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Herbs as an antioxidant arsenal for periodontal diseases

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ABSTRACT

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Received: November 03, 2015 Accepted: January 05, 2016 Published: January 27, 2016 Herbal medicines have long been used as a traditional mode of therapy for various ailments in India. They are being used increasingly as dietary supplements to ward off common diseases. Periodontal diseases are highly prevalent and can affect up to 90% of the world population. Gingivitis is the mild form whereas periodontitis results in an irreversible loss of supporting structures of the teeth. Even though periodontal pathogens form a crucial component in the etiopathogenesis of periodontitis, there is a growing body of evidence suggesting oxidative stress playing a pivotal role in the disease initiation and progression. Studies have shown a direct correlation between increased levels of biomarkers for tissue damage induced by reactive oxygen species (ROS) to the severity of periodontal disease. Thus, the focus of attention has revolved back to herbal medicines due to their wide spectrum of biological and medicinal activities, lower costs, and higher safety margin. Internet databases Pubmed and Google Scholar were searched, and the most relevant articles were considered for review. This review briefly describes the various herbs with antioxidant capacity and their potency in the treating periodontal disease.

KEY WORDS: Antioxidant capacity, herbal medicine, periodontitis

INTRODUCTION

Herbal medicines, including herbs, herbal preparations and finished herbal products, contain as active ingredients parts of plants or other plant materials perceived to have therapeutic benefits [1]. About 80% of the worldwide population use herbal products for their basic health care (primary care) such as extracts, teas and other active principles, a market estimated at US\$ 50 billion per year [2]. Herbal products are preferred over conventional drugs due to wide biological activity, higher safety margin, and lower costs. Furthermore, the conventional drugs are known to cause various side effects, and continuous intake has resulted in antibiotic resistance. Thus, herbal medicines are being used increasingly as dietary supplements to fight or prevent common diseases [3].

Periodontitis is a chronic inflammatory disease which results in the destruction of supporting structures of the teeth. The etiology is multifactorial with periodontopathogens forming a major crux in the initiation and progression of the disease. Plaque build-up allows the growth of anaerobic bacteria [4], which eventually leads to the recruitment and activation of neutrophils. This further results in the upregulation of pro-inflammatory cytokines and also leads to the release of neutrophilic enzymes and ROS. Prolonged exposure of the connective tissue to these insults results in the degradation and subsequent loss of ligamentous support and alveolar bone, eventually leading to tooth loss [5].

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Periodontal therapy includes both surgical and nonsurgical management of the disease process. Various antimicrobials and chemotherapeutic agents, such as chlorhexidine, triclosan, cetylpyridinium chloride, have been tried and tested in the management of periodontal diseases. Due to its multifactorial etiology and complex disease process, the treatment of periodontitis is still a formidable task to dentists. Therefore, herbal remedies have been sought to achieve antimicrobial, antioxidant, antiseptic, anti-inflammatory, and anti-collagenase effects. The following review briefly describes the role of oxidative stress in periodontitis and the various herbs with antioxidant potential used in its management.

OXIDATIVE STRESS IN PERIODONTITIS

It is well-established that oxidative stress is an important cause of cell damage associated with the initiation and progression of many chronic diseases [6-8]. A recent review by Bullon describes the mounting evidence that the basis for the interrelationships between chronic periodontitis and atheromatous disease and diabetes lies at the fundamental intracellular level, namely oxidative stress and mitochondrial dysfunction [9]. Oxidative stress is the disturbance in the pro-oxidant and antioxidant balance, in favor of the former, resulting in potential tissue damage. Consecutive oxidationreduction reactions of molecular oxygen by various enzymes results in the production of molecules such as superoxide anion, hydrogen peroxide, hydroxyl radical, singlet oxygen, nitric oxide, hypochlorous acid which together constitute the term "ROS" [10]. All the cells in the body are capable of generating ROS, of which polymorphonuclear neutrophils are of prime importance with respect to periodontitis. Neutrophils are a part of innate immunity which comprise the first line of host defense and are located at sites of microbial invasion. They are activated by inflammatory mediators and can generate increased levels of ROS, which not only attack the periodontopathogens but also the surrounding tissues [11]. The ROS is generated by the nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (Nox) system present in the neutrophils and it catalyzes the reduction of molecular oxygen to superoxide anion. Subsequent reductions result in the production of hydrogen peroxide and hydroxyl radical. Similarly, superoxide dismutase enzyme present in all the cells catalyzes the dismutation of superoxide radical to hydrogen peroxide. Many studies have shown that ROS regulate the formation and function of osteoclasts, i.e., the activation and bone resorption ability [12-14]. Bone resorption which results in alveolar bone loss and ultimately tooth loss is the hallmark feature of the periodontal disease. NADPH oxidase system plays a role in periodontal pathologies and its involvement is the strongest in aggressive periodontitis [15].

