilica could play a key role as (a) drug delivery vehicles for homeopathic plant or animal source materials that adsorb onto its surfaces [20, 138, 143, 144]; (b) vehicles for epitaxial transfer and even memory of electromagnetic [145] and/or structural information using the specific remedy source materials as structural template "seeds" and biological guides for bottom-up, self-assembled formation of silica nanostructures [20, 138, 146, 147]; and (c) nonspecific biological amplifiers of specific antigen or remedy source effects on immune cells and pathways [112, 130, 148–150]. For example, adding nanosilica significantly increases the apoptotic and growth arrest effects of a traditional snake venom medicine on human breast cancer cells [32]. In brief, while glass-derived nanosilica would not be necessary to make a homeopathic remedy, the silica, when present, would potentially enhance the bioavailability and biological effects of remedy source-specific nanoparticles in the medicines.

Taken together, the data suggest that, homeopathic medicines may be low doses of mineral, plant, and/or animal source nanoparticles. Nanotechnology studies have shown that trituration [151], drug crystal milling [16, 19, 125, 152], and/or various succussion methods [73] at room temperature can release nanoparticles and nanoaggregates of silica (or polypropylene polymers) and/or source material into colloidal solution [73, 134, 136, 139]. The physician-chemist founder of homeopathy, Samuel Hahnemann, deserves credit for what appears to have been the first description of a practical method for making and safely administering nanomedicines. He interpreted his observations on the ability of trituration and succussion to release unique medicinal properties from bulk form materials within the vitalistic and spiritual conceptual framework of his era [126]. However, Hahnemann's life (1755–1843) occurred at a time that predated key scientific discoveries and acceptance of the atomic and molecular nature of matter, as well as the unique characteristics of nanoparticles themselves. Rather than rejecting the entire field of homeopathy for its originally vitalistic *interpretations*, it is time for modern scientists to examine seriously the nanomedicine implications of Hahnemann's *empirical findings* for integrative health care.

That is, such homeopathic preparation steps could represent a crude but effective, top-down technique for mechanically generating nanoparticles [5, 6]. In nanofluids, different amounts of sonication time by itself can increase the size and morphology of nanoparticle and nanocluster aggregates that form from the "same" material [58]. In solution, the enhanced reactivity of NPs can lead to spontaneous changes in shape, NP aggregations and self-assembly [11, 140]. Similarly, homeopathic medicines stored in liquid form at room temperature change their physico-chemical properties over time [153]. Different NPs and aggregates possess different physico-chemical properties from one another because of their

different sizes and shapes [10, 11], even though the original bulk form source was the same substance. [58]. Particle structure, adsorption, aggregation, and self assembled network organization become as important as material source composition in affecting properties at the nano level of scale [147].

Homeopathic Medicines in Infectious Disease Treatment: Cross-Adaptation and Hormesis—Prior research on the basic science of homeopathy [153] converges with evidence from the nanomedicine literature to suggest potential mechanisms of action for low doses of NPs in treating infections. Nanoparticles can exhibit hormetic dose-response patterns [154, 155]. Hormesis is a well-documented nonlinear physiological and cellular phenomenon of adaptation [156–158]. In hormesis, lower doses of a given substance stimulate, whereas higher doses of the same substance inhibit, function in a complex adaptive system [159–164]. The hormetic dose-response range occurs below the noobserved-adverse-effect-level (NOAEL) [165]. Studies suggest that at least some NPs can cause hormesis in low doses [154, 166].

Convergent expert opinion is that hormesis is a nonlinear adaptive, not pharmacological, process that the patient or organism as a complex adaptive system or network generates [154, 167]. The adaptations begin in the cellular defense networks and interact with the other networks of the organism [160, 161, 163, 164, 168–170]. Relevant to their capacity to exert biological effects, as low dose, but highly-catalytic nanoparticles, homeopathic medicines are able to initiate (a) the biphasic dose-response relationship of hormesis [154, 171]; and (b) endogenous time-dependent response amplification and biphasic oscillation processes [7, 8, 172–174]. Integrative medicine researchers have focused on hormesis and complex adaptive systems as potential mechanisms for homeopathic medicine actions at low doses [175]. However, non-homeopathic integrative interventions at relatively low bulk form doses, such as herbs and antioxidant nutriceuticals, also engage hormetic cellular adaptive processes to strengthen host defenses [164, 176]. Nanoforms of nutriceuticals would lower the dose required to evoke the adaptive or hormetic effects within the host.

Given the evidence that homeopathic medicines may be NPs, infectious diseases such as malaria and other epidemics were among the earliest public health problems for which homeopaths reported good outcomes since the inception of this over 200-year old field [126, 127, 177]. Consistent with this claim, animal studies demonstrate an ability of homeopathic *Eupatorium perfoliatum* or *Arsenicum album* to inhibit Plasmodium parasite multiplication [178]. Combinations of other common homeopathic remedies in mice (i.e., *Bryonia alba, Thuja occidentalis, Aconitum napellus, Arsenicum album, and Lachesis*) can inhibit experimental infection with Leishmania amazonensis [179], as can other types of nanoparticles [49, 180–182].

Homeopathic medicines made from botanical sources also exert antiviral effects against multiple human pathogenic respiratory viruses in vitro [183]. Such medicines combine low potencies of *Aconitum napellus, Bryonia alba, Eupatorium, Phosphorus, and Lachesis.* Preliminary clinical trial evidence suggests homeopathic treatment can have beneficial adjunctive benefits in people with HIV/AIDS [184]. Homeopathic diamond given adjunctively to conventional drugs, is one of many remedies anecdotally reported helpful for HIV/AIDS patients in Africa [185] (http://www.homeopathyforhealthinafrica.org/, accessed 06/05/12). Low, but not high, doses of conventional nanodiamond NPs can upregulate phagocytic antibacterial activity of mouse cells *in vitro* [155]. The effects of conventional nanodiamond on antiviral activity are not as yet studied but appear feasible to assess [186, 187].

A large scale public health intervention with prophylactic homeopathically-prepared oral doses of diluted and succussed, inactivated leptospirosis bacteria in 2.3 million people in Cuba demonstrated significant reductions in disease incidence compared with non-intervention regions during a high-risk rainy season [188]. Notably, in conventional nanomedicine research, a one-dose nanoparticle-based vaccination by nasal administration has been shown effective in an animal model of a different bacterial infection, Yesinia pestis (plague) [189]. NP formulations can increase the ability of low doses of a given agent to translocate around the body [190] or stimulate immune responses [71, 72, 155].

As a complete clinical method for administering nanoparticles at nontoxic doses, homeopathy offers significant advantages. In contrast with conventional drugs and many bulk form herbs, drug-drug and drug-herb interactions, are not a problem with homeopathic medicines. Moreover, the likely hormetic adaptive mechanisms of action for homeopathic nanoparticles typically place their dose-response curves into the nontoxic range. Homeopathic treatment also has the advantage of lowering costs via reduced use of symptomatic conventional drugs [121]. Homeopathic medicines themselves are much less expensive, and, for many conditions, require fewer doses than conventional drugs, advantages that can also lower costs and improve patient adherence.

Modern observational, comparative effectiveness, and efficacy studies also indicate a strong track record for homeopathy of faster onset clinical improvements in mild to moderate common acute infections of the upper respiratory tract (e.g., colds, influenza - [191–193]) and ear [194–200]). In vitro, homeopathic remedies made from botanical sources demonstrate antiviral effects and the ability to stimulate patterns of change in pro- and anti-inflammatory cytokine release [183, 201, 202]. Even while dismissing the favorable findings, the otherwise negative (albeit highly flawed - [203–205]) Shang et al's metaanalysis reported a subanalysis of homeopathy efficacy studies restricted to infectious diseases that revealed a strongly significant benefit of homeopathic treatment [206]. In conventional nanomedicine research, nanoparticle-based viral vaccines, gold and other types of NPs are also efficacious against influenza virus [207–209]. Although more clinical research is clearly needed, the evidence exists to support integrating nanoparticle research with homeopathic medicine, in the process of establishing a hormetic or adaptive nanomedicine.

The way in which homeopaths prescribe their medicines differs significantly from conventional pharmacological dosing and relies on small quantities below toxic levels [210]. Homeopathic dosing utilizes not only low doses, but also intermittent pulsed timing of doses, i.e., spaced intermittently in time [211]. Their likely mode of action relates to the ability of low doses of exogenous agents or stress itself to initiate persistent, self-amplified adaptive changes in physiological self-regulation via hormesis and endogenous time-dependent sensitization in the host [7, 8, 174, 212–215]. The low doses of activated nanoparticle forms, by working within the hormetic dose-response range [160–162], would activate broad, endogenous non-pharmacological mechanisms involved in compensatory adaptations to exogenous stressors and stimuli, i.e., the allostatic stress response network [112, 167–169, 216–219]. The stress response network includes heat shock proteins [160–162, 220],cytokines [201, 221, 222], and reactive oxygen species [39]. As a result, pulsed low doses of NPs (i.e., homeopathy) would rely mainly on modulating network-based adaptive, rather than specific local pharmacological, mechanisms of action [7–9].

Many modern nanoformulations are still in preclinical testing phases. However, two centuries of experience with homeopathic medicines have demonstrated that this type of nanoparticle, administered in low intermittent doses, has an exceptionally positive safety profile for public health applications, even in widespread use across large populations [121].

Homeopathic medicines offer a real-world exemplar of low dose, non-pharmacological nanoparticle treatment within an already established whole system of complementary and alternative medicine (CAM). It is possible that the historic term "homeopathy" will ultimately be replaced with a more pragmatic, scientifically accurate, and neutral term – e.g., adaptive network nanomedicine [9].

