

Antidiabetic phytoconstituents and their mode of action on metabolic pathways

Sudhanshu Kumar Bharti, Supriya Krishnan, Ashwini Kumar and Awanish Kumar 🕒



Abstract: Diabetes Mellitus, characterized by persistent hyperglycaemia, is a heterogeneous group of disorders of multiple aetiologies. It affects the human body at multiple organ levels thus making it difficult to follow a particular line of the treatment protocol and requires a multimodal approach. The increasing medical burden on patients with diabetes-related complications results in an enormous economic burden, which could severely impair global economic growth in the near future. This shows that today's healthcare system has conventionally been poorly equipped towards confronting the mounting impact of diabetes on a global scale and demands an urgent need for newer and better options. The overall challenge of this field of diabetes treatment is to identify the individualized factors that can lead to improved glycaemic control. Plants are traditionally used worldwide as remedies for diabetes healing. They synthesize a diverse array of biologically active compounds having antidiabetic properties. This review is an endeavour to document the present armamentarium of antidiabetic herbal drug discovery and developments, highlighting mechanism-based antidiabetic properties of over 300 different phytoconstituents of various chemical categories from about 100 different plants modulating different metabolic pathways such as glycolysis, Krebs cycle, gluconeogenesis, glycogen synthesis and degradation, cholesterol synthesis, carbohydrate metabolism as well as peroxisome proliferator activated receptor activation. dipeptidyl peptidase inhibition and free radical scavenging action. The aim is to provide a rich reservoir of pharmacologically established antidiabetic phytoconstituents with specific references to the novel, cost-effective interventions, which might be of relevance to other lowincome and middle-income countries of the world.

Keywords: antidiabetics, diabetes mellitus, hyperglycaemia, metabolism, phytoconstituents

Received: 1 August 2017; accepted in revised form: 1 November 2017.

Introduction

Diabetes mellitus (DM) is the most common endocrine disorder resulting from a defect in insulin secretion, insulin resistance or both. It is the third leading cause of morbidity and mortality, after heart attack and cancer. In 2015, about 415 million people had diabetes in the world and 78 million people in the Southeast Asia (SEA) region; by 2040 this will rise to 140 million. India is one of the epicentres of the global DM pandemic. There were 69.1 million cases of diabetes in India in 2015.^{1,2} DM characterized by persistent hyperglycaemia is a heterogeneous group of disorders of multiple aetiologies that affect the human body at multiple organ levels, thus making it difficult to

follow a particular line of treatment. The treatment protocol requires a multimodal approach which should be personalized so that it varies from person to person.3 In general, DM is classified into two categories: type 1 and type 2. In type 1 diabetes (T1DM), hormone insulin is not produced due to the destruction of pancreatic β cells, while type 2 diabetes (T2DM) is characterized by a progressive impairment of insulin secretion by pancreatic β cells and by a relative decreased sensitivity of target tissues to the action of this hormone. T2DM leads to other pathological consequences like cardiovascular disorders, nephropathy, neuropathies and the patient becomes prone to a number of infections too.4 The increasing medical burden on

Ther Adv Endocrinol

2018, Vol. 9(3) 81-100

DOI: 10 1177/ 2042018818755019

© The Author(s), 2018. Reprints and permissions: http://www.sagepub.co.uk/ iournalsPermissions.nav

Correspondence to: Awanish Kumar Department of Biotechnology, National Institute of Technology, GE Road, Raipur Chhattisgarh, 492010, India

drawanishkr@gmail.com awanik.bt@nitrr.ac.in

Sudhanshu Kumar Bharti Department of

Biochemistry, Patna University, Patna, Bihar,

Supriya Krishnan Department of PMIR, Patna University, Patna, Bihar, India

Ashwini Kumar Department of Biotechnology, National Institute of Technology. Raipur, Chhattisgarh, India

patients with diabetes-related complications also results in an enormous economic burden, which could severely impair global economic growth in the near future. This shows that today's health system has conventionally been poorly equipped to confront the mounting impact of diabetes on a global scale and demands an urgent need for newer and better options. The overall challenge of this field of diabetic treatment is to identify the individualized factors that can lead to improved glycaemic control.

Besides conventional oral and injectable medications, diabetes treatments include diet modification, regular exercising, lifestyle changes, weight regulation and other alternatives or add on therapies such as herbal therapy.^{5,6} Herbal drugs are prescribed widely as drugs of choice because of their effectiveness, few side effects and relatively low cost.⁷

In present day modern science, concoctions or crude extract based studies are losing their significance and the focus has, for the better, shifted towards discovery and exploitation of specific compounds for their therapeutic Knowledge about specific compounds from various herbal (plant) parts makes the experimental studies easier and helps to focus on better understanding the mechanism of action and future therapeutic potential. Since diabetes is a multifaceted disease with an effect on almost all the organs,4 exploitation of plant resources for better therapeutic molecules needs a boost in research and development. Another advantage of exploiting plant-based resources is the time and money saving since it will surpass the need for drug design and screening. This review documents the present armamentarium of antidiabetic herbal drug discovery and developments, highlighting the mechanism-based antidiabetic properties of over 300 different phytoconstituents of various chemical categories from about 100 different plants modulating different metabolic pathways. The aim is to provide a rich reservoir of pharmacologically established antidiabetic phytoconstituents with specific references to the novel, cost-effective interventions, which might be of relevance to other low-income and middleincome countries of the world.

Materials and methods

This review article is a compilation of the current knowledge and future expectation of various chemical categories of phytoconstituents with

their mode of actions on a single platform, which have been shown to display potent hypoglycaemic activity against DM. We have searched the literature using PubMed, SCOPUS, MEDLINE, and Google scholar with the key words 'diabetes, antidiabetic phytoconstituents, metabolism, their mode of action on metabolic pathways, and induction' to prepare this review article. The literature search included only articles written in the English language. The references lists of all listed articles were searched manually to obtain relevant and additional information. Review and original research articles published between 1984 and 2017 (in English) were included in this review as a reference. The selection of phytoconstituents in this review was on the basis of their antidiabetic activity and ethanopharmacological use.

Carbohydrate metabolism: problem statement

Metabolism in the living system is concerned with managing the material and energy resources within cells involving complex molecules like carbohydrates, lipids and proteins as chief substrates. After a normal meal, the transient increase in plasma glucose, amino acids, triglycerols and chylomicrons is responded to by increased secretion of insulin from pancreatic islet cells, thus enhancing the synthesis of triacylglycerols, glycogen and protein. During this period virtually all tissues use glucose as a fuel.8 Problems with glucose and carbohydrate metabolism are quite rare in cultures adhering to a primitive diet, one low in refined foods, starches and sugars. Although hereditary predispositions, viral and bacterial afflictions of the pancreas and autoantibodies to pancreatic islets do contribute to the development of this disorder, diet, lifestyle and obesity are by far the most significant risk factors.9 In the following section various pathways in which glucose is involved, either as a substrate or liberated as a product, are discussed and the corresponding plant-derived drugs that inhibit or activate the steps in these pathways are listed.

Therapy and management of DM

A combination of side effects, contraindications and lack of effect of synthetic drugs on disease progression highlight the need for newer therapies that minimize the frequency and severity of DM exacerbations. ¹⁰ The plant kingdom historically has been the driving force for the development of novel drugs. Herbal products have been thought

to be inherently safe because of their natural origin and traditional use rather than systemic studies designed to detect adverse effects. Approximately 80% of the world's population relies on biomedicines for their health and wellbeing.11,12 According to ethnobotanical information based on Indian Pharmacopoeia, about 1200 plants with antidiabetic properties have been cited. Of these, around 400 plants and their products have been documented to have antidiabetic properties after significant investigation.¹³ There are unique theories for concepts of aetiology, systems of diagnosis and treatment for plant-derived drugs, which are vital to using them in practice. The mechanisms of action of plant-derived drugs involve regulating glycaemic metabolism, decreasing cholesterol levels, eliminating free radicals, increasing secretion of insulin and improving microcirculation.14 With the background that phytoconstituents form the mainstay of therapy and management of DM, this paper reviews the common Indian antidiabetic plants and their constituents.

Phytoconstituents and their antidiabetic effects

Plants contain numerous chemical compounds having medicinal values and include alkaloids, amino acids, amines and carboxylic acid derivatives, anthranoids, carbohydrates, glycosides, flavanoids, minerals, vitamins and inorganic compounds, peptidoglycans, polyphenol and its derivatives, saponins, and so on.15 These compounds are extracted from different parts of the various plants (root, stem, leaf, flower, fruit, etc.) (Table 1). This review aims to document and summarize the present knowledge about the mechanism-based action of antidiabetic plants, with emphasis on their phytoconstituents that target the various metabolic pathways in humans. The review has been organized according to various categories of phytoconstituents, targeted metabolic pathways and plant sources in Table 1 (A–J), which are also shown in Figures 1–4 at different steps with arrows and phytoconstituent numbers (A-1, B-6, J-2, etc.). Figures 1-4 clearly show the action of various phytoconstituents discussed in this review (Table 1) at different steps of various metabolic pathways.