To combat the oxidative stress, all the cells in the body are equipped with an intrinsic store of molecules known as "antioxidants." Antioxidants may be regarded as "those substances which when present at low concentrations, compared to those of an oxidizable substrate, will significantly delay, or inhibit oxidation of that substrate" [16]. They function by scavenging free radicals as and when they form and thereby preventing oxidative stress. They can also sequester transition metal ions and prevent Fenton's reaction or catalyze the oxidation of other molecules. Various antioxidant molecules include vitamins C, E, coenzyme Q, carotenoids, enzymes such as glutathione reductase, glutathione transferase, superoxide dismutase, and peroxiredoxin.

Numerous studies have shown that the total antioxidant capacity in periodontitis patients is significantly lower when compared to healthy controls [17-19] or in subjects who have received periodontal therapy [20]. These findings have triggered the use of exogenous supplements for the treatment of periodontal disease. Herbal antioxidant remedies have been the focus of research in recent times. A literature search was performed using keywords such as "plant extracts," "herbs," "herbal medicine," "antioxidants," "oxidative stress" "periodontal disease" in PubMed, and Google Scholar. The most relevant articles were included in this review and it gives an overview about the potential of herbal medicine in the management of periodontitis.

GREEN TEA

Components

Polyphenols contained in teas are classified as catechins. There are six primary catechin compounds in green tea: Catechin, gallocatechin, epicatechin, epicatechin, epicatechin, epicatechin gallate (ECg), and epigallocatechin gallate (EGCg). EGCg has been the focus of extensive research among all the other compounds and it is a very potent antioxidant. Green tea also contains carotenoids, tocopherols, ascorbic acid, minerals such as zinc, selenium, chromium, and certain phytochemical compounds.

Role in Periodontitis

Green tea catechins have been observed to have profound effects on periodontal pathogens. Anaerobic bacteria like *Porphyromonas gingivalis* and *Prevotella* spp. are the main etiological agents in periodontitis. *In vitro* studies have shown that these compounds inhibit the growth of *P. gingivalis*, *Prevotella intermedia* and *Prevotella nigrescens* [22-24]. It also prevents the adherence of *P. gingivalis* onto human buccal epithelial cells [25]. A study also showed that both ECg and EGCg inhibited *P. gingivalis* derived collagenase activity [26].

Bone resorption which occurs in periodontitis is due to the interplay between osteoblasts and osteoclasts. In an animal study, it was shown that EGCg reduced lipopolysaccharidemediated bone resorption in both *in-vivo* and *in-vitro* conditions. It also showed that EGCg suppressed LPS mediated gene expression such as RANKL, cyclooxygenase-1 and the cytokine PGE2 in mouse osteoblasts. This clearly suggests that the catechin present in green tea is highly potent in suppressing the bone resorption mediated by an inflammatory response as seen in periodontal disease [27].

There are various reports about the use of green tea in various forms in the management of periodontitis. Pilot studies on the usage of green tea as a dentifrice and a local drug delivery system have observed an improvement in the periodontal status of the patients suffering from chronic periodontitis [28,29]. A clinical trial indicated that green tea mouthwash had a comparable antiplaque efficacy to chlorhexidine gluconate (gold standard) when used for a period of 1 week [30]. These preliminary results show that further research is needed to explore and tap the benefits of green tea, and utilize it in the management of periodontal diseases.