There is now an evidence-based, scientifically testable model for homeopathic remedy effects that overlaps with the empirically-documented properties of nanoparticles [7–9]. Homeopathic remedies appear to contain source and silica nanoparticles and are not inert placebos. NP forms of a given bulk form source are highly reactive and catalytic, prone to adsorbing other nanomaterials on their surface. At moderate or high doses, nanoparticles can trigger stronger pharmacological effects; at low doses, NPs mobilize non-pharmacological, biological adaptation, hormesis and endogenous amplification mechanisms [8, 155, 173, 218]. Together with the state of the organism at the time of administration, the intensity or magnitude of the dose helps determine the direction of adaptive change [7, 8, 174, 223]. The enhanced reactivity of NPs could shift the dose-response range for expressing hormesis even lower than with low bulk form doses [154]. This scientifically-grounded perspective puts the substantial body of preclinical and clinical data on homeopathy and other types of nanoparticles from natural sources in a new light.

Discussion

The Stress Response and Cellular Defense Network and Adaptation

Viewing the organism as a complex adaptive network of networks [167–169, 214, 216, 224–227] is a good starting point for developing preventive strategies and more effective integrative treatments for infectious diseases. Periodic mild stressors from multiple different categories, including herbal and nutritional [164, 228], can all induce hormesis and promote better health and survival [167, 229]. In fact, mild stressors both prepare the organism to fend off higher intensity stressors of the same or cross-adapted type in the future and/or to recover from the adverse effects of prior higher intensity stressors, e.g., infections, once established [156, 160–162].

Complex adaptive systems are self-organizing networks [168, 169] whose functions are shaped by changes in the environment [230, 231]. Adaptations begin at the interface with the environment, within the stress response network, which regulates immune, endocrine, nervous system, and metabolic functions [217, 232]. Changes in the stress response network [169, 214] cascade into an amplified and extensive set of adaptations via their interconnections with organs [227] and the host organism as a whole [216, 225, 226, 230]. The direction of adaptive changes is plastic and bipolar. As in drug-related hormesis, lower intensity biological or psychological stressors also induce persistent changes in systemic responsivity in the opposite direction to those of higher intensity stressors [212]. Global and local levels of systemic organization interact and feedback information to modulate each other's functions and organizational dynamics [225, 233, 234]. Under stress, cellular networks can change their linking pattern structure to improve their chances of survival, but can recover an unstressed linkage pattern when the stress subsides [168, 169].

Local Cellular Defense Mechanisms in Host Adaptation

Effectors in the molecular stress response networks of cells include heat shock proteins, chaperone proteins, lysosomes, DNA repair enzymes, FoxO and other molecular responders to reactive oxygen species (free radicals), sirtuins, and cytokines [167]. Serving as mild environmental stressors for the host organism, natural source, non-drug NPs of plant, mineral, or animal source would modulate and strengthen host adaptive capacities and

resilience [160–162], apart from conventional pharmacological, symptom-suppressing mechanisms. NPs from various natural products, with their enhanced bioavailability, act as hormetins to promote stronger neuroimmune function in the host organism [176, 235–237]. For example, homeopathic NPs can elicit cross-adapted and cross-sensitized hormetic heat shock protein response patterns [160, 161, 220] that would, among other effects, induce innate and adaptive immunity to combat infections and other diseases [238].

One specific example of how either infectious organisms (e.g., bacteria or viruses) or nonviral nanoparticles can initiate such a cascade of adaptive biological events derives from data on the intracellular inflammasomes. Inflammasomes are a multi-protein complex that serves as sensors for microbial DNA patterns and other environmental "danger signals" for the body [148, 239]. A variety of exogenous stimuli, including infectious agents, nanoparticles, and non-infectious crystals of silica, alum, or urate [110, 148, 240, 241], mobilize the immune response by activating the intracellular inflammasomes. Activation of inflammasomes leads to the release of proinflammatory cytokines such as interleukin 1 β along with other interconnected regulatory elements of the stress response network [112, 148, 239, 241–243]. Interleukins can then modulate central nervous system function, evoke fevers and inflammation, and change the host's mood and energy levels [244].

In the setting of established pathogen infection, low dose hormetic nanoparticles salient to the organism's pre-existing adaptational state would signal that a new, low intensity danger or reactive stressor [219], i.e., nontoxic doses of the herbal, nutriceutical, or homeopathic nanoparticles, have entered the body [148, 239, 241, 245]. Then the low dose nanoparticles and nanocrystallites would trigger the beneficial adaptive mechanisms [163, 164, 176], by serving as a low dose "danger" or "alerting" signal to bodily cells in the host to make changes that would lead, for instance, to enhanced immune activation and/or free radical attacks against intracellular infectious agents.

In additional to bidirectionality, the adaptational networks of the body appear to have substantial capacity for cross-adapted and/or cross-sensitized responses [8, 9]. As a result, a stressor from one category can initiate adaptations that modify the reactions to a subsequent stressor from an entirely different category [174, 246, 247]. Thus, adapting to anoxia may improve subsequent ability to tolerate extreme cold temperatures [247]. For instance, repeated intermittent exposures to psychological stress or sucrose ingestion or formaldehyde inhalation may heighten the magnitude or even change the direction of subsequent reactions to amphetamine or cocaine [213, 248–251]. Thus, administering nanoparticle hormetins in low dose has the potential for mobilizing endogenously amplified changes that can improve rather than impair the ability to overcome an infection or other types of stress.

Summary and Conclusions

Nanoparticle research has produced substantial progress toward improving natural product and drug delivery methods for treatment of infectious diseases. Nanomedicine is already integrative, blending state-of-the-science manufacturing technology with herbal and botanical sources to reduce toxicity. NPs offer the promise of more efficient and targeted treatments in a range of infectious diseases, especially intracellular pathogens with important public health implications, such as tuberculosis, HIV/AIDS, leishmania, and malaria [12]. Nanoparticle forms facilitate use of oral and nasal modes of administration [252, 253]. Isopathic public health interventions to reduce epidemics with diluted and succussed but attenuated oral low dose infectious agents, modeled after the leptospirosis project in Cuba [188], appear feasible and merit additional evaluation. At the practical level, modern nanoparticle formulations can lower drug, vaccine, herb, and nutriceutical doses and release their active agent only once inside target cells, thereby lowering drug-drug interactions and side effects [70, 104, 207, 254–256]. The potential benefits are improved outcomes and safety as well as lowered treatment costs as a result of reduced dose levels and oral or inhalation administration. Integrative treatments that support the patient's adaptive networks will lead to functional self reorganization of the patient as a biological network [225]. The result can be host expression of a more robust cellular defense system against infections and other stressors [163, 164, 231]. The advantage for public health is to reduce the risks of fostering drug-resistant organisms.

That is, not only the patient, but also the infectious organisms are complex adaptive systems. Conventional drug treatments for infections mainly ignore the host and, by directly attacking the infectious agent, stimulate dynamical adaptations in the bacteria, viruses, or fungi as complex adaptive systems. As a result, the adapting infectious agent self-reorganizes and evolves to overcome the adverse effects of the drugs for its own survival [168, 169], i.e., develop drug resistance. Thus, wider adoption of integrative strategies discussed in this paper to reduce inappropriate use of conventional antibiotic, antiviral, or antifungal drugs could substantially contribute to improvements in public health.

The development of biodegradable and nontoxic NPs from natural sources constitute an essential advance for translation from bench to integrative clinical application. Traditional herbs from Ayurveda or Chinese medicine [14, 40], as well as antioxidant nutriceuticals [42, 257, 258] that can strengthen immune function and improve resistance to infection are better absorbed and more active in nanoparticle than in ordinary bulk form. Homeopathic remedy (hormetic) nanoparticles offer an over 200 year, real world precedent for the pragmatic utility, safety, and cost-effectiveness of low dose adaptive network nanomedicine [7, 9] for large segments of the population, including in epidemics. The primary integrative therapeutic goal is to stimulate improved adaptive resilience in the host organism as a complex network.

Further research should focus on defining best practices for manufacturing, distributing, and administering nanoparticle-based integrative treatments using natural products for infectious diseases. Given limitations of conventional antibiotic drugs from the emergence of treatment-resistant organisms, developing safe and effective nanomedicines from natural products that bolster host resistance and self healing mechanisms from infections should be a priority for new funding initiatives. Nanomedicine using traditional agents from alternative systems of health care represents an important and timely opportunity for progress in promoting integrative public health in both developed and developing nations.

Acknowledgments

Funding

This study was supported in part by National Center for Complementary and Alternative Medicine grant T32 AT01287 (PI: IRB).

References

- 1. Rakel, D. Integrative Medicine. 3rd Edition. Philadelphia, PA: Saunders; 2012.
- Csermely P, Agoston V, Pongor S. The efficiency of multi-target drugs: the network approach might help drug design. Trends Pharmacol Sci. 2005; 26:178–182. [PubMed: 15808341]
- Bell IR, Koithan M, Pincus D. Research methodological implications of nonlinear dynamical systems models for whole systems of complementary and alternative medicine. Forschende Komplementarmedizin und Klassische Naturheilkunde. 2012; 19:15–21.