Alkaloids

A large number of alkaloids have been isolated from numerous medicinal plants and investigated

by the researchers for their possible antidiabetic activity. 112 Glycolysis is the hub of carbohydrate metabolism because virtually all sugars (whether arising from the diet or from catabolic reactions in the body) ultimately can be converted to glucose *via* a series of 10 reactions with three regulatory steps catalysed by the enzymes hexokinase, phosphofructokinase and pyruvate kinase. The alkaloid berberine, extracted from *Tinospora cordifolia*, enhances the activity of hexokinase and phosphofructokinase, resulting in glucose transport, carbohydrate digestion and absorption. 16

Carbohydrates are the major constituents of the normal diet of humans. Starch and sucrose are its major forms, which supply about 70-80% of the energy requirement to the body. Their digestion starts in the mouth and continues even in the small intestine producing glucose, which is absorbed into the bloodstream through the walls of the intestine, and finally it is transported to different parts of the body through the liver. The digested products are mainly glucose with small amounts of fructose and galactose. Starch is first decomposed into oligosaccharides by the enzyme α-amylase, found in saliva and pancreatic juices. A membrane-bound enzyme α-glucosidase, in the epithelium of the small intestine, catalyses the cleavage of glucose from disaccharides and oligosaccharides. Hence, α -glucosidase inhibition is one of the effective treatments for diabetes since it will delay the time of absorption of glucose.3 There are a number of phytoconstituents known to suppress the activity of α -glucosidase and inhibit the absorption of glucose in both the small intestine and kidney, so that the concentration of glucose in the blood remains constant after a meal. The α -glucosidase inhibitors slow the digestion of starch in the small intestine, so that glucose enters the bloodstream more slowly and can be matched to an impaired insulin response or production. Gluconeogenesis is a ubiquitous multistep process occurring in the liver and kidnev in which pyruvate or a related three-carbon compound like lactate, alanine, is converted to glucose. Seven of the 10 enzymatic reactions of gluconeogenesis are the reverse of glycolysis with four regulatory steps that are catalysed by the enzymes pyruvate carboxylase, phosphoenolpyruvate carboxykinase, fructose-1,6-bisphosphatase and glucose-6-phosphatase. During gluconeogenesis a phytoconstituent barberine reduces the activity of glucose-6-phosphatase enzyme which affects the conversion of d-glucose from glucose-6-phosphate.

Table 1. Chemical categorization of various phytoconstituents having hypoglycaemic potential that regulate intermediates of different metabolic pathways.

Sl. No.	Phytoconstituents	Targeted metabolic pathways	Plant source	References
A	Alkaloids			
1	Barberin	Glucose transport, carbohydrate digestion and absorption, DPP-IV inhibition	Tinospora cordifollia, Barberisaristata	Singh <i>et al.</i> , ¹⁶ Al masri <i>et al.</i> ¹⁷
2	Catharanthine, vindoline, vindolinene vinblastine, vincristine	Free radical scavenging action	Cathanthrus roseus, Vinca rosea	Chattopadhyay, ¹⁸ Jarald <i>et al.</i> , ¹⁹ Kar <i>et al</i> . ²⁰
3	Sotolon [4,5-dimethyl-3-hydroxy-2(5H)-furanone], trigonelline, gentianine, carpaine compounds	Glucose transport, carbohydrate digestion and absorption	Trigonella foenum graecum	Hui <i>et al.,</i> ⁶ Khosla <i>et al.</i> ²¹
4	Ginkgolides	Insulin secretion	Ginkgo biloba	Pinto et al., ²²
5	Allylpropyl disulfide	Glycogen synthesis, insulin secretion	Allium sativum	Sheela <i>et al.</i> , ²³ Kumari and Augusti ²⁴
6	Aegelin, marmesin, marmelosin	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Aegle marmelos	Kamalakkannan and Prince, ²⁵ Ponnachan <i>et al.</i> ²⁶
7	Harmine, pinoline	Insulin secretion and $\beta\text{-cell}$ regeneration	Tribulus terrestris	Cooper <i>et al.</i> , ²⁷ Kirtikar and Basu ²⁸
8	Betaine, achyranthine, β -ecdysone	Carbohydrate digestion and absorption	Achyranthus aspera	Akhtar and Iqbal ²⁹
9	Castanospermine, epifagomine,	Carbohydrate digestion and absorption, insulin secretion	Xanthocercis zambesiaca	Akhtar ³⁰
10	Castanospermine, australine	DPP-IV inhibition	Castanospermum australe	Bharti <i>et al.</i> , ¹¹ Orwa <i>et al.</i> ³¹
В	Amino acids, amines and car	boxylic acid derivatives		
1	Allicin, apigenin, alliin	Cholesterol synthesis, glycogen synthesis	Allium sativum	Gholap and Kar, ³² Kumar and Reddy ³³
2	Gurmarin, betaine, choline, trimethylamine	Regeneration of pancreatic β cells and insulin secretion	Gymnema sylvestre	Sugihara <i>et al.</i> , ³⁴ Preuss <i>et al</i> . ³⁵
3	(-) Hydroxycitric acid	Insulin secretion	Garcinia cambogia, Gymnema sylvestre	Preuss <i>et al.</i> , ³⁵ Hayamizu <i>et al</i> . ³⁶
5	Ferulic acid	Free radical scavenging activity, insulin secretion	Curcuma longa	Ohnishi <i>et al</i> . ³⁷
6	Leucine, isoleucine, alanin	Insulin secretion	Aloe vera	Ajabnoor ³⁸
7	Mallic acid, chlorogenic acid	Krebs cycle	Caralluma edulis, Syzygium cumini, Acacia Arabica	Wadood and Shah ³⁹
8	4-Hydroxyisoleucine, n-hydroxyisoleucine	Glucose transport, carbohydrate metabolism	Trigonella foenum graecum	Hui <i>et al.,⁶</i> Khosla <i>et al.</i> ²¹
9	Polypeptide-P	Insulin secretion, glycogen synthesis	Momordica charantia	Chao and Huang, ⁴⁰ Sarkar <i>et al</i> . ⁴¹
10	S-methyl cysteine sulfoxide, S-allyl cysteine sulfoxide	Glycolysis, cholesterol synthesis	Alium sepa	Kumari and Augusti, ²⁴ Roman-Ramos <i>et al</i> . ⁴²
11	Nitrosamines	Carbohydrate digestion and absorption	Areca catechu	Mannan <i>et al</i> . ⁴³

Table 1. (Continued)

Sl. No.	Phytoconstituents	Targeted metabolic pathways	Plant source	References
12	Brevifolin carboxylic acid, ethyl brevifolin carboxylate	Carbohydrate digestion and absorption	Phyllanthus amarus	Ali et al. ⁴⁴
13	Lectins, mistletoe lectin I, II, III, viscotoxin B, cycliton	Insulin secretion, glycogen synthesis	Viscum album	Adaramoye <i>et al.</i> , ⁴⁵ Eno <i>et al.</i> , ⁴⁶ Gray and Flatt ⁴⁷
14	Furfural, caprylic acid	Insulin secretion, glycogen synthesis	Agaricus campestris	Manohar <i>et al.</i> , ⁴⁸ Gray and Flatt ⁴⁹
15	Procyanidins	Antihyperglycaemic	Grape seed	Pinent <i>et al.</i> ⁵⁰
16	Bis (2-ethyl hexyl) phthalate (DEHP)	Insulin secretion, glycogen synthesis	Cassia auriculata	Abesundara <i>et al.</i> ⁵¹
17	Raisin	Insulinonematic activity	Vitis vinifera	Rankin <i>et al.</i> ⁵²
С	Anthranoids			
1	Aloin, barbaloin, isobarbaloine, aloetic acid, aloe-emodin, emodin, cinnamic acid, crysophanic acid	Insulin secretion and synthesis	Aloe vera, Cassia tora	Ajabnoor ³⁸
2	Vicine	Insulin secretion	Momordica charantia	Chao and Huang, ⁴⁰ Sarkar <i>et al</i> . ⁴¹
3	Torachrysone, toralactone, rhein, alaternin	Insulin secretion	Cassia tora	Nam and Choi ⁵³
4	Camphor, eugenol, trans-β- ocimene, geraniol, α-pinene, limonene, p-cymene, 1,8-cineole, thujone	Insulin secretion, regeneration of pancreatic β cells	Ocimum canum, Coriandrum sativum, Artemisia roxburghiana, Syzygium aromaticum	Hannan <i>et al.</i> , ⁵⁴ Hussain <i>et al.</i> , ⁵⁵ Broadhurst <i>et al</i> . ⁵⁶
D	Carbohydrates			
1	Glucomannan	Insulin secretion, carbohydrate digestion	Aloe vera	Van de Venter <i>et al</i> . ⁵⁷
2	Caryophylline	Insulin secretion, carbohydrate digestion and absorption	Ocimum sanctum, Syzygium aromaticum	Van de Venter <i>et al.</i> ⁵⁷
3	Protein-bound polysaccharide	Insulin secretion, carbohydrate digestion and absorption	Alpinia galangal, Aloe vera, Ocimum sanctum	Van de Venter <i>et al.</i> ⁵⁷
4	Guar gum, pectin and pectin fibres, mucilaginous fibre	Glucose transport, carbohydrate metabolism, stabilizing agents	Trigonella foenum graecum, Citrus sinensis, Coccinia indica	Kar <i>et al.</i> , ²⁰ Nandini <i>et al</i> . ⁵⁸
5	Cellulose, mannose	Carbohydrate digestion and absorption	Aloe vera	Van de Venter <i>et al</i> . ⁵⁷
6	D-threitol, D-arabinitol, palmitic acid	Carbohydrate digestion and absorption	Hericium erinaceus	Khan <i>et al</i> ., ⁵⁹ Liang <i>et al</i> . ⁶⁰
7	L-arabino-D-xylan, cinnzeylanin, cinnzeylanol, D-glucan	Carbohydrate digestion and absorption	Cinnamomum zeylanicum	Solomon and Blannin ⁶¹
8	Mucopolysaccharide	Carbohydrate metabolism, cholesterol synthesis	Opuntia ficus indica	Godard <i>et al.</i> ⁶²
9	Inulin, laevulin	Glucose transport, carbohydrate digestion and absorption	Taraxacum officinale	Godard <i>et al.</i> , ⁶² Onal <i>et al</i> . ⁶³

(Continued)

Table 1. (Continued)