TRIPHALA

Components

Triphala is a well-known powdered preparation in ayurvedic medicine used since ancient time. It consists of equal parts of Amalaki (*Emblica officinalis*), Haritaki (*Terminalia chebula*) and Bahera (*Terminalia belerica*). Amalaki is an excellent source of vitamin C and also contains carotene, nicotinic acid, D-glucose, D-fructose, riboflavin, empicol, and mucic and phyllemblic acids. Haritaki contains anthraquinone glycoside, chebulinic acid, tannic acid, terchebin, vitamin C, and arachidonic, linoleic, oleic, palmitic, and stearic acids. Bahera contains chebulagic acid, ellagic acid and its ethyl ester, gallic acid, fructose, galactose, glucose, mannitol, and rhamnose. All the three components present in triphala have a wide range of pharmacological activity and are potent antioxidants.

Role in Periodontitis

Triphala has a strong antimicrobial, antioxidant and anticollagenase properties. The antioxidants present in Triphala reduce the oxidative burden and protect cells from the damage caused by free radicals [31,32]. Bahera is the most active antioxidant followed by Amalaki and Haritaki. A clinical trial has shown that Triphala mouthwash is as efficacious as 0.2% chlorhexidine in antiplaque and anti-inflammatory activities [33].

The antibacterial effect has been assessed in an *in-vitro* study where Triphala concentrations of 50 μ g/ml inhibited *Streptococcus mutans* species. This antiplaque effect may be attributable to tannic acid present in Triphala, which is well adsorbed on the surface of bacterial cells resulting in protein denaturation and ultimately to bacterial cell death [34].

Triphala has also been known to inhibit the collagenases derived from polymorphonuclear leukocytes which are responsible for connective tissue destruction in periodontal disease. This has been corroborated by an *ex-vivo* study where tissue samples were treated with triphala, kamillosan extracts, and doxycycline, and gelatin zymography was done. Triphala showed 76.6% reduction of matrix metalloproteinase-9 (MMP-9) activity, whereas kamillosan and doxycycline showed 46.36% and 58.7%, respectively, at concentrations of 1500 μ g/ml [35].

RUBIA CORDIFOLIA

The roots of this plant have been used in ayurvedic medicine. It also contains an organic compound known as Alizarin, which gives the red color to textile dyes. Mollugin, a major component of R. cordifolia has been shown to possess antiinflammatory property [36]. A recent study showed that mollugin inhibited RANKL-induced osteoclast differentiation and bone resorbing activity of mature osteoclasts. Mollugin reduced the phosphorylation of signaling pathways activated in the early stages of osteoclast differentiation, including the MAPK, Apt, and GSK3β and inhibited the different genes associated with osteoclastogenesis such as Osteoclast-associated receptor, tartrate resistant acid phosphatase, ICAM-1, cathepsin K, DC-STAMP and OC-STAMP. Furthermore mice treated with mollugin showed significant restoration of lipopolysaccharideinduced bone loss as indicated by micro-computed tomography and histological analysis of femurs [37]. However, further studies are required to use this herbal product as a novel therapeutic approach to treat bone degenerative disorders such as periodontitis, rheumatoid arthritis, and osteoporosis.

PIPERINE

It is an alkaloid which is present in plants such as *Piper nigrum* and *Piper longum*. It is shown to have antioxidant and anti-

inflammatory properties. In an animal model, LPS stimulated mice when treated with piperine showed reductions in the nitrite level and lowered the TNF- α level. This study corroborates the free radical scavenging activity of piperine [38]. Another study on rat periodontitis model revealed that piperine significantly down-regulated the production of interleukin-1 β , MMP-8, and MMP-13. Piperine clearly inhibited alveolar bone loss and reformed trabecular microstructures in a dose-dependent manner. Histological staining showed that piperine significantly reduced the infiltration of inflammation in soft tissues [39].