- Fønnebø V, Grimsgaard S, Walach H, Ritenbaugh C, Norheim AJ, MacPherson H, Lewith G, Launsø L, Koithan M, Falkenberg T, Boon H, Aickin M. Researching complementary and alternative treatments--the gatekeepers are not at home. BMC Med Res Methodol. 2007; 7:7. [PubMed: 17291355]
- Chikramane PS, Suresh AK, Bellare JR, Kane SG. Extreme homeopathic dilutions retain starting materials: A nanoparticulate perspective. Homeopathy. 2010; 99:231–242. [PubMed: 20970092]
- 6. Upadhyay RP, Nayak C. Homeopathy emerging as nanomedicine. International Journal of High Dilution Research. 2011; 10:299–310.
- Bell IR, Koithan M. A model for homeopathic remedy effects: low dose nanoparticles, allostatic cross-adaptation, and time-dependent sensitization in a complex adaptive system. BMC Complement Altern Med. 2012 in press.
- 8. Bell IR, Koithan M, Brooks AJ. Testing the nanoparticle-allostatic cross-adaptation-sensitization model for homeopathic remedy effects. Homeopathy. 2012 in press.
- 9. Bell IR, Schwartz GE. Adaptive network nanomedicine: an integrated model for homeopathic medicine. Frontiers in Bioscience (Elite Ed). 2012 in press.
- Buzea C, Pacheco II, Robbie K. Nanomaterials and nanoparticles: sources and toxicity. Biointerphases. 2007; 2:MR17–MR71. [PubMed: 20419892]
- Roduner E. Size matters: why nanomaterials are different. Chemical Society Reviews. 2006; 35:583–592. [PubMed: 16791330]
- Armstead AL, Li B. Nanomedicine as an emerging approach against intracellular pathogens. Int J Nanomedicine. 2011; 6:3281–3293. [PubMed: 22228996]
- Devi VK, Jain N, Valli KS. Importance of novel drug delivery systems in herbal medicines. Pharmacogn Rev. 2010; 4:27–31. [PubMed: 22228938]
- Huang S, Chang WH. Advantages of nanotechnology-based Chinese herb drugs on biological activities. Curr Drug Metab. 2009; 10:905–913. [PubMed: 20214585]
- Jia L. Nanoparticle Formulation Increases Oral Bioavailability of Poorly Soluble Drugs: Approaches Experimental Evidences and Theory. Curr Nanosci. 2005; 1:237–243. [PubMed: 19865587]
- DeCastro, CL.; Mitchell, BS. Nanoparticles from mechanical attrition. In: Baraton, MI., editor. Synthesis, Functionalization, and Surface Treatment of Nanoparticles. Valencia, CA: American Scientific Publisher; 2002. p. 1-15.
- Merisko-Liversidge E, Liversidge GG. Nanosizing for oral and parenteral drug delivery: a perspective on formulating poorly-water soluble compounds using wet media milling technology. Adv Drug Deliv Rev. 2011; 63:427–440. [PubMed: 21223990]
- Caron V, Willart JF, Lefort R, Derollez P, Danede F, Descamps M. Solid state amorphization kinetic of alpha lactose upon mechanical milling. Carbohydr Res. 2011; 346:2622–2628. [PubMed: 21983262]
- 19. Verma S, Gokhale R, Burgess DJ. A comparative study of top-down and bottom-up approaches for the preparation of micro/nanosuspensions. Int J Pharm. 2009; 380:216–222. [PubMed: 19596059]
- Belton DJ, Deschaume O, Perry CC. An overview of the fundamentals of the chemistry of silica with relevance to biosilicification and technological advances. FEBS J. 2012; 279:1710–1720. [PubMed: 22333209]
- Gwinn MR, Vallyathan V. Nanoparticles: health effects--pros and cons. Environ Health Perspect. 2006; 114:1818–1825. [PubMed: 17185269]
- Seaton A, Tran L, Aitken R, Donaldson K. Nanoparticles, human health hazard and regulation. J R Soc Interface. 2010; 7(Suppl 1):S119–S129. [PubMed: 19726441]
- 23. Cao, G.; Wang, Y. Nanostructures and Nanomaterials: Synthesis, Properties, and Applications. 2nd Edition. New Jersey: World Scientific; 2011.
- 24. Ambrogio MW, Thomas CR, Zhao YL, Zink JI, Stoddart JF. Mechanized Silica Nanoparticles: A New Frontier in Theranostic Nanomedicine. Acc Chem Res. 2011
- 25. Singh N, Manshian B, Jenkins GJ, Griffiths SM, Williams PM, Maffeis TG, et al. NanoGenotoxicology: the DNA damaging potential of engineered nanomaterials. Biomaterials. 2009; 30:3891–3914. [PubMed: 19427031]

- 26. Winnik FM, Maysinger D. Quantum Dot Cytotoxicity and Ways To Reduce It. Acc Chem Res. 2012
- Napierska D, Thomassen LC, Rabolli V, Lison D, Gonzalez L, Kirsch-Volders M, et al. Sizedependent cytotoxicity of monodisperse silica nanoparticles in human endothelial cells. Small. 2009; 5:846–853. [PubMed: 19288475]
- Passagne I, Morille M, Rousset M, Pujalte I, L'Azou B. Implication of oxidative stress in sizedependent toxicity of silica nanoparticles in kidney cells. Toxicology. 2012; 299:112–124. [PubMed: 22627296]
- 29. Sayes CM, Wahi R, Kurian PA, Liu Y, West JL, Ausman KD, et al. Correlating nanoscale titania structure with toxicity: a cytotoxicity and inflammatory response study with human dermal fibroblasts and human lung epithelial cells. Toxicol Sci. 2006; 92:174–185. [PubMed: 16613837]
- 30. Song M, Yuan S, Yin J, Wang X, Meng Z, Wang H, et al. Size-dependent toxicity of nano-C60 aggregates: more sensitive indication by apoptosis-related Bax translocation in cultured human cells. Environ Sci Technol. 2012; 46:3457–3464. [PubMed: 22352688]
- Shi Z, Huang X, Liu B, Tao H, Cai Y, Tang R. Biological response of osteosarcoma cells to sizecontrolled nanostructured hydroxyapatite. J Biomater Appl. 2010; 25:19–37. [PubMed: 19726533]
- 32. Al-Sadoon MK, Abdel-Maksoud MA, Rabah DM, Badr G. Induction of Apoptosis and Growth Arrest in Human Breast Carcinoma Cells by a Snake (Walterinnesia aegyptia) Venom Combined With Silica Nanoparticles: Crosstalk Between Bcl2 and Caspase 3. Cell Physiol Biochem. 2012; 30:653–665. [PubMed: 22854437]
- 33. Lim KJ, Bisht S, Bar EE, Maitra A, Eberhart CG. A polymeric nanoparticle formulation of curcumin inhibits growth, clonogenicity and stem-like fraction in malignant brain tumors. Cancer biology & therapy. 2011; 11:464–473. [PubMed: 21193839]
- Harhaji L, Isakovic A, Raicevic N, Markovic Z, Todorovic-Markovic B, Nikolic N, et al. Multiple mechanisms underlying the anticancer action of nanocrystalline fullerene. Eur J Pharmacol. 2007; 568:89–98. [PubMed: 17560995]
- Akhtar MJ, Ahamed M, Kumar S, Khan MM, Ahmad J, Alrokayan SA. Zinc oxide nanoparticles selectively induce apoptosis in human cancer cells through reactive oxygen species. Int J Nanomedicine. 2012; 7:845–857. [PubMed: 22393286]
- Nair HB, Sung B, Yadav VR, Kannappan R, Chaturvedi MM, Aggarwal BB. Delivery of antiinflammatory nutraceuticals by nanoparticles for the prevention and treatment of cancer. Biochem Pharmacol. 2010; 80:1833–1843. [PubMed: 20654584]
- Burnett ME, Wang SQ. Current sunscreen controversies: a critical review. Photodermatol Photoimmunol Photomed. 2011; 27:58–67. [PubMed: 21392107]
- Chonpathompikunlert P, Fan CH, Ozaki Y, Yoshitomi T, Yeh CK, Nagasaki Y. Redox nanoparticle treatment protects against neurological deficit in focused ultrasound-induced intracerebral hemorrhage. Nanomedicine (Lond). 2012
- Ghosh D, Choudhury ST, Ghosh S, Mandal AK, Sarkar S, Ghosh A, et al. Nanocapsulated curcumin: oral chemopreventive formulation against diethylnitrosamine induced hepatocellular carcinoma in rat. Chem Biol Interact. 2012; 195:206–214. [PubMed: 22197969]
- Miroliaee AE, Esmaily H, Vaziri-Bami A, Baeeri M, Shahverdi AR, Abdollahi M. Amelioration of experimental colitis by a novel nanoselenium-silymarin mixture. Toxicol Mech Methods. 2011; 21:200–208. [PubMed: 21247366]
- Morganti P, Fabrizi G, Palombo P, Palombo M, Guarneri F, Cardillo A, et al. New chitin complexes and their anti-aging activity from inside out. J Nutr Health Aging. 2012; 16:242–245. [PubMed: 22456780]
- Neves AR, Lucio M, Lima JL, Reis S. Resveratrol in medicinal chemistry: a critical review of its pharmacokinetics, drug-delivery, and membrane interactions. Curr Med Chem. 2012; 19:1663– 1681. [PubMed: 22257059]
- 43. Keijzer C, Slutter B, van der Zee R, Jiskoot W, van Eden W, Broere F. PLGA, PLGA-TMC and TMC-TPP nanoparticles differentially modulate the outcome of nasal vaccination by inducing tolerance or enhancing humoral immunity. PLoS ONE. 2011; 6:e26684. [PubMed: 22073184]