Sl. No.	Phytoconstituents	Targeted metabolic pathways	Plant source	References
10	Fructo-oligosaccharide	Decrease glycosuria and AGEs	Aureobasidium pullulans	Bharti <i>et al.</i> ¹²
E	Glycosides			
1	Gymnemic acid, gymnemosides	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Gymnema sylvestre	Sugihara <i>et al.</i> , ³⁴ Preuss <i>et al</i> . ³⁵
2	Vin α -ginsenoside R3	Insulin secretion	Panax quinquefolium	Vuksan <i>et al.</i> ⁶⁴
3	Astragalin, scopolin, skimmin, roscoside II	Regeneration of pancreatic β cells and insulin secretion	Morus alba	Gulubova and Boiadzhiev ⁶⁵
4	C-glycosides	Glucose transport, carbohydrate metabolism	Trigonella foenum graecum	Gupta <i>et al.</i> , ⁶⁶ Kluwer ⁶⁷
5	Momordin, momordicine, charantin	Insulin secretion, glycogen synthesis	Momordica charantia	Chao and Huang, ⁴⁰ Sarkar <i>et al</i> . ⁴¹
6	Tinosporine, cordifolide, tinosporide, cordifole, columbin	Cholesterol synthesis, glycolysis	Tinospora cordifollia, Tinospora crispa	Hui <i>et al.</i> , ⁶ Kar <i>et al.</i> , ²⁰ Van de Venter <i>et al.</i> , ⁵⁷ Noor and Ashcroft ⁶⁸
7	Momorcharaside A and B, momorcharin A and B	Insulin secretion, glycogen synthesis	Momordica charantia	Chao and Huang, ⁴⁰ Sarkar <i>et al</i> . ⁴¹
8	Cucurbitacin B, isocucurbitacin B	Insulin secretion, glycogen synthesis	Helicteres isora	Lemus <i>et al.</i> ⁶⁹
9	Momordin-a, luffin-a	Insulin secretion, glycogen synthesis	Luffa cylindnica	Lemus <i>et al.</i> ⁶⁹
10	Kotalanol, salacinol	Insulin secretion, glycogen synthesis	Salacia reticulate, Salacia oblonga	Huang <i>et al.</i> ⁷⁰
11	Arbutin, eriolin	Insulin secretion, glycogen synthesis	Arctostaphylos uvaursi	Moon et al. ⁷¹
12	Citrullol, colocynthin, elaterin, elatericin B, colosynthetin	Insulin secretion, glycogen synthesis	Cifrullus colocynthis	González-Tejero <i>et al.</i> , ⁷² Ziyyat <i>et al.</i> ⁷³
13	Leucocyanidin, pelarogonidin	Insulin secretion, glycogen synthesis	Ficus bengalensis	Singh <i>et al.</i> , ⁷⁴ Cherian <i>et al.</i> , ⁷⁵ Kumar <i>et al.</i> ⁷⁶
14	Taraxacin	Insulin secretion	Taraxacum officinale	Broadhurst et al. ⁵⁶
F	Flavanoids			
1	Chrysin, isoquercitrin	Insulin secretion	Morus alba	Roman-Ramos et al. ⁴²
3	Epigallocatechin-gallate, gallocatechin, epicatechin, (+) catechin, (-) epicatechin	Free radical scavenging activity, insulinonematic activity	Camellia sinensis, Punica granatum, Satureja khuzestanica, Bauhinia forficata	Hii and Howell, ⁷⁷ Waltner-Law <i>et al.</i> , ⁷⁸ Vessal <i>et al</i> ., ⁷⁹ Li <i>et al</i> . ⁸⁰
4	Myrciaphenones A and B, myrciacitrins I and II	Insulin secretion	Myrcia multiflora	Chattopadhyay, ¹⁸ Ngueyem <i>et al.</i> ⁸¹
5	$\alpha\text{-}Cephalin,myricetin\text{-}3'\text{-}$ glucoside, ambrettolide	Insulin secretion	Abelmoschus moschatus	Chattopadhyay, ¹⁸ Ngueyem <i>et al</i> . ⁸¹
6	Cytrus bioflavonoids (hesperidin, naringin)	Glycogen synthesis, glycolysis, gluconeogenesis	Camellia sinensis	Jung <i>et al.</i> ⁸²
7	Flavanols, flavones, flavanones	Insulin secretion	Panax notoginseng	Liu <i>et al.</i> , ⁸³ Vuksan <i>et al</i> . ⁶⁴

Table 1. (Continued)

Sl. No.	Phytoconstituents	Targeted metabolic pathways	Plant source	References
8	Quercetin, quercetrin, apigenin, rutin, apigenin-7- O-glucoside	Insulin secretion	Urtica dioica, Bauhinia varigtla, Ginkgo biloba	Hussain <i>et al.</i> , ⁵⁵ Jellin <i>et al</i> . ⁸⁴
9	Naringenin	Insulin secretion	Camellia sinensis	Waltner-Law et al. ⁷⁸
10	Soy isoflavones (genistein, diadzein)	Lipid and glucose metabolism, PPAR activation	Glycin max, Curcuma longa	Howes <i>et al.</i> , ⁸⁵ Mezei <i>et al</i> . ⁸⁶
11	Proanthocyanidins	Insulinonematic activity	Vitis vinifera	Gray and Flatt, ⁴⁹ Pinent <i>et al.</i> , ⁵⁰ Rankin <i>et al</i> . ⁵²
12	lpha-Terpineol, hexanol	Insulin secretion	Agaricus campestris	Gray and Flatt, ⁴⁹ Pinent <i>et al.</i> , ⁵⁰ Rankin <i>et al.</i> ⁵²
13	Kaempferitrin	Glycolysis	Bauhinia candicans, Bauhinia forficata	Lemus <i>et al.,⁶⁹</i> Jorge <i>et al.⁸⁷</i>
15	(+) Catechin, (–) epicatechin, chiorogenic acid, liquiritigenin, isoliquiritigerin	Insulinomematic activity	Phylanthus embelica, Acacia Arabica, Pterocarpus marsupium, Phylanthus embelica	Grover and Vats, ⁷ Kar <i>et al.</i> , ²⁰ Wadood and Shah, ³⁹ Van de Venter <i>et al</i> . ⁵⁷
16	Silymarin, silybin, silychristin, silidianin	HMG Co A suppression	Silybum marianum	Huseini <i>et al</i> . ⁸⁸
17	Kaempferol, isorhamnetin	Free radical scavenging activity	Ginkgo biloba	Jellin <i>et al.</i> ⁸⁴
18	Amarogentin, swerchirin, chirantin, gentiopicrin	Insulin secretion, glycogen synthesis	Swertia chirayita	Van de Venter <i>et al</i> . ⁵⁷
19	Tribulusamides A and B, kaempferol-3-β-D-(6'P- coumaroyl)glucoside, kaempferol-3-glucoside	Insulin secretion, free radical scavenging activity	Tribulus terrestris	Cooper <i>et al.</i> , ²⁷ Kirtikar and Basu ²⁸
20	Shamimin	Insulin secretion	Biophytum sensitivum	Puri and Baral, ⁸⁹ Puri <i>et al.</i> ⁹⁰
21	Leucopelargonidin, dulcitol	Insulin secretion	Casearia esculenta	Prakasam <i>et al.</i> 91
22	Matteuorien, matteuorienin matteuorienate A, B, C	Insulin secretion	Matteuccia orientalis	Shane-McWhorter ⁹²
G	Minerals, vitamins and inorg	anic compounds		
1	Zinc	Insulin secretion	Aloe vera	Wijesekara <i>et al.</i> , ⁹³
3	Vitamin A,E	Free radical scavenging activity	Cucurbita pepo	Bharti <i>et al.</i> ¹²
Н	Peptidoglycans			
1	Fenugreekine	Glucose transport, carbohydrate digestion and absorption	Trigonella foenum graecum	Khosla <i>et al.</i> ²¹
2	Gluten, taraxacerin	Glucose transport, carbohydrate digestion and absorption	Taraxacum officinale	Hussain <i>et al.</i> , ⁵⁵ Yarnell and Abascal ⁹⁴
3	Glucosamines	Insulin secretion, carbohydrate digestion and absorption	Aloe vera	Ajabnoor ³⁸

(Continued)

Table 1. (Continued)

Sl. No.	Phytoconstituents	Targeted metabolic pathways	Plant source	References		
ı	Polyphenol and its derivatives					
1	Curcumin, turmerone, germacrone, zingiberene	Carbohydrate digestion and absorption, insulin secretion	Curcuma longa	Kar <i>et al.,</i> ²⁰ Zhang <i>et al.</i> ⁹⁵		
2	Ellagic acid and its derivatives	Carbohydrate digestion and absorption, insulin secretion	Potentilla candican, Phyllanthus niruri, Caesalpinia ferrea, Arctostaphylos uvaursi	Ueda <i>et al</i> . ⁹⁶		
3	Ellagic acid, corosolic acid, 4-hydroxybenzoic acid, 3-0-methylprotocatechuic acid, caffeic acid, p-coumaric acid, kaempferol	Carbohydrate digestion and absorption, insulin secretion	Lagerstroemia speciosa, Acacia Arabica	Naisheng <i>et al.</i> ⁹⁷		
4	Tannins, gallotannic acid	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Syzygium aromaticum	Hannan <i>et al</i> . ⁵⁴		
5	Wedelolactone, dimethyl wedelolactone	Insulin secretion, carbohydrate digestion and absorption	Eclipta alba	Ananthi <i>et al.</i> ⁹⁸		
6	Carvacrol, linalool	Insulin secretion, carbohydrate digestion and absorption	Ocimum sanctum	Hannan <i>et al.</i> , ⁵⁴ Broadhurst <i>et al</i> . ⁵⁶		
7	Mangiferin	$\alpha\text{-}Glucosidase\text{-}inhibiting$ activity	Salacia	Yoshikawa <i>et al.</i> ⁹⁹		
J	Saponins					
1	Stigmasterol, quercitol, gymnenic acid IV	Regeneration of pancreatic β cells, insulin secretion	Gymnema sylvestre	Sugihara <i>et al.</i> , ³⁴ Preuss <i>et al</i> . ³⁵		
2	Quinquenoside L3 and L9	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Panax quinquefolium	Vuksan <i>et al</i> . ⁶⁴		
3	Andrographolide	Regeneration of pancreatic β cells, insulin secretion	Andrographis paniculata	Yu et al. ¹⁰⁰		
4	3-O-β-D-glucopyranoside	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Myrtus communis	Alipour et al. 101		
5	3-Hepatadecanone, 8-hexadecenoic acid hexadecenoic acid	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Asparagus adscendens	Mathews <i>et al.</i> ¹⁰²		
6	Ginsenosides Rg2, panaxan A, B, C, D, E	Regeneration of pancreatic β cells, free radical scavenging	Panax ginseng	Ma <i>et al.</i> , ¹⁰³ Attele <i>et al</i> . ¹⁰⁴		
7	Lactucain C	Regeneration of pancreatic β cells, insulin secretion	Lactuca indica	Hou <i>et al.</i> ¹⁰⁵		
9	e-Glucoside, mangiferin, salacinol, kotalanol, epigallocatechin	Regeneration of pancreatic β cells, insulin secretion	Salacia reticulate, Salacia oblonga	Krishnakumar <i>et al</i> . ¹⁰⁶		
10	Allo-aromadendrene, T-cadinol, α-gurjunene, β-eudesmol, β-ubebene, aromadendrene	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Artemisia pallens	Ruikar <i>et al.</i> ¹⁰⁷		