SUMAC

Sumac (*Rhus coriaria*) is a well-known spice used widely as an herbal medicine for its anti-inflammatory, antimicrobial, and antioxidant properties [40]. The existing literature on sumac fruit extracts show that they has marked antioxidant activity against lipid peroxidation and free radicals *in vitro* [41,42]. In an experimental animal study, Wistar rats with ligature-induced periodontitis were administered with sumac extracts orogastrically at a dosage of 20 mg/kg/day. Serum total oxidant status (TOS) and oxidative stress index (OSI) were significantly reduced in the sumac extract treated rats. Furthermore, the serum total antioxidant status was similar to the non-ligated rats. Sumac extracts have the potential to reduce alveolar bone loss by affecting TOS and OSI levels in periodontal disease in rats [43].

GINKBO BILOBA

G. biloba (EGb) leaf extract is among the widely used herbal dietary supplement in the US [44]. It is composed of ginkgo flavone glycosides (24%), terpenoids (6%) and less than 5ppm of ginkgolic acid. Its purported biological effects include: Scavenging free radicals [45], lowering oxidative stress [46], and anti-inflammation [47]. In ligature-induced periodontitis rat model, systemic administration of EGb (28-56 mg/kg/day) resulted in reduced osteoclastic counts, decreased inflammation and induced osteoblastic activity [48].

PSIDIUM GUAJAVA

Guava has an excellent antioxidant property because it is primarily rich in Vitamin C (Ascorbic acid). It also has quercetin, carotenoids, and polyphenols which augment its antioxidant action [49,50]. Guava leaf extracts and essential oil from the stem have the ability to scavenge hydrogen peroxide, superoxide anion and inhibit the formation of hydroxyl radical [51,52]. The decoction of the root bark is recommended as a mouthwash and decoction of leaves as an effective gargle for bleeding gums [53].

CURRENT DEVELOPMENTS

Recent *in vitro* studies have shown that herbs such as *Lythrum* salicaria and Ascophyllum nodosum have shown to possess potent antioxidant properties. *L. salicaria* aqueous extracts inhibited ROS production from stimulated neutrophils which were isolated and cultured from humans [54]. A polyphenol rich

extract from A. *nodosum* demonstrated significant anti-lipid peroxidation activity and antioxidant activity by scavenging superoxide anion, hydroxyl, and peroxyl radicals [55]. Preliminary studies on a mouthwash containing microencapsulated natural extracts such as avocado oil, manuka oil, propolis oil, grapeseed extract, *Aloe vera*, green tea, coenzyme Q10 (6% GingiNat) have shown significant efficiency on plaque, gingivitis and halitosis due to its antioxidant and immunoregulatory properties [56,57]. Pradeep *et al.* showed that a gel and powder formulation derived from *Acacia arabica* demonstrated a significant improvement in plaque and gingival scores when compared to 1% chlorhexidine in gingivitis patients [58].

CONCLUSION

The herbal medicines have shown to possess a wide array of biological properties such as antimicrobial, antioxidant, and anti-inflammatory effects. The natural phytochemicals present in these herbs aid in suppressing the alveolar bone loss, which is the striking feature in periodontitis. Furthermore, the oxidative burden established due to the chronicity of the disease can be alleviated with the antioxidant property of these herbs. Although many studies, have shown the potency of herbal medicines as an alternative to conventional therapy, there still lies a void in research with respect to the clinical application of these agents in periodontics. Future targeted trials in learning the mechanism of action of these herbal remedies are warranted.