- 44. Zhang H, He X, Zhang Z, Zhang P, Li Y, Ma Y, et al. Nano-CeO2 exhibits adverse effects at environmental relevant concentrations. Environ Sci Technol. 2011; 45:3725–3730. [PubMed: 21428445]
- Rao KS, Ghorpade A, Labhasetwar V. Targeting anti-HIV drugs to the CNS. Expert Opin Drug Deliv. 2009; 6:771–784. [PubMed: 19566446]
- 46. McMillan J, Batrakova E, Gendelman HE. Cell delivery of therapeutic nanoparticles. Prog Mol Biol Transl Sci. 2011; 104:563–601. [PubMed: 22093229]
- Boyapalle S, Mohapatra S. Nanotechnology Applications to HIV Vaccines and Microbicides. J Glob Infect Dis. 2012; 4:62–68. [PubMed: 22529630]
- 48. Ranjan A, Pothayee N, Seleem MN, Boyle SM, Kasimanickam R, Riffle JS, et al. Nanomedicine for intracellular therapy. FEMS Microbiol Lett. 2012
- 49. Tiuman TS, Santos AO, Ueda-Nakamura T, Filho BP, Nakamura CV. Recent advances in leishmaniasis treatment. Int J Infect Dis. 2011; 15:e525–e532. [PubMed: 21605997]
- Nnamani PO, Scoles G, Krol S. Preliminary characterization of N-trimethylchitosan as a nanocarrier for malaria vaccin. J Vector Borne Dis. 2011; 48:224–230. [PubMed: 22297285]
- Santos-Magalhaes NS, Mosqueira VC. Nanotechnology applied to the treatment of malaria. Adv Drug Deliv Rev. 2010; 62:560–575. [PubMed: 19914313]
- 52. Gnanadesigan M, Anand M, Ravikumar S, Maruthupandy M, Vijayakumar V, Selvam S, et al. Biosynthesis of silver nanoparticles by using mangrove plant extract and their potential mosquito larvicidal property. Asian Pac J Trop Med. 2011; 4:799–803. [PubMed: 22014736]
- Debbage P. Targeted drugs and nanomedicine: present and future. Curr Pharm Des. 2009; 15:153– 172. [PubMed: 19149610]
- Montagnier L, Aissa J, Ferris S, Montagnier J-L, Lavallee C. Electromagnetic signals are produced by aqueous nanostructures derived from bacterial DNA sequences. Interdisciplinary Sci Comput Life Sci. 2009; 1:81–90.
- Dave SR, Gao X. Monodisperse magnetic nanoparticles for biodetection, imaging, and drug delivery: a versatile and evolving technology. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2009; 1:583–609. [PubMed: 20049819]
- 56. Sandhu A, Handa H, Abe M. Synthesis and applications of magnetic nanoparticles for biorecognition and point of care medical diagnostics. Nanotechnology. 2010; 21:442001. [PubMed: 20935358]
- Hossu M, Ma L, Chen W. Nonlinear enhancement of spontaneous biophoton emission of sweet potato by silver nanoparticles. J Photochem Photobiol B. 2010; 99:44–48. [PubMed: 20207158]
- 58. Ruan B, Jacobi M. Ultrasonication effects on thermal and rheological properties of carbon nanotube suspensions. Nanoscale Research Letters. 2012; 7:127. [PubMed: 22333487]
- Yao P, Hughes S. Macroscopic entanglement and violation of Bell's inequalities between two spatially separated quantum dots in a planar photonic crystal system. Opt Express. 2009; 17:11505–11514. [PubMed: 19582066]
- Bhakta G, Shrivastava A, Maitra A. Magnesium phosphate nanoparticles can be efficiently used in vitro and in vivo as non-viral vectors for targeted gene delivery. J Biomed Nanotechnol. 2009; 5:106–114. [PubMed: 20055113]
- Bisht S, Bhakta G, Mitra S, Maitra A. pDNA loaded calcium phosphate nanoparticles: highly efficient non-viral vector for gene delivery. Int J Pharm. 2005; 288:157–168. [PubMed: 15607268]
- Huang S, Chen JC, Hsu CW, Chang WH. Effects of nano calcium carbonate and nano calcium citrate on toxicity in ICR mice and on bone mineral density in an ovariectomized mice model. Nanotechnology. 2009; 20:375102. [PubMed: 19706952]
- Tavares Cardoso MA, Talebi M, Soares PA, Yurteri CU, van Ommen JR. Functionalization of lactose as a biological carrier for bovine serum albumin by electrospraying. Int J Pharmaceutics. 2011; 414:1–5.
- 64. Bhattacharyya SS, Paul S, Khuda-Bukhsh AR. Encapsulated plant extract (Gelsemium sempervirens) poly (lactide-co-glycolide) nanoparticles enhance cellular uptake and increase bioactivity in vitro. Exp Biol Med (Maywood). 2010; 235:678–688. [PubMed: 20511672]

- Bhakta G, Mitra S, Maitra A. DNA encapsulated magnesium and manganous phosphate nanoparticles: potential non-viral vectors for gene delivery. Biomaterials. 2005; 26:2157–2163. [PubMed: 15576191]
- Bhatti M, Yahioglu G, Milgrom LR, Garcia-Maya M, Chester KA, Deonarain MP. Targeted photodynamic therapy with multiply-loaded recombinant antibody fragments. Int J Cancer. 2008; 122:1155–1163. [PubMed: 17973256]
- 67. Deonarain MP, Milgrom LR. A Conjugation Platform for the Targeted Delivery of Anticancer Agents. Innovations in Pharmaceutical Technology. 2011:56–59.
- 68. Jain AK, Das M, Swarnakar NK, Jain S. Engineered PLGA nanoparticles: an emerging delivery tool in cancer therapeutics. Crit Rev Ther Drug Carrier Syst. 2011; 28:1–45. [PubMed: 21395514]
- 69. Hueber AJ, Stevenson R, Stokes RJ, Graham D, Garside P, McInnes IB. Imaging inflammation in real time--future of nanoparticles. Autoimmunity. 2009; 42:368–372. [PubMed: 19811304]
- Ahmad Z, Pandey R, Sharma S, Khuller GK. Alginate nanoparticles as antituberculosis drug carriers: formulation development, pharmacokinetics and therapeutic potential. Indian J Chest Dis Allied Sci. 2006; 48:171–176. [PubMed: 18610673]
- Bershteyn A, Hanson MC, Crespo MP, Moon JJ, Li AV, Suh H, et al. Robust IgG responses to nanograms of antigen using a biomimetic lipid-coated particle vaccine. J Control Release. 2012; 157:354–365. [PubMed: 21820024]
- Diwan M, Elamanchili P, Cao M, Samuel J. Dose sparing of CpG oligodeoxynucleotide vaccine adjuvants by nanoparticle delivery. Curr Drug Deliv. 2004; 1:405–412. [PubMed: 16305402]
- Bhattacharyya SS, Mandal SK, Biswas R, Paul S, Pathak S, Boujedaini N, et al. In vitro studies demonstrate anticancer activity of an alkaloid of the plant Gelsemium sempervirens. Exp Biol Med (Maywood). 2008; 233:1591–1601. [PubMed: 18997108]
- Prakash DJ, Arulkumar S, Sabesan M. Effect of nanohypericum (Hypericum perforatum gold nanoparticles) treatment on restraint stress induced behavioral and biochemical alteration in male albino mice. Pharmacognosy Res. 2010; 2:330–334. [PubMed: 21713134]
- Chifiriuc MC, Grumezescu V, Grumezescu AM, Saviuc CM, Lazar V, Andronescu E. Hybrid magnetite nanoparticles/Rosmarinus officinalis essential oil nanobiosystem with antibiofilm activity. Nanoscale Res Lett. 2012; 7:209. [PubMed: 22490675]
- He S, Zhou B, Zhang S, Lei Z, Zhang Z. Preparation of nanoparticles of Magnolia bark extract by rapid expansion from supercritical solution into aqueous solutions. J Microencapsul. 2011; 28:183–189. [PubMed: 21425944]
- 77. Han L, Fu Y, Cole AJ, Liu J, Wang J. Co-encapsulation and sustained-release of four components in ginkgo terpenes from injectable PELGE nanoparticles. Fitoterapia. 2012
- Felippi CC, Oliveira D, Stroher A, Carvalho AR, Van Etten EA, Bruschi M, et al. Safety and efficacy of antioxidants-loaded nanoparticles for an anti-aging application. J Biomed Nanotechnol. 2012; 8:316–321. [PubMed: 22515083]
- Lopes CM, Martins-Lopes P, Souto EB. Nanoparticulate carriers (NPC) for oral pharmaceutics and nutraceutics. Pharmazie. 2010; 65:75–82. [PubMed: 20225647]
- Meng X, Zu Y, Zhao X, Li Q, Jiang S, Sang M. Characterization and pharmacokinetics of coenzyme Q10 nanoparticles prepared by a rapid expansion of supercritical solution process. Pharmazie. 2012; 67:161–167. [PubMed: 22512087]
- Ghosh D, Ghosh S, Sarkar S, Ghosh A, Das N, Das Saha K, et al. Quercetin in vesicular delivery systems: evaluation in combating arsenic-induced acute liver toxicity associated gene expression in rat model. Chem Biol Interact. 2010; 186:61–71. [PubMed: 20371363]
- Anitha A, Maya S, Deepa N, Chennazhi KP, Nair SV, Jayakumar R. Curcumin-Loaded N,O-Carboxymethyl Chitosan Nanoparticles for Cancer Drug Delivery. J Biomater Sci Polym Ed. 2011
- Koufaki M, Detsi A, Kiziridi C. Multifunctional lipoic acid conjugates. Curr Med Chem. 2009; 16:4728–4742. [PubMed: 19903137]
- da Silva ER, Maquiaveli Cdo C, Magalhaes PP. The leishmanicidal flavonols quercetin and quercitrin target Leishmania (Leishmania) amazonensis arginase. Exp Parasitol. 2012; 130:183– 188. [PubMed: 22327179]
- Zandi K, Teoh BT, Sam SS, Wong PF, Mustafa MR, Abubakar S. Antiviral activity of four types of bioflavonoid against dengue virus type-2. Virol J. 2011; 8:560. [PubMed: 22201648]