Table 1. (Continued)

Sl. No.	Phytoconstituents	Targeted metabolic pathways	Plant source	References
11	Diosgenin	Glucose transport, carbohydrate metabolism	Trigonella foenum graecum	Khosla <i>et al.</i> ²¹
12	Sotolon [3.hydroxy-4,5- dimethyl-2(5H)-furanone], Trigonellin	Regeneration of pancreatic β cells, insulin secretion	Trigonella foenum- graecum	Khosla <i>et al.</i> ²¹
13	Ursolic acid, mulberrofuran-U	Regeneration of pancreatic $\boldsymbol{\beta}$ cells and insulin secretion	Morus insignis, Myrtus communis	Basnet <i>et al</i> . ¹⁰⁸
14	Kotalagenin-16-acetate, diterpene, triterpens	Carbohydrate digestion and absorption	Salacia oblongaq, Croton cajucara	Krishnakumar <i>et al.</i> , ¹⁰⁶ Silva <i>et al</i> . ¹⁰⁹
15	Muinol, azorellanol, mulin- 11,3-dien-20-oic-acid, mulinolic acid	Regeneration of pancreatic β cells and insulin secretion	Azorella compacta	Borquez <i>et al.</i> , ¹¹⁰ Fuentes <i>et al.</i> ¹¹¹
PPAR, peroxisome proliferator activated receptor.				

Catharanthine, vindoline and vindolinine, obtained from Catharanthus roseus lower the blood sugar level and show free radical scavenging action. 18,19 Glucose takes part in the glycation of the membrane lipid and its peroxidation to produce free radicals. In DM, the glucose concentration is very high and so is the amount of free radicals in the body which are highly reactive. To prevent their deleterious effect, our body has a defence system comprising several enzymes, which include superoxide dispiutase, catalase, reduced glutathione and glutathione-Stransferases. 113 Catharanthine, vindoline and vindolinine activate these free radical scavenging enzymes and prevent our body from their adverse effects. Vinblastine and vincristine are isolated from Vinca rosea, which also activate free radical scavenging enzymes.^{20,57} Sotolon [4,5-dimethyl-3-hydroxy-2(5H)-furanone], trigonelline, gentianine and carpaine compounds are extracted from Trigonella foenum graecum and downregulate the activity of fructose-1,6-bisphosphatase and check the dephosphorylation of fructose-1,6-bisphosphate.^{21,66,67,84} Ginkgolides found in the Ginkgo biloba plant have been reported to have an antihyperglycaemic effect on in vitro models.²² The mangiferin, a xanthone glucoside found in the leaves of Mangifera indica, has antidiabetic and antihyperlipidaemic properties. 114 The Allium sativum plant is a rich source of alkaloid allylpropyl disulfide that is involved in glycogen synthesis and insulin secretion.²³ Glycogen synthesis is a multistep process, but allylpropyl disulfide checks the conversion of pyruvate into lactate by

reducing the activity of the lactate dehydrogenase enzyme.

Aegelin, marmesin and marmelosin are the major alkaloids from the plant Aegle marmelos^{25,26} that causes regeneration of pancreatic β cells and insulin secretion. Insulin produced by the pancreatic β cells is one of the most important peptide hormones coordinating the utilization of fuels by tissues whose metabolic effects are anabolic, favoring, for example, the synthesis of glycogen, triacylglycerols and protein. The aim of this holistic approach by these botanicals is to repair pancreatic β cells and maintain the proper amount of insulin by increasing the expression of the insulin gene, increasing the secretion of insulin and inhibiting their degradation. Patients with T1DM have virtually no functional β cells (implicated due to genetic, autoimmune, environmental or viral factors) which leads to gradual depletion of the cellular population. They can neither respond to variations in circulating fuels nor maintain a basal secretion of insulin. Patients with T1DM must rely on exogenous insulin to control hyperglycaemia and ketoacidosis. The β carbolines (harmine, nor-harmine, pinoline) are believed to promote insulin secretion by β-cell regeneration and are the extracts of Tribulus terrestris. 27,28 Carbohydrate digestion and absorption is affected by betaine, achyranthine and β ecdysone isolated from Achyranthes aspera.²⁹ Castanospermine, epifagomine and fagomine are the chief phytoconstituents of Xanthocercis zambesiaca that are actively involved in carbohydrate digestion, absorption and insulin secretion.30

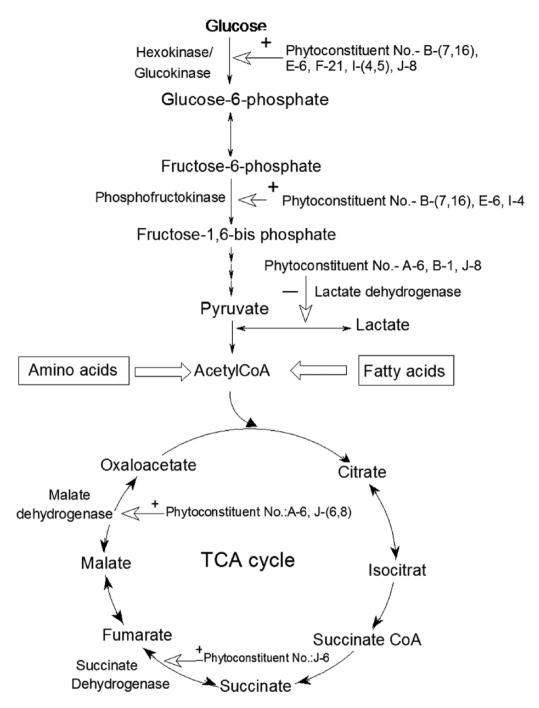


Figure 1. Phytoconstituent regulation of glycolysis and Krebs cycle with sources and fate of acetyl coenzyme A.

Berberine, found in the plant *Berberis aristata*, has been shown to have dipeptidyl peptidase IV (DPP-IV)-inhibiting activity. ¹⁷ The seed extract of *Castanospermum australe* contains three alkaloids, namely castanospermine, 7-deoxy-6-epi-castanospermine and australine, which have been shown to have DPP-IV inhibition activity and are effective in controlling the hyperglycaemic state in experimental rats. ^{11,31}

Amino acids, amines and carboxylic acid derivatives

The compounds allicin, apigenin and alliin extracted from *Allium sativum* target cholesterol and glycogen synthesis pathways.^{32,33} Cholesterol is the most abundant sterol in our body and is essential for normal functioning of the cells. If the cholesterol level exceeds the normal value, the chances of cardiovascular diseases increase. Cholesterol

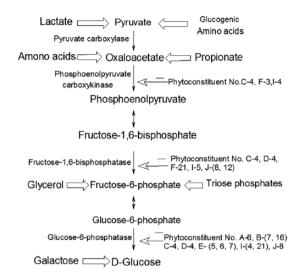


Figure 2. Illustration of the gluconeogenesis pathway with major substrate precursors and regulation by phytoconstituents.

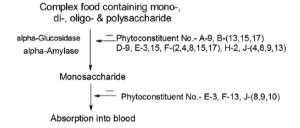


Figure 3. Regulation of carbohydrate metabolism by phytoconstituents.

synthesis is approximately a 30-step process with acetyl coenzyme A (CoA) as its precursor. Regeneration of pancreatic β cells and insulin secretion are activated by gurmarin, betaine, choline, gymnemic acid IV and trimethylamine isolated from *Gymnema sylvestre*. ^{32,34,35} (–) Hydroxycitric acid and 2-heptyl acetate, 2-methyl butyl acetate and isoamyl acetate are carboxylic acid derivatives from *Garcinia cambogi* and *Gymnema sylvestre* respectively that induce insulin secretion. ^{35,36} Erulic acid from *Curcuma longa* activates free radical scavenging activity and insulin secretion. ³⁷ *Aloe vera* extracts contain leucine, isoleucin and alanine, which trigger insulin secretion. ³⁸

Some plant extracts of *Caralluma edulis*, *Syzygium cumini* and *Acacia Arabica* contain malic acid and chiorogenic acid that check the steps of Krebs cycle.³⁹ The Krebs cycle is the central pathway for energy production in the mitochondrial matrix. Here pyruvate gets oxidized to CO₂ and H₂O *via*

acetyl CoA with the synthesis of energy equivalent Nicotinamide Adenine Dinucleotide (NADH), which ultimately is oxidized to produce energy via the electron transport chain. Out of seven enzymes involved in the cycle, only two, succinate dehydrogenase and malate synthase, are regulated by these botanicals. The compound polypeptide-P in Momordica charantia extract is shown to regulate insulin secretion and glycogen synthesis. 40,41 The compounds S-methyl cysteine sulfoxide and S-allyl cysteine sulfoxide derived from Allium cepa act on glycolysis and cholesterol synthesis. 24,42 Contrarily, the nitrosamines, nitrosated derivatives found in Areca catechu, are a great hyperglycaemia risk factor in the Asian population.⁴³ The compounds brevifolin carboxylic acid and ethyl brevifolin carboxylate extracted from Phyllanthus amarus are also involved in carbohydrate digestion and absorption.44 Viscum album extracts have been shown to have antihyperglycaemic effects and insulin-releasing effects in streptozotocin-induced diabetic rats and glucose-sensitive insulin-releasing pancreatic cell lines. 45-47 Coriander, a common household food ingredient used worldwide, has been shown to have significant insulin-like activity and helps in insulin secretion too.115 The compounds furfural and captylic acid from Agaricus campestris, 48,49 and bis-(2-ethyl) hexyl phthalate from Cassia auriculata, enhance insulin secretion and glycogen synthesis.⁵¹ The raisins from Vitis vinifera have insulin-mimetic activity.⁵² Procyanidins, extracted from grape seeds, have insulin-mimetic properties.⁵⁰