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REFERENCES

- World Health Organization. Traditional Medicine. Geneva; 2008. Available from: http://www.who.int/medicines/areas/traditional/en/. [Last Accssed date 2015 Nov 01].
- World Health Organization. The World Medicines Situation 2011. Traditional Medicines: Global Situation, Issues and Challenges. Geneva: WHO; 2011. p. 1-14.
- Tambekar DH, Dahikar SB, Lahare MD. Antibacterial potentials of some herbal preparations available in India. Res J Med Med Sci 2009;4:224-7.
- Listgarten MA. Pathogenesis of periodontitis. J Clin Periodontol 1986;13:418-30.
- 5. Laine ML, Crielaard W, Loos BG. Genetic susceptibility to periodontitis. Periodontol 2000 2012;58:37-68.
- Matthews AT, Ross MK. Oxyradical stress, endocannabinoids, and atherosclerosis. Toxics 2015;3:481-498.
- Fischer BM, Voynow JA, Ghio AJ. COPD: Balancing oxidants and antioxidants. Int J Chron Obstruct Pulmon Dis 2015;10:261-76.
- D'Aiuto F, Nibali L, Parkar M, Patel K, Suvan J, Donos N. Oxidative stress, systemic inflammation, and severe periodontitis. J Dent Res 2010;89:1241-6.
- Bullon P, Newman HN, Battino M. Obesity, diabetes mellitus, atherosclerosis and chronic periodontitis: A shared pathology via oxidative stress and mitochondrial dysfunction? Periodontol 2000 2014;64:139-53.
- Devasagayam TP, Tilak JC, Boloor KK, Sane KS, Ghaskadbi SS, Lele RD. Free radicals and antioxidants in human health: Current status and future prospects. J Assoc Physicians India 2004;52:794-804.
- 11. Nagata M. Inflammatory cells and oxygen radicals. Curr Drug Targets Inflamm Allergy 2005;4:503-4.