- 86. Ganesan S, Faris AN, Comstock AT, Wang Q, Nanua S, Hershenson MB, et al. Quercetin inhibits rhinovirus replication in vitro and in vivo. Antiviral Res. 2012
- Choi HJ, Song JH, Kwon DH. Quercetin 3-rhamnoside exerts antiinfluenza A virus activity in mice. Phytother Res. 2012; 26:462–464. [PubMed: 21728202]
- Li H, Zhao X, Ma Y, Zhai G, Li L, Lou H. Enhancement of gastrointestinal absorption of quercetin by solid lipid nanoparticles. J Control Release. 2009; 133:238–244. [PubMed: 18951932]
- Cao X, Fu M, Wang L, Liu H, Deng W, Qu R, et al. Oral bioavailability of silymarin formulated as a novel 3-day delivery system based on porous silica nanoparticles. Acta Biomater. 2012; 8:2104– 2112. [PubMed: 22343518]
- Bisht S, Khan MA, Bekhit M, Bai H, Cornish T, Mizuma M, et al. A polymeric nanoparticle formulation of curcumin (NanoCurc) ameliorates CCl4-induced hepatic injury and fibrosis through reduction of pro-inflammatory cytokines and stellate cell activation. Lab Invest. 2011; 91:1383– 1395. [PubMed: 21691262]
- 91. Agalya Priyadarshini K, Murugan K, Panneerselvam C, Ponarulselvam S, Hwang JS, Nicoletti M. Biolarvicidal and pupicidal potential of silver nanoparticles synthesized using Euphorbia hirta against Anopheles stephensi Liston (Diptera: Culicidae). Parasitol Res. 2012
- 92. Sonkaew P, Sane A, Suppakul P. Antioxidant Activities of Curcumin and Ascorbyl Dipalmitate Nanoparticles and Their Activities after Incorporation into Cellulose-Based Packaging Films. J Agric Food Chem. 2012
- Antoniades C, Psarros C, Tousoulis D, Bakogiannis C, Shirodaria C, Stefanadis C. Nanoparticles: a promising therapeutic approach in atherosclerosis. Curr Drug Deliv. 2010; 7:303–311. [PubMed: 20695841]
- 94. Lee JE, Lee N, Kim T, Kim J, Hyeon T. Multifunctional mesoporous silica nanocomposite nanoparticles for theranostic applications. Acc Chem Res. 2011; 44:893–902. [PubMed: 21848274]
- Pattekari P, Zheng Z, Zhang X, Levchenko T, Torchilin V, Lvov Y. Top-down and bottom-up approaches in production of aqueous nanocolloids of low solubility drug paclitaxel. Phys Chem Chem Phys. 2011; 13:9014–9019. [PubMed: 21442095]
- 96. Singh SK, Srinivasan KK, Gowthamarajan K, Singare DS, Prakash D, Gaikwad NB. Investigation of preparation parameters of nanosuspension by top-down media milling to improve the dissolution of poorly water-soluble glyburide. Eur J Pharm Biopharm. 2011; 78:441–446. [PubMed: 21439378]
- Cai Z, Wang Y, Zhu LJ, Liu ZQ. Nanocarriers: a general strategy for enhancement of oral bioavailability of poorly absorbed or pre-systemically metabolized drugs. Curr Drug Metab. 2010; 11:197–207. [PubMed: 20384585]
- Wong HL, Chattopadhyay N, Wu XY, Bendayan R. Nanotechnology applications for improved delivery of antiretroviral drugs to the brain. Adv Drug Deliv Rev. 2010; 62:503–517. [PubMed: 19914319]
- Chadwick S, Kriegel C, Amiji M. Nanotechnology solutions for mucosal immunization. Adv Drug Deliv Rev. 2010; 62:394–407. [PubMed: 19931581]
- 100. Reis CP, Damge C. Nanotechnology as a promising strategy for alternative routes of insulin delivery. Methods Enzymol. 2012; 508:271–294. [PubMed: 22449931]
- 101. Fang IJ, Trewyn BG. Application of mesoporous silica nanoparticles in intracellular delivery of molecules and proteins. Methods Enzymol. 2012; 508:41–59. [PubMed: 22449920]
- 102. Uskokovic V, Uskokovic DP. Nanosized hydroxyapatite and other calcium phosphates: chemistry of formation and application as drug and gene delivery agents. J Biomed Mater Res B Appl Biomater. 2011; 96:152–191. [PubMed: 21061364]
- 103. Asano H, Muraki S, Endo H, Bandow S, Iijima S. Strong magnetism observed in carbon nanoparticles produced by the laser vaporization of a carbon pellet in hydrogen-containing Ar balance gas. J Phys Condens Matter. 2010; 22:334209. [PubMed: 21386499]
- 104. Ahmad Z, Khuller GK. Alginate-based sustained release drug delivery systems for tuberculosis. Expert Opin Drug Deliv. 2008; 5:1323–1334. [PubMed: 19040395]

- 105. Al-Jamal WT, Kostarelos K. Liposomes: from a clinically established drug delivery system to a nanoparticle platform for theranostic nanomedicine. Acc Chem Res. 2011; 44:1094–1104. [PubMed: 21812415]
- 106. Hadrup N, Loeschner K, Mortensen A, Sharma AK, Qvortrup K, Larsen EH, et al. The similar neurotoxic effects of nanoparticulate and ionic silver in vivo and in vitro. Neurotoxicology. 2012; 33:416–423. [PubMed: 22531227]
- 107. Liu Y, Gao Y, Zhang L, Wang T, Wang J, Jiao F, et al. Potential health impact on mice after nasal instillation of nano-sized copper particles and their translocation in mice. J Nanosci Nanotechnol. 2009; 9:6335–6343. [PubMed: 19908531]
- 108. Almeida JP, Chen AL, Foster A, Drezek R. In vivo biodistribution of nanoparticles. Nanomedicine (Lond). 2011; 6:815–835. [PubMed: 21793674]
- 109. Nowrouzi A, Meghrazi K, Golmohammadi T, Golestani A, Ahmadian S, Shafiezadeh M, et al. Cytotoxicity of subtoxic AgNP in human hepatoma cell line (HepG2) after long-term exposure. Iran Biomed J. 2010; 14:23–32. [PubMed: 20683495]
- 110. Park EJ, Park K. Oxidative stress and pro-inflammatory responses induced by silica nanoparticles in vivo and in vitro. Toxicol Lett. 2009; 184:18–25. [PubMed: 19022359]
- 111. Liu T, Li L, Teng X, Huang X, Liu H, Chen D, et al. Single and repeated dose toxicity of mesoporous hollow silica nanoparticles in intravenously exposed mice. Biomaterials. 2011; 32:1657–1668. [PubMed: 21093905]
- 112. Mohamed BM, Verma NK, Prina-Mello A, Williams Y, Davies AM, Bakos G, et al. Activation of stress-related signalling pathway in human cells upon SiO2 nanoparticles exposure as an early indicator of cytotoxicity. J Nanobiotechnology. 2011; 9:29. [PubMed: 21801388]
- 113. Seleem MN, Munusamy P, Ranjan A, Alqublan H, Pickrell G, Sriranganathan N. Silica-antibiotic hybrid nanoparticles for targeting intracellular pathogens. Antimicrob Agents Chemother. 2009; 53:4270–4274. [PubMed: 19667284]
- 114. Raffa V, Vittorio O, Riggio C, Cuschieri A. Progress in nanotechnology for healthcare. Minim Invasive Ther Allied Technol. 2010; 19:127–135. [PubMed: 20497066]
- 115. Ghosh S, Patil S, Ahire M, Kitture R, Gurav DD, Jabgunde AM, et al. Gnidia glauca flower extract mediated synthesis of gold nanoparticles and evaluation of its chemocatalytic potential. J Nanobiotechnology. 2012; 10:17. [PubMed: 22548753]
- 116. Daisy P, Saipriya K. Biochemical analysis of Cassia fistula aqueous extract and phytochemically synthesized gold nanoparticles as hypoglycemic treatment for diabetes mellitus. Int J Nanomedicine. 2012; 7:1189–1202. [PubMed: 22419867]
- 117. Das S, Das J, Samadder A, Bhattacharyya S, Das D, Khuda-Bukhsh AR. Biosynthesized silver nanoparticles by ethanolic extracts of Phytolacca decandra, Gelsemium sempervirens, Hydrastis canadensis and Thuja occidentalis induce differential cytotoxicity through G2/M arrest in A375 cells. Colloids and Surfaces B: Biointerfaces. 2013; 101:325–336.
- 118. Philip D. Green synthesis of gold and silver nanoparticles using Hibiscus rosa sinensis. Physica E: Low Dimens Sys Nanostruct. 2010; 42:1417–1424.
- 119. Nune SK, Chanda N, Shukla R, Katti K, Kulkarni RR, Thilakavathi S, et al. Green Nanotechnology from Tea: Phytochemicals in Tea as Building Blocks for Production of Biocompatible Gold Nanoparticles. J Mater Chem. 2009; 19:2912–2920. [PubMed: 20161162]
- 120. Bhattacharyya SS, Das J, Das S, Samadder A, Das D, De A, et al. Rapid green synthesis of silver nanoparticles from silver nitrate by a homeopathic mother tincture Phytolacca Decandra. Zhong Xi Yi Jie He Xue Bao. 2012; 10:546–554. [PubMed: 22587977]
- 121. Bornhoft, G.; Matthiessen, PF. Homeopathy in Healthcare -- Effectiveness, Appropriateness, Safety, Costs. Springer; 2011.
- 122. Witt, C.; Albrecht, H. New Directions in Homeopathy Research. Essen, Germany: KVC Verlag; 2009.
- 123. Witt CM, Luedtke R, Baur R, Willich SN. Homeopathic Medical Practice: Long-term results of a Cohort Study with 3981 Patients. BMC Public Health. 2005; 5:115. epub. [PubMed: 16266440]
- 124. Bronstein AC, Spyker DA, Cantilena LR, Green J, Rumack BH, Giffin SL. 2008 Annual Report of the American Association of Poison Control Centers' National Poison Data System (NPDS): 26th Annual Report. Clin Toxicol (Phila). 2009; 47:911–1084. [PubMed: 20028214]