Anthranoids

Anthranoid compounds like aloin, barbaloin, isobarbaloine, aloetic acid, aloe-emodin, emodin, cinnamic acid and crysophanic acid from *Aloe vera and Cassia tora* initiate insulin secretion/synthesis. Momordica charantia is a rich source of vicine which acts on insulin secretion and glycogen synthesis. 40,41 Extracts from *Cassia tora* also stimulate insulin release. Compounds like camphor, eugenol, trans- β -ocimene, geraniol, α -pinene, limonene, p-cymene, 1,8-cineole and thujone, which help in pancreatic β -cell restoration and insulin secretion, are reported to be found in *Ocimum sanctum*, *Coriandrum sativum*, *Artemisia roxburghiana* and *Syzygium aromaticum*. $^{54-56}$

Carbohydrates

Plants like Aloe vera, Ocimum sanctum, Alpinia galangal, among others, contain polysaccharides

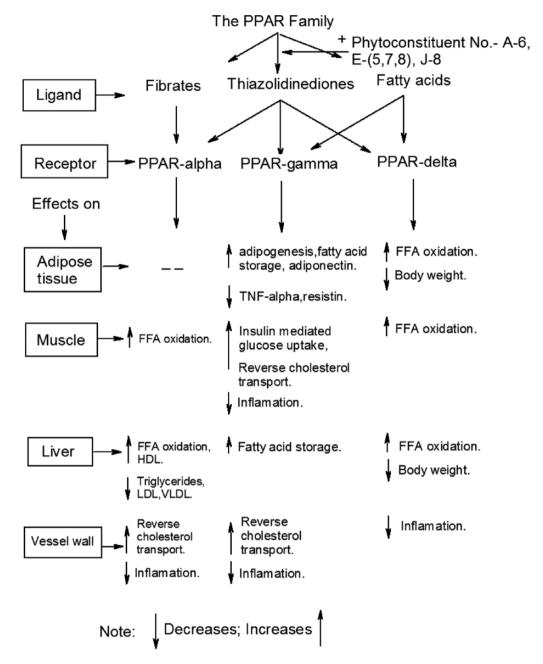


Figure 4. Mechanism of action of the peroxisome proliferator activated receptor (PPAR) family and their regulation by phytoconstituents.

The PPARs are ligand-activated nuclear receptors (α , δ and γ isoforms of PPAR) that can be activated by a range of fatty acids and derivatives, and they function as regulators in the biosynthesis, metabolism and storage of fats. PPAR ligands have displayed the importance of these receptors in the regulation of lipid and glucose homeostasis.

which have a considerable hypoglycaemic effect. Glucomannan, caryophylline, protein-bound polysaccharide, cellulose and mannose from these plants are either directly or indirectly involved in insulin secretion, carbohydrate digestion and absorption.⁵⁷ Guar gum, pectin and pectin fibres and mucilaginous fibre are secretory and excretory products of *Trigonella foenum graecum*, *Citrus*

sinensis and Coccinia indica that initiate insulin secretion, carbohydrate digestion and absorption. 20,58 Hericium erinaceus contain many β -glucan polysaccharides such as D-threitol and D-arabinitol which have antihyperglycaemic action. 59,60 Carbohydrate digestion and absorption are also regulated by L-arabino-D-xylan, cinnzeylanin, cinnzeylanol and D-glucan, which are

extracted from Cinnamomum zeylanicum blume.⁶¹ Opuntia ficus indica, Myrtus cummunis and Taraxacum officinale probably inhibit α-glucosidase, leading to slow absorption of carbohydrates.^{62,63} Fructo-oligosaccharide extract of plant and microbial origin significantly decreases glycosuria, advanced glycation end products (AGEs) and plasma triglycerides, as well as very low density lipoproteins.⁹

Glycosides

Gymnemic acid and gymnemosides from Gymnema sylvestre, 34,35 and astragalin, scopolin, skimmin and roscoside II from Morus alba65 are mainly involved in the restoration of pancreatic β cells and insulin secretion. Some major glycosides that control the process of insulin secretion and glycogen synthesis are vin α-ginsenoside R3 from Panax quinquefolium,64 momordin, momordicine, charantin, momorcharaside A and B, and momorcharin A and B from Momordica charantia, 40,41 cucurbitacin B and isocucurbitacin B from Helicteres isora, 69 momordina and luffina from Luffa cylindnica,69 kotalanol and salacinol from Salacia reticulate and Salacia oblonga,70 arbutin and eriolin from Arctostaphylos uvaursi, 71 citrullol, colocynthin, elaterin, elatericin B and colosynthetin from Cifrullus colocynthis,72,73 leucopelargonidin, leucocyanidin and pelarogonidin from Ficus bengalensis,74-76 and taraxacin from Taraxacum officinale.56 Tinospora cordifolli and T. crispa are the major sources of tinosporine, cordifolide, tinosporide, cordifole and columbin that regulate cholesterol synthesis and glycolysis. 6,20,57,68 Glucose transport and carbohydrate metabolism are the targeted pathways of C-glycosides which are extracted from the plant Trigonella foenum graecum, 66,67

Flavonoids

Flavonoids are poly-hydroxy poly-phenolic compounds which have a wide ranging herbal presence. Flavonoids are classified into categories like flavanols, flavones and flavanones, and have numerous medicinal effects including antidiabetic properties. Chrysin and isoquercitrin isolated from *Morus alba* are involved in insulin secretion. Free radical scavenging and insulinonematic activity have been shown by epigallocatechin gallate (EGCG), epigallocatechin (EGC), epicatechin, catechin and quercetin extracted from *Camellia sinensis*, *Punica granatum*, *Satureja khuzestanica* and *Bauhinia forficate*. 77–80

Myrcia multiflora and Abelmoschus moschatus are important sources of myrciaphenones A and B, and myrciacitrins I and II.18,81 Citrus bioflavonoids (hesperidin and naringin) are extracts of Camellia sinensis which target glycogen synthesis, glycolysis and gluconeogenesis.82 Flavanoids such as quercetin, quercetrin, apigenin, rutin, apigenin-7-O-glucoside and naringenin are important phytoconstituents of Panax notoginseng, 83,64 Urtica dioica, Bauhinia varigtla^{55,84} and Camellia sinensis78 which are actively involved in the restoration of pancreatic β-cell and insulin secretion. Sov isoflavones (genistein and diadzein) are major chemical constituents of Glycin max and Curcuma longa and are involved in lipid and glucose metabolism by activation of peroxisome proliferator activated receptors (PPARs).85,86 The PPARs bind DNA as heterodimers with the retinoid X receptors to the peroxisome proliferator response elements identified in the promoter region of a number of genes involved in lipid and carbohydrate metabolism. 11,12 The three human isoforms of PPAR, α , δ and γ , show distinct patterns of tissue distribution and ligand preference, and control different biological activities (Figure 4). PPARα is a regulator of fatty acid catabolism and peroxisome proliferation in the liver, while PPARy plays a key role in adipogenesis. All three isoforms are expressed in macrophages where they are implicated in the control of cholesterol efflux. The use of synthetic PPAR ligands has demonstrated the importance of these receptors in the regulation of lipid and glucose homeostasis and today PPARs are established molecular targets for the treatment of T2DM and cardiovascular disease. 11,12,116 Phytoconstituents like aegelin, marmesin, marmelosin, momordin, momordicine, charantin, momorcharaside A and B, momorcharin A and B, cucurbitacin B, isocucurbitacin B and β-sitosterol (Table 1) increase the expression of PPARy and decrease insulin resistance. The thiazolidinediones class of drugs (troglitazone, pioglitazone, etc.) is known to consist of activators of PPARy, which are used pharmacologically as insulin sensitizers. With the growing understanding of PPAR biology, it has become evident that novel herbal drugs modulating PPAR activity could improve current diabetes treatment. 11,12,116

Bauhinia candicans and Bauhinia forficata produce kaempferitrin that affects glycolysis. ^{69,87} Proanthocyanidins, α-terpineol and hexanol obtained from *Vitis vinifera* and *Agaricus campes*tris have insulinomimetic activity. ^{49,50,52} The

compounds catechin, epicatechin, chiorogenic acid, liquiritigenin and isoliquiritigerin have been extracted from a number of plants, namely, Phylanthus embelica, Acacia Arabica, Pterocarpus marsupium and Phylanthus embelica. 7,20,39,57 Insulinomimetic activity was also shown by the potential applications of silvmarin, silvbin, silvchristin and silidianin (extracts of Silvbum marianum) along with 3-hydroxy-3-methylglutaryl Coenzyme A (HMG CoA) suppression activity.88 Insulin secretion and glycogen synthesis are also targeted by amarogentin, swerchirin, chirantin and gentiopicrin (extracts of Swertia chirayita),57 shamimin (extracts of Biophytum sensitivum),89,90 leucopelargonidin and dulcitol (extracts of Casearia esculenta), 91 isorhamnetin, quercetin and kaempferol (extracts of Matteuccia orientalis) and anthocyanosides (bioflavonoids found in bilberry).92 Among all the reported flavonoids in Table 1, some have potential antidiabetic effects, like quercetin, naringenin, chrysin,²⁰ citrus bioflavonoids like hesperidin and naringin, 117 anthocyanidins, 92 soy isoflavones genistein or daidzein,86 kaempferitrin [kaempferol-3,7-O-(alpha)-l-dirhamnoside],69,87 tea flavonoid, EGCG and epicatechin.78

Minerals and vitamins

Zinc has been shown to be associated with proper functioning of pancreatic β cells and maturation of insulin secretory granules. A high serum level of zinc has been related to improved insulin sensitivity. The antioxidant property of zinc has been related to the prevention of oxidative stress.⁹³ It has been shown that oxidative stress plays an important role in DM and reactive oxygen/nitrogen species (ROS/RNS: superoxides, hydrogen peroxide, hydroxyl anions, singlet oxygen and nitric oxide) are believed to be important independent risk factors that are developed in DM and known as autooxidative glycosylation (a process which is relevant at elevated blood glucose level).118 Once they have formed, they react with cellular components such as DNA, or the cell membrane and cellular damage starts due to a chain reaction. Cells may function poorly or die if this occurs. Many phytoconstituents have antioxidant properties that inhibit the formation of free radicals and lipid peroxidation or neutralize them in cells to prevent the propagation reaction from continuing. Tocopherol and carotenoids, the two common natural vitamins, from the seeds of Cucurbita pepo (pumpkin) have been shown to have antidiabetic effects on experimental diabetic

rats. ¹² Vitamin D has a strong relation with pathogenesis of T2DM. Vitamin D level and β-cell functioning are positively correlated. Vitamin D deficiency leads to T2DM and people with T2DM are also prone to vitamin D deficiency and related pathologies. ^{119,120} Vitamin D supplementation improves fasting plasma glucose and insulin level. ¹²¹

Peptidoglycans

A few phytoconstituents of this category like fenugreekine (extract of *Trigonella foenum graecum*),²¹ inulin, taraxacosides (extract of *Taraxacum officinale*)^{55,94} and glucosamines (extract of *Aloe vera*)³⁸ are efficiently involved in glucose transport, carbohydrate digestion and absorption.