- Garrett IR, Boyce BF, Oreffo RO, Bonewald L, Poser J, Mundy GR. Oxygen-derived free radicals stimulate osteoclastic bone resorption in rodent bone *in vitro* and *in vivo*. J Clin Invest 1990;85:632-9.
- Bax BE, Alam AS, Banerji B, Bax CM, Bevis PJ, Stevens CR, *et al.* Stimulation of osteoclastic bone resorption by hydrogen peroxide. Biochem Biophys Res Commun 1992;183:1153-8.
- Lean JM, Jagger CJ, Kirstein B, Fuller K, Chambers TJ. Hydrogen peroxide is essential for estrogen-deficiency bone loss and osteoclast formation. Endocrinology 2005;146:728-35.
- Giannopoulou C, Krause KH, Müller F. The NADPH oxidase NOX2 plays a role in periodontal pathologies. Semin Immunopathol 2008;30:273-8.
- Halliwell B. Antioxidants: The basics What they are and how to evaluate them. Adv Pharmacol 1997;38:3-20.
- Chapple IL, Mason GI, Garner I, Matthews JB, Thorpe GH, Maxwell SR, *et al.* Enhanced chemiluminescent assay for measuring the total antioxidant capacity of serum, saliva and crevicular fluid. Ann Clin Biochem 1997;34:412-21.
- Sculley DV, Langley-Evans SC. Periodontal disease is associated with lower antioxidant capacity in whole saliva and evidence of increased protein oxidation. Clin Sci (Lond) 2003;105:167-72.
- Brock GR, Butterworth CJ, Matthews JB, Chapple IL. Local and systemic total antioxidant capacity in periodontitis and health. J Clin Periodontol 2004;31:515-21.
- Shirzaiy M, Ansari SM, Dehghan JH, Ghaeni SH. Total anti-oxidant capacity of saliva in chronic periodontitis patients before and after periodontal treatment. J Nepal Health Res Counc 2014;12:172-6.
- Cabrera C, Artacho R, Giménez R. Beneficial effects of green tea A review. J Am Coll Nutr 2006;25:79-99.
- Asahi Y, Noiri Y, Miura J, Maezono H, Yamaguchi M, Yamamoto R, *et al.* Effects of the tea catechin epigallocatechin gallate on *Porphyromonas gingivalis* biofilms. J Appl Microbiol 2014;116:1164-71.
- Araghizadeh A, Kohanteb J, Fani MM. Inhibitory activity of green tea (*Camellia sinensis*) extract on some clinically isolated cariogenic and periodontopathic bacteria. Med Princ Pract 2013;22:368-72.
- Okamoto M, Leung KP, Ansai T, Sugimoto A, Maeda N. Inhibitory effects of green tea catechins on protein tyrosine phosphatase in *Prevotella intermedia*. Oral Microbiol Immunol 2003;18:192-5.
- Sakanaka S, Aizawa M, Kim M, Yamamoto T. Inhibitory effects of green tea polyphenols on growth and cellular adherence of an oral bacterium, *Porphyromonas gingivalis*. Biosci Biotechnol Biochem 1996;60:745-9.
- Makimura M, Hirasawa M, Kobayashi K, Indo J, Sakanaka S, Taguchi T, et al. Inhibitory effect of tea catechins on collagenase activity. J Periodontol 1993;64:630-6.
- Tominari T, Matsumoto C, Watanabe K, Hirata M, Grundler FM, Miyaura C, *et al.* Epigallocatechin gallate (EGCG) suppresses lipopolysaccharide-induced inflammatory bone resorption, and protects against alveolar bone loss in mice. FEBS Open Bio 2015;5:522-7.
- Hrishi T, Kundapur P, Naha A, Thomas B, Kamath S, Bhat G. Effect of adjunctive use of green tea dentifrice in periodontitis patients - A randomized controlled pilot study. Int J Dent Hyg 2015.
- Hirasawa M, Takada K, Makimura M, Otake S. Improvement of periodontal status by green tea catechin using a local delivery system: A clinical pilot study. J Periodontal Res 2002;37:433-8.
- Kaur H, Jain S, Kaur A. Comparative evaluation of the antiplaque effectiveness of green tea catechin mouthwash with chlorhexidine gluconate. J Indian Soc Periodontol 2014;18:178-82.
- Vani T, Rajani M, Sarkar S, Shishoo CJ. Antioxidant properties of the ayurvedic formulation triphala and its constituents. Pharm Biol 1997;35:313-7.
- Padmawar A, Bhadoriya U. Phytochemical investigation and comparative evaluation of in vitro free radical scavenging activity of triphala and curcumin. Asian J Pharm Med Sci 2011;1:9-12.
- Naiktari RS, Gaonkar P, Gurav AN, Khiste SV. A randomized clinical trial to evaluate and compare the efficacy of triphala mouthwash with 0.2% chlorhexidine in hospitalized patients with periodontal diseases. J Periodontal Implant Sci 2014;44:134-40.
- Jagdish L, Anand Kumar VK, Kaviyarasan V. Effect of triphala on dental biofilm. Indian J Sci Technol 2009;2:30-3.
- Abraham S, Kumar MS, Sehgal PK, Nitish S, Jayakumar ND. Evaluation of the inhibitory effect of triphala on PMN-type matrix metalloproteinase (MMP-9). J Periodontol 2005;76:497-502.
- 36. Zhu ZG, Jin H, Yu PJ, Tian YX, Zhang JJ, Wu SG. Mollugin inhibits the

inflammatory response in lipopolysaccharide-stimulated RAW264.7 macrophages by blocking the Janus kinase-signal transducers and activators of transcription signaling pathway. Biol Pharm Bull 2013;36:399-406.