- 125. Merisko-Liversidge E, Liversidge GG, Cooper ER. Nanosizing: a formulation approach for poorly-water-soluble compounds. Eur J Pharm Sci. 2003; 18:113–120. [PubMed: 12594003]
- 126. Hahnemann, S. Organon of the Medical Art. 6th ed.. Redmond, WA: Birdcage Books; 1843.
- 127. Kayne, SB. Homeopathic Practice. London: Pharmaceutical Press; 2008.
- 128. Kim TH, Kim M, Park HS, Shin US, Gong MS, Kim HW. Size-dependent cellular toxicity of silver nanoparticles. J Biomed Mater Res A. 2012
- 129. Barik TK, Sahu B, Swain V. Nanosilica-from medicine to pest control. Parasitol Res. 2008; 103:253–258. [PubMed: 18438740]
- 130. Wang T, Jiang H, Zhao Q, Wang S, Zou M, Cheng G. Enhanced mucosal and systemic immune responses obtained by porous silica nanoparticles used as an oral vaccine adjuvant: Effect of silica architecture on immunological properties. Int J Pharm. 2012; 436:351–358. [PubMed: 22721849]
- 131. Boericke, W. Pocket Manual of Homeopathic Materia Medica. Santa Rosa, CA: Boericke and Tafel, Inc.; 1927.
- 132. Rao ML, Roy R, Bell IR. The defining role of structure (including epitaxy) in the plausibility of homeopathy. Homeopathy. 2007; 96:175–182. [PubMed: 17678814]
- 133. Keck CM, Muller RH. Drug nanocrystals of poorly soluble drugs produced by high pressure homogenisation. Eur J Pharm Biopharm. 2006; 62:3–16. [PubMed: 16129588]
- 134. Ives JA, Moffett JR, Arun P, Lam D, Todorov TI, Brothers AB, et al. Enzyme stabilization by glass-derived silicates in glass-exposed aqueous solutions. Homeopathy. 2010; 99:15–24. [PubMed: 20129173]
- Demangeat JL. NMR relaxation evidence for solute-induced nanosized superstructures in ultramolecular aqueous dilutions of silica-lactose. Journal of Molecular Liquids. 2010; 155:71– 79.
- 136. Milgrom LR, King KR, Lee J, Pinkus AS. On the investigation of homeopathic potencies using low resolution NMR T2 relaxation times: an experimental and critical survey of the work of Roland Conte et al. British Homoeopathic Journal. 2001; 90:5–13. [PubMed: 11212090]
- 137. Liu L, Randolph TW, Carpenter JF. Particles shed from syringe filters and their effects on agitation-induced protein aggregation. J Pharm Sci. 2012; 101:2952–2959. [PubMed: 22674153]
- 138. Perry CC, Keeling-Tucker T. Crystalline silica prepared at room temperature from aqueous solution in the presence of intrasilica bioextracts. Chem Commun (Camb). 1998:2587–2588.
- Anick DJ, Ives JA. The silica hypothesis for homeopathy: physical chemistry. Homeopathy. 2007; 96:189–195. [PubMed: 17678816]
- 140. Liu Y, Kathan K, Saad W, Prudhomme RK. Ostwald ripening of B-carotene nanoparticles. Physical Review Letters. 2007; 98:1–4.
- 141. Tang C, Zhou T, Yang J, Zhang Q, Chen F, Fu Q, et al. Wet-grinding assisted ultrasonic dispersion of pristine multi-walled carbon nanotubes (MWCNTs) in chitosan solution. Colloids Surf B Biointerfaces. 2011; 86:189–197. [PubMed: 21530188]
- 142. Abbasi AR, Morsali A. Influence of solvents on the morphological properties of AgBr nanostructures prepared using ultrasound irradiation. Ultrason Sonochem. 2012; 19:540–545. [PubMed: 21963874]
- 143. Perry CC. An overview of silica in biology: its chemistry and recent technological advances. Progress in molecular and subcellular biology. 2009; 47:295–313. [PubMed: 19198783]
- 144. Song L, Yang K, Jiang W, Du P, Xing B. Adsorption of bovine serum albumin on nano and bulk oxide particles in deionized water. Colloids Surf B Biointerfaces. 2012; 94:341–346. [PubMed: 22405471]
- 145. Relaix S, Leheny RL, Reven L, Sutton M. Memory effect in composites of liquid crystal and silica aerosil. Phys Rev E Stat Nonlin Soft Matter Phys. 2011; 84 061705.
- 146. Baca HK, Carnes EC, Ashley CE, Lopez DM, Douthit C, Karlin S, et al. Cell-directed-assembly: directing the formation of nano/bio interfaces and architectures with living cells. Biochim Biophys Acta. 2011; 1810:259–267. [PubMed: 20933574]
- 147. Kaehr B, Townson JL, Kalinich RM, Awad YH, Swartzentruber BS, Dunphy DR, et al. Cellular complexity captured in durable silica biocomposites. Proc Natl Acad Sci U S A. 2012

- 148. Hornung V, Bauernfeind F, Halle A, Samstad EO, Kono H, Rock KL, et al. Silica crystals and aluminum salts activate the NALP3 inflammasome through phagosomal destabilization. Nat Immunol. 2008; 9:847–856. [PubMed: 18604214]
- 149. Zhu M, Li Y, Shi J, Feng W, Nie G, Zhao Y. Exosomes as extrapulmonary signaling conveyors for nanoparticle-induced systemic immune activation. Small. 2012; 8:404–412. [PubMed: 22144073]
- 150. Zhu M, Tian X, Song X, Li Y, Tian Y, Zhao Y, et al. Nanoparticle-Induced Exosomes Target Antigen-Presenting Cells to Initiate Th1-Type Immune Activation. Small. 2012
- Ive EC, Couchman IM, Reddy L. Therapeutic effect of Arsenicum album on leukocytes. Int J Mol Sci. 2012; 13:3979–3987. [PubMed: 22489193]
- 152. Singare DS, Marella S, Gowthamrajan K, Kulkarni GT, Vooturi R, Rao PS. Optimization of formulation and process variable of nanosuspension: An industrial perspective. Int J Pharm. 2010; 402:213–220. [PubMed: 20933066]
- 153. Elia V, Napoli E, Germano R. The 'Memory of Water': an almost deciphered enigma. Dissipative structures in extremely dilute aqueous solutions. Homeopathy. 2007; 96:163–169. [PubMed: 17678812]
- 154. Iavicoli I, Calabrese EJ, Nascarella MA. Exposure to nanoparticles and hormesis. Dose Response. 2010; 8:501–517. [PubMed: 21191487]
- 155. Karpukhin AV, Avkhacheva NV, Yakovlev RY, Kulakova II, Yashin VA, Lisichkin GV, et al. Effect of detonation nanodiamonds on phagocyte activity. Cell Biol Int. 2011; 35:727–733. [PubMed: 21155712]
- 156. Calabrese EJ. Converging concepts: adaptive response, preconditioning, and the Yerkes-Dodson Law are manifestations of hormesis. Ageing Res Rev. 2008; 7:8–20. [PubMed: 17768095]
- 157. Calabrese EJ. Hormesis is central to toxicology, pharmacology and risk assessment. Hum Exp Toxicol. 2010; 29:249–261. [PubMed: 20332169]
- 158. Calabrese EJ, Bachmann KA, Bailer AJ, Bolger PM, Borak J, Cai L, et al. Biological stress response terminology: Integrating the concepts of adaptive response and preconditioning stress within a hormetic dose-response framework. Toxicol Appl Pharmacol. 2007; 222:122–128. [PubMed: 17459441]
- 159. Mattson MP. Hormesis defined. Aging Research Rev. 2008; 7:1-7.
- 160. Van Wijk R, Wiegant FA. Postconditioning hormesis and the homeopathic Similia principle: molecular aspects. Hum Exp Toxicol. 2010; 29:561–565. [PubMed: 20558607]
- 161. Van Wijk R, Wiegant FA. Postconditioning hormesis and the similia principle. Front Biosci (Elite Ed). 2011; 3:1128–1138. [PubMed: 21622119]
- 162. Wiegant FA, Prins HA, Van Wijk R. Postconditioning hormesis put in perspective: an overview of experimental and clinical studies. Dose Response. 2011; 9:209–224. [PubMed: 21731537]
- 163. Calabrese V, Cornelius C, Cuzzocrea S, Iavicoli I, Rizzarelli E, Calabrese EJ. Hormesis, cellular stress response and vitagenes as critical determinants in aging and longevity. Mol Aspects Med. 2011; 32:279–304. [PubMed: 22020114]
- 164. Calabrese V, Cornelius C, Dinkova-Kostova AT, Iavicoli I, Di Paola R, Koverech A, et al. Cellular stress responses, hormetic phytochemicals and vitagenes in aging and longevity. Biochim Biophys Acta. 2012; 1822:753–783. [PubMed: 22108204]
- 165. Calabrese EJ. Paradigm lost, paradigm found: the re-emergence of hormesis as a fundamental dose response model in the toxicological sciences. Environ Pollut. 2005; 138:379–411. [PubMed: 16098930]
- 166. Nascarella MA, Calabrese EJ. A method to evaluate hormesis in nanoparticle dose-responses. Dose Response. 2012; 10:344–354. [PubMed: 22942868]
- 167. Demirovic D, Rattan SI. Establishing cellular stress response profiles as biomarkers of homeodynamics, health and hormesis. Exp Gerontol. 2012
- 168. Mihalik A, Csermely P. Heat shock partially dissociates the overlapping modules of the yeast protein-protein interaction network: a systems level model of adaptation. PLoS Comput Biol. 2011; 7 e1002187.