Polyphenol and its derivatives

Polyphenolic phytochemicals are ubiquitous in plants, in which they function in various protective roles. It is suggested that polyphenols, and particularly curcuminoids might be of value as a complement to pharmaceutical treatment, but also prebiotic treatment, in conditions proven to be rather therapy resistant, such as Crohn's disease, long-stay patients in intensive care units, but also for conditions such as cancer, liver cirrhosis, chronic renal disease, chronic obstructive lung disease, diabetes and Alzheimer's disease. Curcuma longa is the chief source of curcumin, turmerone, germacrone and zingiberene which improve glucose metabolism.^{20,95} There are so many plants such as Potentilla candican, Phyllanthus niruri, Caesalpinia ferrea and Arctostaphylos uvaursi which produce ellagic acid, helpful in carbohydrate digestion and absorption, and insulin secretion.⁹⁶ A number of phytoconstituents like corosolic acid, 4-hydroxybenzoic acid, 3-O-methylprotocatechuic acid, caffeic acid, p-coumaric acid and kaempferol are actively involved in carbohydrate digestion and absorption, and insulin secretion. These phytoconstituents are extracted from Lagerstroemia speciosa and Acacia arabica. 97 Compounds like wedelolactone and dimethyl wedelolactone, extracted from Eclipta alba, are involved in insulin secretion and carbohydrate digestion.98 Phytoconstituents of Ocimum sanctum (carvacrol, linalool) regulate insulin secretion, carbohydrate digestion and absorption.^{54,56} Mangiferin extracted from Salacia species has α-glucosidase-inhibiting activity, making it an effective antihyperglycaemic agent.99

Saponins

Saponins are bioactive compounds present naturally in many plants and known to possess potent antihyperglycaemic activity.26 Stigmasterol, quercitol, gymnenic acid IV (extract of Gymnema sylvestre),34,35 quinquenoside L3 and L9 (extract of Panax quinquefolium),64 andrographolide (extract of Andrographis paniculata), 100 myrtucommulone and limonene (extract of Myrtus communis), 101 3-hepatadecanone and 8-hexadecenoic acid (extract of Asparagus adscendens)102 and ginsenosides Rg2 and panaxan A, B, C, D, E (extract of Panax quinquefolium)^{103,104} are efficiently involved in the restoration of pancreatic β -cell and insulin secretion. Lactucain C obtained from Lactuca indica was found to produce significant antihyperglycaemic activity. 105 Salacinol, kotalanol and EGC obtained from Salacia reticulate and Salacia oblonga were found to possess significant antihyperglycaemic activity. 70,106 Several polyphenols obtained from the plant of Artemisia pallens exhibit potent antioxidant and hypoglycaemic activity. 107,122 Diosgenin from Trigonella foenum graecum regulates glucose transport and carbohydrate metabolism but the exact action mechanism of the constituents is not properly understood.²¹ [3-hydroxy-4,5-dimethyl-2(5H)-furanonel and trigonellin are other compounds of Trigonella foenum graecum which restore pancreatic β cells for proper insulin secretion.²¹ Ursolic acid and mulberrofuran-U of Morus insignis have antihyperglycaemic activity in both types of diabetes.¹⁰⁸ Kotalagenin-16-acetate, diterpene and triterpens are a few saponins extracted from the plants Salacia oblongaq and Croton cajucara. 106,109 They are either directly or indirectly involved in carbohydrate digestion and absorption. Extract of Azorella compacta contains muinol, azorellanol, mulin-11, 3-dien-20-oic-acid and mulinolic acid, 110,111 which restore pancreatic β cells and increase insulin secretion.

Conclusion

Diabetes is a disorder of carbohydrate, fat and protein metabolism attributed to the diminished production of insulin or mounting resistance to its action. In spite of all the advances in therapeutics, diabetes still remains a major cause of morbidity and mortality in the world. The most commonly used drugs of modern medicine such as aspirin, antimalarials, anticancers, digitalis, among others, originated from plant sources. Considering the safety, efficacy and time tested utility in humans under different traditional

systems of medicines, plant sources are regarded as safe. Thus, plants offer a natural alternative or an adjunct to conventional agents with fewer side effects. However, for concrete evidence and application as drugs in the stricter norms of drug development, more studies are required to evaluate their activities and associated benefits in the prevention or treatment of diabetes in humans. Several plant-derived drugs have been scientifically validated as potent antidiabetics and include flavonoids (queretin, neringerin and chrysin), alkaloids (berberin, catharenthine and vindolin), glycosides and saponins (triterpenoid and steroidal glycosides such as charantin, lactucain C, β-sitosterol and gymnemic acid), glycolipids, dietary fibres, imidazole compounds, polysaccharides, peptidoglycans, carbohydrates and amino acids. Among these, the alkaloids, flavonoids and saponins show diverse effects. Most of the plants having antihyperglycaemic activity also show other functions that are beneficial to patients with DM. Taken together, the data on botanical compounds compiled in this review provide a lead with respect to diabetes management, showing the regulatory effects on various steps of different metabolic pathways that may have therapeutic and other applications. Although recent progress has been made in understanding the underlying mechanisms and diverse activities of these plantderived drugs, further studies are required to firmly establish the mechanisms of actions.

Future prospects

Using biotechnological tools, the future would be better equipped to offer personalized approaches to preventive diabetology. Advances in plant genomics would facilitate individualized diets customized to a person's genetic profile to maximize health and wellbeing. A futuristic doctor's desk reference would contain information on individual genetic profiles to be matched specific phytochemical interventions. Simultaneously, toxicity to specific ingredients would be minimal, as recommendations would be based on an individual's genetic profiles and susceptibility data. Armed with a cornucopia of phytoconstituents, and a dazzling array of genomic evidence, preventive diabetology is all set to trace the footprints of ancient wisdom. Also, these drugs are absolutely natural and very economical, which could make them applicable for the masses at large. In addition, many herbal remedies used today have not undergone careful scientific assessment and some have the potential

to cause serious toxic effects and major drugdrug interactions.

Acknowledgements

The authors are grateful to the Department of Biotechnology, National Institute of Technology, Raipur, India, and the Department of Biochemistry, Patna University, Patna, India for providing facilities, space and resources.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

Conflict of interest statement

The authors declare that there is no conflict of interest.

ORCID iD

Awanish Kumar https://orcid.org/0000-0001-8735-479X

References

- Unnikrishnan R, Anjana RM and Mohan V. Diabetes mellitus and its complications in India. Nat Rev Endocrinol 2016; 12: 357–370.
- International Diabetes Federation. IDF diabetes atlas. 7th ed. Brussels: International Diabetes Federation, 2015.
- 3. Kumar A, Bharti SK and Kumar A. Therapeutic molecules against type 2 diabetes: what we have and what are we expecting? *Pharmacol Rep* 2017; 69: 959–970.
- 4. Kumar A, Bharti SK and Kumar A. Type 2 diabetes mellitus: the concerned complications and target organs. *Apollo Med* 2014; 11: 161–166.
- Bharti SK, Krishnan S and Gupta AK. Herbal formulation to combat type 2 diabetes mellitus. Germany: LAMBERT Academic Publishing, 2013.
- 6. Hui H, Tang G and Go VL. Hypoglycemic herbs and their action mechanisms. *Chin Med* 2009; 12: 4–11.
- Grover JK and Vats V. Shifting paradigm from conventional to alternate medicine: an introduction on traditional Indian medicine. *Asia Pac Biotech News* 2000; 5: 28–32.
- 8. Ruderman MB, Tornheim K and Goodman MN. Fuel homeostasis and intermediary metabolism of carbohydrate, fat and protein. In: Becker KL, Bilezikian JP, Bremner WJ, et al. (eds) *Principles and practice of endocrinology and*