- Baek JM, Kim JY, Jung Y, Moon SH, Choi MK, Kim SH, *et al*. Mollugin from *Rubea cordifolia* suppresses receptor activator of nuclear factorκB ligand-induced osteoclastogenesis and bone resorbing activity in vitro and prevents lipopolysaccharide-induced bone loss *in vivo*. Phytomedicine 2015;22:27-35.
- Pradeep CR, Kuttan G. Effect of piperine on the inhibition of nitric oxide (NO) and TNF-alpha production. Immunopharmacol Immunotoxicol 2003;25:337-46.
- Dong Y, Huihui Z, Li C. Piperine inhibit inflammation, alveolar bone loss and collagen fibers breakdown in a rat periodontitis model. J Periodontal Res 2015;50:758-65.
- 40. Rayne S, Mazza G. Biological activities of extracts from sumac (Rhus spp.): A review. Plant Foods Hum Nutr 2007;62:165-75.
- Bursal E, Koksal E. Evaluation of reducing power and radical scavenging activities of water and ethanol extracts from sumac (*Rhus coriaria* L.) Food Res Int 2011;44:2217-21.
- Candan F, Sökmen A. Effects of *Rhus coriaria* L (Anacardiaceae) on lipid peroxidation and free radical scavenging activity. Phytother Res 2004; 18:84-6.
- Saglam M, Köseoglu S, Hatipoglu M, Esen HH, Köksal E. Effect of sumac extract on serum oxidative status, RANKL/OPG system and alveolar bone loss in experimental periodontitis in rats. J Appl Oral Sci 2015;23:33-41.
- 44. Shankland WE 2nd. Four common herbs seen in dental practice: Properties and potential adverse effects. Cranio 2009;27:118-24.
- Yuan F, Yu R, Yin Y, Shen J, Dong Q, Zhong L, *et al.* Structure characterization and antioxidant activity of a novel polysaccharide isolated from *Ginkgo biloba*. Int J Biol Macromol 2010;46:436-9.
- Wang CG, Dai Y, Li DL, Ma KY. *Ginkgo biloba* leaf extract action in scavenging free radicals and reducing mutagenicity and toxicity of cigarette smoke *in vivo*. J Environ Sci Health A Tox Hazard Subst Environ Eng 2010;45:498-505.
- Thorpe LB, Goldie M, Dolan S. Central and local administration of *Gingko biloba* extract EGb 761[®] inhibits thermal hyperalgesia and inflammation in the rat carrageenan model. Anesth Analg 2011;112:1226-31.
- Sezer U, Kara MI, Erciyas K, Ozdemir H, Üstün K, Ozer H, et al. Protective effects of Ginkgo biloba extract on ligature-induced

periodontitis in rats. Acta Odontol Scand 2013;71:38-44.

- Qian H, Nihorimbere V. Antioxidant power of phytochemicals from *Psidium guajava* leaf. J Zhejiang Univ Sci 2004;5:676-83.
- Dakappa SS, Adhikari R, Timilsina SS, Sunita S. A review on the medicinal plant *Psidium guajava* Linn. (Myrtaceae). J Drug Deliv Ther 2013;3:162-8.
- Ogunlana OE, Ogunlana OO. *In vitro* assessment of the free radical scavenging activity of *Psidium guajava*. Res J Agric Biol Sci 2008;4:666-71.
- Chen HY, Yeh GC. Antioxidant activity and free radical-scavenging capacity of extracts from guava (*Psidium guajava* L.) leaves. Food Chem 2007;101:686-94.
- Mittal P, Gupta V, Kaur G, Garg AK, Singh A. Phytochemistry and pharmacological activities of *Psidium guajava*: A review. Int J Pharm Sci Res 2010;1:9-19.
- Piwowarski JP, Kiss AK. Contribution of C-glucosidic ellagitannins to *Lythrum salicaria* L. influence on pro-inflammatory functions of human neutrophils. J Nat Med 2015;69:100-10.
- Tamanai-Shacoori Z, Chandad F, Rébillard A, Cillard J, Bonnaure-Mallet M. Silver-zeolite combined to polyphenol-rich extracts of *Ascophyllum nodosum*: Potential active role in prevention of periodontal diseases. PLoS One 2014;9:e105475.
- Mouhyi J, Del Corso M, Hippolyte MP, Sammartino G, Dohan Ehrenfest DM. Mouthwash solutions containing microencapsuled natural extracts: *In vitro* evaluation of antioxidant properties (dental plaque and gingivitis). Rev Stomatol Chir Maxillofac 2010;111:140-3.
- Mouhyi J, Del Corso M, Hippolyte MP, Sammartino G, Dohan Ehrenfest DM. Mouthwash solutions containing microencapsulated natural extracts: Clinical results on dental plaque and gingivitis. Rev Stomatol Chir Maxillofac 2010;111:144-7.
- Pradeep AR, Agarwal E, Bajaj P, Naik SB, Shanbhag N, Uma SR. Clinical and microbiologic effects of commercially available gel and powder containing *Acacia arabica* on gingivitis. Aust Dent J 2012;57:312-8.

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