- 169. Szalay MS, Kovacs IA, Korcsmaros T, Bode C, Csermely P. Stress-induced rearrangements of cellular networks: Consequences for protection and drug design. FEBS Lett. 2007; 581:3675– 3680. [PubMed: 17433306]
- 170. Vidal M, Cusick ME, Barabasi AL. Interactome networks and human disease. Cell. 2011; 144:986–998. [PubMed: 21414488]
- 171. Ives JA, Jonas WB, Frye JC. Do serial dilutions really dilute? Homeopathy. 2010; 99:229–230. [PubMed: 20970091]
- 172. Davidson J. Psychiatry and homeopathy. Basis for a dialogue. British Homoeopathic Journal. 1994; 83:78–83.
- 173. Bell IR, Howerter A, Jackson N, Brooks AJ, Schwartz GE. Multi-week resting EEG cordance change patterns from repeated olfactory activation with two constitutionally-salient homeopathic remedies in healthy young adults. J Alternative and Complementary Medicine. 2012; 18:445– 453.
- 174. Antelman SM, Levine J, Gershon S. Time-dependent sensitization: the odyssey of a scientific heresy from the laboratory to the door of the clinic. Molecular Psychiatry. 2000; 5:350–356. [PubMed: 10889544]
- 175. Calabrese EJ, Jonas WB. Homeopathy: clarifying its relationship to hormesis. Hum Exp Toxicol. 2010 Jul; 29(7):531–536. 2010;29:531-6. [PubMed: 20558601]
- 176. Calabrese EJ, Mattson MP, Calabrese V. Resveratrol commonly displays hormesis: occurrence and biomedical significance. Hum Exp Toxicol. 2010; 29:980–1015. [PubMed: 21115559]
- 177. Fisher P. What is homeopathy? An introduction. Front Biosci (Elite Ed). 2012; 4:1669–1682. [PubMed: 22201984]
- 178. Lira-Salazar G, Marines-Montiel E, Torres-Monzon J, Hernandez-Hernandez F, Salas-Benito JS. Effects of homeopathic medications Eupatorium perfoliatum and Arsenicum album on parasitemia of Plasmodium berghei-infected mice. Homeopathy. 2006; 95:223–228. [PubMed: 17015193]
- 179. Pereira WK, Lonardoni MV, Grespan R, Caparroz-Assef SM, Cuman RK, Bersani-Amado CA. Immunomodulatory effect of Canova medication on experimental Leishmania amazonensis infection. J Infect. 2005; 51:157–164. [PubMed: 16038768]
- 180. Basu MK, Lala S. Macrophage specific drug delivery in experimental leishmaniasis. Curr Mol Med. 2004; 4:681–689. [PubMed: 15357216]
- 181. Danesh-Bahreini MA, Shokri J, Samiei A, Kamali-Sarvestani E, Barzegar-Jalali M, Mohammadi-Samani S. Nanovaccine for leishmaniasis: preparation of chitosan nanoparticles containing Leishmania superoxide dismutase and evaluation of its immunogenicity in BALB/c mice. Int J Nanomedicine. 2011; 6:835–842. [PubMed: 21589651]
- 182. Tafaghodi M, Khamesipour A, Jaafari MR. Immunization against leishmaniasis by PLGA nanospheres encapsulated with autoclaved Leishmania major (ALM) and CpG-ODN. Parasitol Res. 2011; 108:1265–1273. [PubMed: 21125294]
- 183. Glatthaar-Saalmuller B. In vitro evaluation of the antiviral effects of the homeopathic preparation Gripp-Heel on selected respiratory viruses. Can J Physiol Pharmacol. 2007; 85:1084–1090. [PubMed: 18066110]
- 184. Ullman D. Controlled clinical trials evaluating the homeopathic treatment of people with human immunodeficiency virus or acquired immune deficiency syndrome. J Altern Complement Med. 2003; 9:133–141. [PubMed: 12676041]
- 185. Sherr, J. Dynamic Provings Volume 1. Malvern, Worcester UK: Dynamis Books; 1997.
- 186. Lam R, Chen M, Pierstorff E, Huang H, Osawa E, Ho D. Nanodiamond-embedded microfilm devices for localized chemotherapeutic elution. ACS Nano. 2008; 2:2095–2102. [PubMed: 19206456]
- 187. Mochalin VN, Shenderova O, Ho D, Gogotsi Y. The properties and applications of nanodiamonds. Nat Nanotechnol. 2012; 7:11–23. [PubMed: 22179567]
- 188. Bracho G, Varela E, Fernandez R, Ordaz B, Marzoa N, Menendez J, et al. Large-scale application of highly-diluted bacteria for Leptospirosis epidemic control. Homeopathy. 2010; 99:156–166. [PubMed: 20674839]

- 189. Ulery BD, Kumar D, Ramer-Tait AE, Metzger DW, Wannemuehler MJ, Narasimhan B. Design of a protective single-dose intranasal nanoparticle-based vaccine platform for respiratory infectious diseases. PLoS ONE. 2011; 6:e17642. [PubMed: 21408610]
- 190. Tang M, Zhang T, Xue Y, Wang S, Huang M, Yang Y, et al. Dose dependent in vivo metabolic characteristics of titanium dioxide nanoparticles. J Nanosci Nanotechnol. 2010; 10:8575–8583. [PubMed: 21121368]
- 191. Ferley JP, Zmirou D, D'Adhemar D, Balducci F. A controlled evaluation of a homoeopathic preparation in the treatment of influenza-like syndromes. British Journal of Clinical Pharmacology. 1989; 27:329–335. [PubMed: 2655683]
- 192. Papp R, et al. Oscillococcinum in patients with influenza-like syndromes: a placebo-controlled double blind evaluation. British Homeopathy Journal. 1998; 87:69–76.
- 193. Vickers AJ, Smith C. Homoeopathic Oscillococcinum for preventing and treating influenza and influenza-like syndromes. Cochrane Database Syst Rev. 2006; 19 CD001957.
- 194. Haidvogl M, Riley DS, Heger M, Brien S, Jong M, Fischer M, Lewith GT, Jansen G, Thurneysen AE. Homeopathic and conventional treatment for acute respiratory and ear complaints: a comparative study on outcome in the primary care setting. BMC Complement Altern Med. 2007; 2:7. [PubMed: 17335565]
- 195. Trichard M, Chaufferin G, Nicoloyannis N. Pharmacoeconomic comparison between homeopathic and antibiotic treatment strategies in recurrent acute rhinopharyngitis in children. Homeopathy Journal of the Faculty of Homeopathy. 2005; 94:3–9.
- 196. Friese KH, Kruse S, Ludtke R, Moeller H. The homoeopathic treatment of otitis media in children--comparisons with conventional therapy. Int J Clin Pharmacol Ther. 1997; 35:296–301. [PubMed: 9247843]
- 197. Sinha MN, Siddiqui VA, Nayak C, Singh V, Dixit R, Dewan D, et al. Randomized controlled pilot study to compare homeopathy and conventional therapy in acute otitis media. Homeopathy. 2012; 101:5–12. [PubMed: 22226309]
- 198. Bellavite P, Marzotto M, Chirumbolo S, Conforti A. Advances in homeopathy and immunology: a review of clinical research. Front Biosci (Schol Ed). 2011; 3:1363–1389. [PubMed: 21622275]
- 199. Ramchandani NM. Homoeopathic treatment of upper respiratory tract infections in children: evaluation of thirty case series. Complement Ther Clin Pract. 2010; 16:101–108. [PubMed: 20347842]
- 200. Schmiedel V, Klein P. A complex homeopathic preparation for the symptomatic treatment of upper respiratory infections associated with the common cold: An observational study. Explore (NY). 2006; 2:109–114. [PubMed: 16781624]
- 201. Ramachandran C, Nair PK, Clement RT, Melnick SJ. Investigation of cytokine expression in human leukocyte cultures with two immune-modulatory homeopathic preparations. J Altern Complement Med. 2007; 13:403–407. [PubMed: 17532732]
- 202. Roeska K, Seilheimer B. Antiviral activity of Engystol(R) and Gripp-Heel(R): an in-vitro assessment. J Immune Based Ther Vaccines. 2010; 8:6. [PubMed: 21080959]
- 203. Aickin M. The end of biomedical journals: there is madness in their methods. Journal of Alternative & Complementary Medicine. 2005; 11:755–757.
- 204. Fisher P, Berman B, Davidson J, Reilly D, Thompson T. Are the clinical effects of homoeopathy placebo effects? Lancet. 2005; 366:2082–2083. [PubMed: 16360780]
- 205. Ludtke R, Rutten AL. The conclusions on the effectiveness of homeopathy highly depend on the set of analyzed trials. J Clin Epidemiol. 2008; 61:1197–1204. [PubMed: 18834714]
- 206. Shang A, Huwiler-Muntener K, Nartey L, Juni P, Dorig S, Sterne JA, Pewsner D, Egger M. Are the clinical effects of homoeopathy placebo effects? Comparative study of placebo-controlled trials of homoeopathy and allopathy. Lancet. 2005; 366:726–732. [PubMed: 16125589]
- 207. Adair BM. Nanoparticle vaccines against respiratory viruses. Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2009; 1:405–414. [PubMed: 20049806]
- 208. Papp I, Sieben C, Ludwig K, Roskamp M, Bottcher C, Schlecht S, et al. Inhibition of influenza virus infection by multivalent sialic-acid-functionalized gold nanoparticles. Small. 2010; 6:2900– 2906. [PubMed: 21104827]

- 209. Tai W, Roberts L, Seryshev A, Gubatan JM, Bland CS, Zabriskie R, et al. Multistrain influenza protection induced by a nanoparticulate mucosal immunotherapeutic. Mucosal Immunol. 2011; 4:197–207. [PubMed: 20736998]
- 210. Calabrese EJ, Mattson MP. Hormesis provides a generalized quantitative estimate of biological plasticity. J Cell Commun Signal. 2011; 5:25–38. [PubMed: 21484586]
- 211. Vithoulkas, G. The Science of Homeopathy. N.Y.: Grove Weidenfeld; 1980.
- 212. Antelman SM, Caggiula AR, Kocan D, Knopf S, Meyer D, Edwards DJ, et al. One experience with 'lower' or 'higher' intensity stressors, respectively enhances or diminishes responsiveness to haloperidol weeks later: implications for understanding drug variability. Brain Research. 1991; 566:276–283. [PubMed: 1814544]
- 213. Sorg B, Bailie T, Tschirgi M, Li N, Wu W. Exposure to repeated low-level formaldehyde in rats increases basal corticosterone levels and enhances the corticosterone response to subsequent formaldehyde. Brain Res. 2001; 898:314–320. [PubMed: 11306018]
- Csermely P. Chaperone overload is a possible contributor to 'civilization diseases'. Trends Genet. 2001; 17:701–704. [PubMed: 11718923]
- 215. Csermely P, Korcsmaros T, Kovacs IA, Szalay MS, Soti C. Systems biology of molecular chaperone networks. Novartis Found Symp. 2008; 291:45–54. discussion -8, 137-40. [PubMed: 18575265]
- 216. Karatsoreos IN, McEwen BS. Psychobiological allostasis: resistance, resilience and vulnerability. Trends Cogn Sci. 2011; 15:576–584. [PubMed: 22078931]
- 217. McEwen BS. Protective and damaging effects of stress mediators: central role of the brain. Dialogues Clin Neurosci. 2006; 8:367–381. [PubMed: 17290796]
- 218. Li C, Li X, Suzuki AK, Fujitani Y, Jigami J, Nagaoka K, et al. Effects of exposure to nanoparticle-rich diesel exhaust on adrenocortical function in adult male mice. Toxicol Lett. 2012; 209:277–281. [PubMed: 22260943]
- 219. Rattan SI, Deva T. Testing the hormetic nature of homeopathic interventions through stress response pathways. Hum Exp Toxicol. 2010; 29:551–554. [PubMed: 20558605]
- 220. Wiegant FA, Spieker N, van Wijk R. Stressor-specific enhancement of hsp induction by low doses of stressors in conditions of self- and cross-sensitization. Toxicology. 1998; 127:107–119. [PubMed: 9699798]
- 221. Coelho Moreira CO, de Fatima Ferreira Borges da Costa J, Leal MF, Ferreira de Andrade E, Rezende AP, Imbeloni AA, et al. Lymphocyte proliferation stimulated by activated Cebus apella macrophages treated with a complex homeopathic immune response modifiers. Homeopathy. 2012; 101:74–79. [PubMed: 22226318]
- 222. Naser B, Bodinet C, Tegtmeier M, Lindequist U. Thuja occidentalis (Arbor vitae): A Review of its Pharmaceutical, Pharmacological and Clinical Properties. Evid Based Complement Alternat Med. 2005; 2:69–78. [PubMed: 15841280]
- 223. Bellavite P, Ortolani R, Pontarollo F, Pitari G, Conforti A. Immunology and Homeopathy. 5. The Rationale of the 'Simile'. Evid Based Complement Alternat Med. 2007; 4:149–163. [PubMed: 17549232]
- 224. Bell IR, Caspi O, Schwartz GE, Grant KL, Gaudet TW, Rychener D, et al. Integrative medicine and systemic outcomes research: issues in the emergence of a new model for primary health care. Archives of Internal Medicine. 2002; 162:133–140. [PubMed: 11802746]
- 225. Koithan M, Bell IR, Niemeyer K, Pincus D. A complex systems science perspective for whole systems of CAM research. Forschende Komplementarmedizin und Klassische Naturheilkunde. 2012; 19:7–14.
- 226. Barabasi AL, Gulbahce N, Loscalzo J. Network medicine: a network-based approach to human disease. Nat Rev Genet. 2011; 12:56–68. [PubMed: 21164525]
- 227. Brumovsky PR, Gebhart GF. Visceral organ cross-sensitization an integrated perspective. Auton Neurosci. 2010; 153:106–115. [PubMed: 19679518]
- 228. Calabrese V, Cornelius C, Trovato A, Cavallaro M, Mancuso C, Di Rienzo L, et al. The hormetic role of dietary antioxidants in free radical-related diseases. Curr Pharm Des. 2010; 16:877–883. [PubMed: 20388101]