- *metabolism*. Philadelphia: JB Lippincott, 2001, pp.1257–1271.
- Bharti SK, Krishnan S, Kumar A, et al.
 Antidiabetic activity and molecular docking of fructooligosaccharides produced by Aureobasidium pullulans in poloxamer-407-induced T2DM rats. Food Chem 2013; 136: 813–821.
- Park K. Park's textbook of preventive and social medicine. 22th ed. Jabalpur: M/S Banarasidas Bhanot, 2013, pp.302–309.
- Bharti SK, Krishnan S, Kumar A, et al.
 Antihyperglycemic activity with DPP-IV inhibition of alkaloids from seed extract of Castanospermum australe: investigation by experimental validation and molecular docking. Phytomedicine 2012; 20: 24–31.
- 12. Bharti SK, Kumar A, Sharma NK, *et al.*Tocopherol from seeds of *Cucurbita pepo* against diabetes: validation by in vivo experiments supported by computational docking. *J Formos Med Assoc* 2013; 112: 676–690.
- 13. Alarcon-Aguilara FJ, Roman-Ramos R, Perez-Gutierrez S, et al. Study of the antihyperglycemic effect of plants used as antidiabetics. J Ethnopharmacol 1998; 61: 101–110.
- Luo J, Fort DM, Carlson TJ, et al. Cryptolepis sanguinolenta: an ethnobotanical approach to drug discovery and the isolation of a potentially useful new antihyperglycaemic agent. Diabet Med 1998; 15: 367–374.
- Bharti SK, Krishnan S and Kumar A. Phytotherapy for diabetes mellitus: back to nature. *Minerva Endocrinol* 2016; 41: 143–146.
- Singh SS, Pandey SC, Srivastava S, et al.
 Chemistry and medicinal properties of *Tinospora cordifolia* (Guduchi). *Indian J Pharmacol* 2003;
 35: 83–91.
- 17. Al masri IM, Mohammad MK and Tahaa MO. Inhibition of dipeptidyl peptidase IV (DPP-IV) is one of the mechanisms explaining the hypoglycemic effect of berberine. J Enzyme Inhib Med Chem 2009; 24: 1061–1066.
- 18. Chattopadhyay RR. A comparative evaluation of some blood sugar lowering agents of plant origin. *J Ethnopharmacol* 1999; 67: 367–372.
- 19. Jarald EE, Sheeja E, Motwani S, et al.
 Comparative evaluation of antihyperglycaemic and hypoglycaemic activity of various parts of Catharanthus roseus Linn. Res J Med Plant 2008; 2: 10–15.
- 20. Kar A, Choudhary BK and Bandyopadhyay NG. Comparative evaluation of hypoglycaemie

- activity of some Indian medicinal plants in alloxan diabetic rats. *J Ethnopharmacol* 2003; 84: 105–108.
- Khosla P, Gupta DD and Nagpal RK. Effect of *Trigonella foenum graecum* (Fenugreek) on serum lipids in normal and diabetic rats. *Indian J Physiol Pharmacol* 1995; 27: 89–93.
- 22. Pinto MDS, Kwon YI, Apostolidis E, *et al*. Potential of *Ginkgo biloba* L. leaves in the management of hyperglycemia and hypertension using in vitro models. *Bioresour Technol* 2009; 100: 6599–6609.
- Sheela CG, Kumud K and Augusti KT. Antidiabetic effects of onion and garlic sulfoxide amino acids in rats. *Planta Med* 1995; 61: 356–357.
- 24. Kumari K and Augusti KT. Lipid lowering effect of S-methyl cysteine sulfoxide from *Allium cepa* Linn in high cholesterol diet fed rats. *J Ethnopharmacol* 2007; 109: 367–371.
- 25. Kamalakkannan N and Prince PSM. The effect of *Aegle marmelos* fruit extract in streptozotocin diabetes: a histopathological study. *J Herb Pharmacol* 2005; 5: 87–96.
- 26. Ponnachan PT, Paulose CS and Panikkar KR. Effect of leaf extract of *Aegle marmelose* in diabetic rats. *Indian J Exp Biol* 1993; 31: 345–347.
- 27. Cooper EJ, Hudson AL, Parker CA, et al. Effects of the beta-carbolines, harmane and pinoline, on insulin secretion from isolated human islets of Langerhans. Eur J Pharmacol 2003; 482: 189–196.
- 28. Kirtikar KR and Basu BD. *Indian medicinal plants*, vol. 1. 1998.
- 29. Akhtar MS and Iqbal J. Evaluation of the hypoglycaemic effect of *Achyranihes aspera* in normal and alloxan-diabetic rabbits. *J Ethnopharmacol* 1991; 31: 49–57.
- Akhtar MS. Hypoglycaemic activities of some indigenous medicinal plants traditionally used as antidiabetic drugs. J Pak Med Assoc 1992; 42: 271–277.
- 31. Orwa C, Mutua A, Kindt R, et al. Agroforestree database: a tree reference and selection guide version 4.0. Kenya: World Agroforestry Centre, 2009.
- 32. Gholap S and Kar A. Hypoglycaemic effects of some plant extracts are possibly mediated through inhibition in corticosteroid concentration. *Pharmazie* 2004; 59: 876–878.
- 33. Kumar GR and Reddy KP. Reduced nociceptive responses in mice with alloxan

- induced hyperglycemia after garlic treatment. *Indian 7 Exp Biol* 1999; 37: 662–666.
- 34. Sugihara Y, Nojima H, Matsuda H, et al. Antihyperglycemic effects of gymnemic acid IV, a compound derived from *Gymnema sylvestre* leaves in streptozotocin-diabetic mice. *J Asian Nat Prod Res* 2000; 2: 321–327.
- 35. Preuss HG, Bagchi D, Bagchi M, et al. Effects of a natural extract of (-)-hydroxycitric acid (HCA-SX) and a combination of HCA-SX plus niacin-bound chromium and *Gymnema sylvestre* extract on weight loss. *Diabetes Obes Metab* 2004; 6: 171–180.
- Hayamizu K. Effect of *Garcinia cambogia* extract on serum leptin and insulin in mice. *Fitoterapia* 2003; 74: 267–273.
- Ohnishi M, Matuo T, Tsuno T, et al.
 Antioxidant activity and hypoglycemic effect of ferulic acid in STZ-induced diabetic mice and KK-Ay mice. Biofactors 2004; 21: 315–319.
- 38. Ajabnoor MA. Effect of aloes on blood glucose levels in normal and alloxan diabetic mice. *J Ethnopharmacol* 1990; 28: 215–220.
- 39. Wadood AN and Shah SA. Effects of *Acacia arabica* and *Caralluma edulis* on blood glucose levels of normal and alloxan diabetic rabbits. *J Pak Med Assoc* 1989; 39: 208–212.
- 40. Chao CY and Huang CJ. Bitter gourd (Momordica charantia) extract activates peroxisome proliferator-activated receptors and upregulates the expression of the acyl CoA oxidase gene in H4IIEC3 hepatoma cells. J Biomed Sci 2003; 10: 782–791.
- 41. Sarkar S, Pranava M and Marita R.
 Demonstration of the hypoglycemic action
 of *Momordica charantia* in a validated animal
 model of diabetes. *Pharmacol Res* 1996; 33: 1–4.
- 42. Roman-Ramos R, Flores-Saenz JL and Alarcon-Aguilar FL. Anti-hyperglycemic effect of some edible plants. *J Ethnopharmacol* 1995; 48: 25–32.
- 43. Mannan N, Boucher BJ and Evans SJW. Increased waist size and weight in relation to consumption of *Areca catechu* (betel-nut); a risk factor for increased glycaemia in Asians in East London. *Br J Nutr* 2000; 83: 267–275.
- 44. Ali H, Houghton PJ and Soumyanath A. α-Amylase inhibitory activity of some Malaysian plants used to treat diabetes; with particular reference to *Phyllanthus amarus*. J Ethnopharmacol 2006; 107: 449–455.
- 45. Adaramoye O, Amanlou M, Habibi-Rezaei M, *et al.* Methanolic extract of African mistletoe (*Viscum album*) improves

- carbohydrate metabolism and hyperlipidemia in streptozotocin-induced diabetic rats. *Asian Pac* 7 *Trop Med* 2012; 5: 427–433.
- 46. Eno AE, Ofem OE, Nku CO, et al. Stimulation of insulin secretion by Viscum album (mistletoe) leaf extract in streptozotocin-induced diabetic rats. Afr J Med Med Sci 2008; 37: 141–147.
- 47. Gray AM and Flatt PR. Insulin-secreting activity of the traditional antidiabetic plant *Viscum album* (mistletoe). *J Endocrinol* 1999; 160(3): 409–414.
- 48. Manohar V, Talpur NA, Echard BW, et al. Effects of a water-soluble extract of maitake mushroom on circulating glucose/insulin concentrations in KK mice. *Diabetes Obes Metab* 2002; 4: 43–48.
- 49. Gray AM and Flatt PR. Insulin-releasing and insulin-like activity of *Agaricus campestris* (mushroom). *J Endocrinol* 1998; 157: 259–266.
- 50. Pinent M, Blay M, Blade MC, *et al.* Grape seed-derived procyanidins have an antihyperglycemic effect in streptozotocininduced diabetic rats and insulinomimetic activity in insulin-sensitive cell lines. *Endocrinology* 2004; 145: 4985–4990.
- 51. Abesundara KJM, Mastui T and Matsumoto K. α-Glucoidase inhibitory activity of some Sri Lanka plant extracts, one of which, *Cassia auriculata*, exerts a strong antihyperglycemic effect in rats comparable to the therapeutic drug acarbose. *J Agric Food Chem* 2004; 52: 2541–2545.
- 52. Rankin JW, Andreae MC, Oliver Chen CY, et al. Effect of raisin consumption on oxidative stress and inflammation in obesity. *Diabetes Obes Metab* 2008; 10: 86–96.
- 53. Nam J and Choi H. Effect of butanol fraction from *Cassia tora* L. seeds on glycemic control and insulin secretion in diabetic rats. *Nutr Res Pract* 2008; 2: 240–246.
- Hannan JM, Marcnah L, Au L, et al. Ocimum sanctum leaf extracts stimulate insulin secretion from perfused pancreas, isolated islets and clonal pancreatic β-cells. J Endocrinol 2006; 189: 127–136.
- 55. Hussain Z, Waheed A, Qurshi RA, et al. The effect of medicinal plants of Islamabad and Murree region of Pakistan on insulin secretion from INS-1 cells. Phytother Res 2004; 18: 73–77.
- Broadhurst CL, Polansky MM and Anderson RA. Insulin-like biological activity of culinary and medicinal plant aqueous extracts in-vitro. J Agric Food Chem 2000; 48: 849–852.
- 57. Van de Venter M, Roux S, Bungu LC, *et al.* Antidiabetic screening and scoring of 11