- 229. Vaiserman AM. Hormesis and epigenetics: is there a link? Ageing Res Rev. 2011; 10:413–421. [PubMed: 21292042]
- 230. Danese A, McEwen BS. Adverse childhood experiences, allostasis, allostatic load, and agerelated disease. Physiol Behav. 2012; 106:29–39. [PubMed: 21888923]
- 231. Pincus D, Metten A. Nonlinear dynamics in biopsychosocial resilience. Nonlinear Dynamics Psychol Life Sci. 2010; 14:353–380. [PubMed: 20887686]
- 232. McEwen BS. Central effects of stress hormones in health and disease: Understanding the protective and damaging effects of stress and stress mediators. Eur J Pharmacol. 2008; 583:174–185. [PubMed: 18282566]
- 233. Gross T, Blasius B. Adaptive coevolutionary networks: a review. J R Soc Interface. 2008; 5:259– 271. [PubMed: 17971320]
- 234. Vasquez A, Dobrin R, Sergi D, Eckmann JP, Oltvai ZN, Barabasi AL. The topological relationship between the large-scale attributes and local interaction patterns of complex networks. Proceedings of the National Academy of Sciences of the United States of America. 2004; 101:17940–17945. [PubMed: 15598746]
- 235. Calabrese V, Cornelius C, Mancuso C, Pennisi G, Calafato S, Bellia F, et al. Cellular stress response: a novel target for chemoprevention and nutritional neuroprotection in aging, neurodegenerative disorders and longevity. Neurochem Res. 2008; 33:2444–2471. [PubMed: 18629638]
- 236. Mattson MP, Son TG, Camandola S. Viewpoint: mechanisms of action and therapeutic potential of neurohormetic phytochemicals. Dose Response. 2007; 5:174–186. [PubMed: 18648607]
- 237. Veeraapandian S, Sawant SN, Doble M. Antibacterial and antioxidant activity of protein capped silver and gold nanoparticles synthesized with Escherichia coli. J Biomed Nanotechnol. 2012; 8:140–148. [PubMed: 22515102]
- 238. Kaul G, Thippeswamy H. Role of Heat Shock Proteins in Diseases and Their Therapeutic Potential. Indian J Microbiol. 2011; 51:124–131. [PubMed: 22654152]
- Petrilli V, Dostert C, Muruve DA, Tschopp J. The inflammasome: a danger sensing complex triggering innate immunity. Curr Opin Immunol. 2007; 19:615–622. [PubMed: 17977705]
- 240. Winter M, Beer HD, Hornung V, Kramer U, Schins RP, Forster I. Activation of the inflammasome by amorphous silica and TiO2 nanoparticles in murine dendritic cells. Nanotoxicology. 2011; 5:326–340. [PubMed: 20846021]
- 241. Demento SL, Eisenbarth SC, Foellmer HG, Platt C, Caplan MJ, Mark Saltzman W, et al. Inflammasome-activating nanoparticles as modular systems for optimizing vaccine efficacy. Vaccine. 2009; 27:3013–3021. [PubMed: 19428913]
- 242. Muruve DA, Petrilli V, Zaiss AK, White LR, Clark SA, Ross PJ, et al. The inflammasome recognizes cytosolic microbial and host DNA and triggers an innate immune response. Nature. 2008; 452:103–107. [PubMed: 18288107]
- 243. Petrilli V, Martinon F. The inflammasome, autoinflammatory diseases, and gout. Joint Bone Spine. 2007; 74:571–576. [PubMed: 17714972]
- 244. Elenkov IJ, Iezzoni DG, Daly A, Harris AG, Chrousos GP. Cytokine dysregulation, inflammation and well-being. Neuroimmunomodulation. 2005; 12:255–269. [PubMed: 16166805]
- 245. Stern S, Dror T, Stolovicki E, Brenner N, Braun E. Genome-wide transcriptional plasticity underlies cellular adaptation to novel challenge. Mol Syst Biol. 2007; 3:106. [PubMed: 17453047]
- 246. Hale HB. Cross-adaptation. Environmental Research. 1969; 2:423–434. [PubMed: 4909667]
- 247. Launay JC, Besnard Y, Guinet-Lebreton A, Savourey G. Acclimation to intermittent hypobaric hypoxia modifies responses to cold at sea level. Aviat Space Environ Med. 2006; 77:1230–1235. [PubMed: 17183918]
- 248. Avena NM, Rada P, Hoebel BG. Evidence for sugar addiction: Behavioral and neurochemical effects of intermittent, excessive sugar intake. Neurosci Biobehav Rev. 2008; 32:20–39. [PubMed: 17617461]
- 249. Antelman SM, Caggiula AR. Oscillation follows drug sensitization: implications. Critical Reviews in Neurobiology. 1996; 10:101–117. [PubMed: 8853956]

- 250. Sorg BA, Swindell S, Tschirgi ML. Repeated low level formaldehyde exposure produces enhanced fear conditioning to odor in male, but not female, rats. Brain Research. 2004; 1:11–19. [PubMed: 15081377]
- 251. Sorg B, Tschirgi M, Swindell S, Chen L, Fang J. Repeated formaldehyde effects in an animal model for multiple chemical sensitivity. Ann N Y Acad Sci. 2001; 933:57–67. [PubMed: 12000036]
- 252. Bharali DJ, Mousa SA, Thanavala Y. Micro- and nanoparticle-based vaccines for hepatitis B. Adv Exp Med Biol. 2007; 601:415–421. [PubMed: 17713030]
- 253. Camacho AI, de Souza J, Sanchez-Gomez S, Pardo-Ros M, Irache JM, Gamazo C. Mucosal immunization with Shigella flexneri outer membrane vesicles induced protection in mice. Vaccine. 2011; 29:8222–8229. [PubMed: 21911022]
- 254. Duceppe N, Tabrizian M. Advances in using chitosan-based nanoparticles for in vitro and in vivo drug and gene delivery. Expert Opin Drug Deliv. 2010; 7:1191–1207. [PubMed: 20836623]
- 255. Mansour HM, Rhee YS, Wu X. Nanomedicine in pulmonary delivery. Int J Nanomedicine. 2009; 4:299–319. [PubMed: 20054434]
- 256. Laroui H, Sitaraman SV, Merlin D. Gastrointestinal Delivery of Anti-inflammatory Nanoparticles. Methods Enzymol. 2012; 509:101–125. [PubMed: 22568903]
- 257. Bansal SS, Goel M, Aqil F, Vadhanam MV, Gupta RC. Advanced drug delivery systems of curcumin for cancer chemoprevention. Cancer Prev Res (Phila). 2011; 4:1158–1171. [PubMed: 21546540]
- 258. Beg S, Javed S, Kohli K. Bioavailability enhancement of coenzyme Q10: an extensive review of patents. Recent Pat Drug Deliv Formul. 2010; 4:245–255. [PubMed: 20863275]

Table 1

Exemplar Studies of Herbal-Facilitated Nanoparticle Product Production

Herb or Botanical	Nanoparticle Type	References
Hypericum	Gold	[74]
Gelsemium sempervirens	Poly(lactide-co-glycolide)	[64]
Gelsemium sempervirens	Silica (glass), Polypropylene	[73]
Rosmarinus officinalis	Magnetite (iron oxide/oleic acid)	[75]
Magnolia bark	Magnolia bark extract	[76]
Gingko biloba	monomethyl poly(ethylene glycol)–poly (lactide-co-glycolide)–monomethyl-poly(ethylene-glycol), MeO–PEG–PLGA–PEG–OMe polymers	[77]
Cassia fistula	Gold	[116]
Thuja occidentalis Phytolacca decandra Hydrastis Canadensis Gelsemium sempervirens	Silver	[117]
Equisetum telmateia	Silica	[138]