- plants traditionally used in South Africa. *J Ethnopharmacol* 2008; 119: 81–86.
- 58. Nandini CD, Sambaiah K and Salimath PV. Dietary fibres ameliorate decreased synthesis of heparan sulphate in streptozotocin induced diabetic rats. *J Nutr Biochem* 2003; 14: 203–210.
- Khan MA, Tania M, Liu R, et al. Hericium erinaceus: an edible mushroom with medicinal values. J Complement Integr Med 2013; 10(1): 1–6.
- 60. Liang B, Guo Z, Xie F, et al. Antihyperglycemic and antihyperlipidemic activities of aqueous extract of *Hericium erinaceus* in experimental diabetic rats. *BMC Complement Altern Med* 2013; 13: 253.
- Solomon TP and Blannin AK. Effects of shortterm cinnamon ingestion on in vivo glucose tolerance. *Diabetes Obes Metab* 2007; 9: 895–901.
- 62. Godard MP, Ewing BA, Pischel I, *et al.* Acute blood glucose lowering effects and long-term safety of OpunDiaTM supplementation in prediabetic males and females. *J Ethnopharmacol* 2010; 130: 631–634.
- 63. Onal S, Timur S, Okutucu B, *et al.* Inhibition of alpha-glucosidase by aqueous extracts of some potent antidiabetic medicinal herbs. *Prep Biochem Biotechnol* 2005; 35: 29–36.
- 64. Vuksan V, Sievenpiper JL, Koo VY, et al. American ginseng (*Panax quinquefolius* L) reduces postprandial glycemia in nondiabetic subjects and subjects with type 2 diabetes mellitus. *Arch Intern Med* 2000; 160: 1009–1013.
- 65. Gulubova R and Boiadzhiev TS. Morphological changes in the endocrine pancreas of the rabbit after the administration of a *Morus alba* extract. *Eksp Med Morfol* 1975; 14: 166–171.
- 66. Gupta D, Raju J and Baquer NZ. Modulation of some gluconeogenic enzyme activities in diabetic rat liver and kidney: effect of antidiabetic compounds. *Indian J Exp Biol* 1999; 37: 196–199.
- 67. Kluwer WC. The review of natural products by facts and comparisons. St Louis, MO: Facts & Comparisons®, 1999.
- 68. Noor H and Ashcroft SJH. Insulinotropic activity of *Tinospora crispa* extract: effect on β-cell Ca²+ handling. *Phytother Res* 1998; 12: 98–102.
- Lemus I, Garcia R, Dclvillar E, et al.
 Hypoglycemic activity of four plants used in Chilean popular medicine. Phytother Res 1999; 13: 91–94.
- Huang TH, He L, Qin Q, et al. Salacia oblonga root decreases cardiac hypertrophy in Zucker diabetic fatty rats: inhibition of cardiac expression of angiotensin II type 1 receptor. Diabetes Obes Metab 2008; 10: 574–585.

- Moon YH, Nam SH, Kang J, et al. Enzymatic synthesis and characterization of arbutin glucosides using glucansucrase from Leuconostoc mesenteroides. Appl Microbiol Biotechnol 2007; 27: 559–567.
- 72. González-Tejero MR, Casares-Porcel M, Sánchez-Rojas CP, *et al.* Medicinal plants in the Mediterranean area: synthesis of the results of the project Rubia. *J Ethnopharmacol* 2008; 116: 341–357.
- Ziyyat A, Lcgssyer A, Mekhfi HR, et al. Phytotherapy of hypertension and diabetes in oriental Morocco. J Ethnopharmacol 1997; 58: 45–54.
- 74. Singh RK, Mehta S, Jaiswal D, et al.
 Antidiabetic effect of *Ficus bengalensis* aerial roots in experimental animals. *J Ethnopharmacol* 2009; 123: 110–114.
- 75. Cherian S, Sheela CG and Augusti KT. Insulin sparing action of leucopelergonidin derivative isolated from *Ficus bengalensis* Linn. *Indian J Exp Biol* 1995; 33: 608–611.
- Kumar RV and Augusti KT. Insulin sparing action of leucocyanidin derivative isolated from Ficus bengalensis Linn. Indian J Biochem Biophys 1994; 31: 73–76.
- 77. Hii SCT and Howell SL. Effects of epicatechin on rat islets of Langerhans. *Diabetes* 1984; 33: 291–296.
- 78. Waltner-Law ME, Wang XL, Law BK, *et al.* Epigallocatechin gallate: a constituent of green tea, represses hepatic glucose production. *J Biol Chem* 2002; 277: 34933–34940.
- 79. Vessal M, Hemmati M and Vasei M. Hypoglycemic effects of quercetin in streptozocin-induced diabetic rats: comparative biochemistry and physiology. *Toxicol Pharmacol* 2003; 135: 357–364.
- 80. Li Y, Qi Y, Huang TH, et al. Pomegranate flower: a unique traditional antidiabetic medicine with dual PPAR-alpha/-gamma activator properties. *Diabetes Obes Metab* 2008; 10: 10–17.
- 81. Ngueyem TA, Brusotti G, Caccialanza G, *et al.* The genus Bridelia: a phytochemical and ethnopharmacological review. *J Ethnopharmacol* 2009; 124: 339–349.
- 82. Jung UJ, Lee MK, Jeong KS, *et al.* The hypoglycemic effects of hesperidin and naringin are partly mediated by hepatic glucoseregulating enzymes in C57BL/KsJ-db/db mice. *J Nutr* 2004; 134: 2499–2503.
- 83. Liu KZ, Li JB, Lu HL, et al. Effects of Asiragalus and saponins of *Panax notoginseng* on MMP-9 in patients with type 2 diabetic. *Macroangiopathy* 2004; 29: 264–266.

- 84. Jellin JM, Batz F and Hitchens K. *Pharmacist's letter/prescriber's letter natural medicines comprehensive database*. Stockton, CA: Therapeutic Research Faculty, 1999.
- 85. Howes JB, Tran D, Brillante D, et al. Effects of dietary supplementation with isoflavones from red clover on ambulatory blood pressure and endothelial function in postmenopausal type 2 diabetes. Diabetes Obes Metab 2003; 5: 325–332.
- 86. Mezei O, Banz WJ, Steger RW, et al.
 Soy isoflavones exert hypoglycemic and
 hypolipidemic effects through the PPAR
 pathways in obese Zucker rats and murine RAW
 264.7 cells. J Nutr 2003; 133: 1238–1243.
- 87. Jorge AP, Horst H, de Sousa E, *et al*. Insulinomimetic effects of kaempferitrin on glycaemia and on glucose uptake in rat soleus muscle. *Chem Biol Interact* 2004; 149: 89–96.
- 88. Huseini HF, Larijani B, Heshmat R, et al.

 The efficacy of Silybum marianum (L.) Gaertn
 (Silymarin) in the treatment of type 2 diabetes:
 a randomized, double-blind, placebo-controlled
 clinical trial. Phytother Res 2006; 20: 1036–1039.
- Puri D and Baral N. Hypoglycemic effect of Biophytum sensitivum in the alloxan diabetic rabbits. Indian J Physiol Pharmacol 1998; 42: 401–406.
- 90. Puri D. The insulinotropic activity of a Nepalese medicinal plant *Biophytum* sensitivum: preliminary experimental study. *J* Ethnopharmacol 2001: 78: 89–93.
- 91. Prakasam A, Sethupathy S and Pugalendia KV. Antiperoxidative and antioxidant effects of *Casearia esculenta* root extract in streptozotocin induced diabetic rats. *Yale J Biol Med* 2005; 78: 15–23.
- Shane-McWhorter L. Biological complementary therapies: a focus on botanical products in diabetes. *Diabetes Spectr* 2001; 14: 199–208.
- 93. Wijesekara N, Chimienti F and Wheeler MB. Zinc, a regulator of islet function and glucose homeostasis. *Diabetes Obes Metab* 2009; 11(Suppl. 4): 202–214.
- 94. Yarnell E and Abascal K. Dandelion (*Taraxacum officinale* and *T. mongolicum*). *Integ Med* 2009; 8: 35–38.
- 95. Zhang D, Fu M, Gao SH, et al. Curcumin and diabetes: a systematic review. Evid Based Complement Alternat Med 2013; 2013: 636053.
- 96. Ueda H, Kawanishi K and Moriyasu M. Effects of ellagic acid and 2-(2,3,6-trihydroxy-4-carboxyphenyl) ellagic acid on sorbitol accumulation *in vitro* and *in vivo*. *Biol Pharm Bull* 2004; 27: 1384–1387.

- 97. Naisheng B, Kan H, Roller M, et al. Active compounds from Lagerstroemia speciosa, insulinlike glucose uptake-stimulatory/inhibitory and adipocyte differentiation-inhibitory activities in 3T3-L1 cells. J Agric Food Chem 2008; 56: 11668–11674.
- 98. Ananthi J, Prakasam A and Pugalendi KV. Antihyperglycemic activity of *Eclipta alba* leaf on alloxan-induced diabetic rats. *Yale J Biol Med* 2003; 76: 97–102.
- 99. Yoshikawa M, Nishida N, Shimoda H, et al. Polyphenol constituents from Salacia species: quantitative analysis of mangiferin with alphaglucosidase and aldose reductase inhibitory activities. Yakugaku Zasshi 2001; 121: 371–378.
- 100. Yu BC, Hung CR, Chen WC, *et al.*Antihyperglycemic effect of andrographolide in streptozotocin induced diabetic rats. *Planta Med* 2003; 69: 1075–1079.
- Alipour G, Dashti S and Hosseinzadeh H. Review of pharmacological effects of *Myrtus communis* L. and its active constituents. *Phytother Res* 2014; 28: 1125–1136.
- 102. Mathews JN, Flatt PR and Abdel-Wahab YH. *Asparagus adseendens* (Shweta musali) stimulates insulin secretion, insulin action and inhibits starch digestion. *Br J Nutr* 2006; 95: 576–581.
- 103. Ma SW, Benzie IF, Chu TT, et al. Effect of Panax ginseng supplementation on biomarkers of glucose tolerance, antioxidant status and oxidative stress in type 2 diabetic subjects: results of a placebo-controlled human intervention trial. Diabetes Obes Metab 2008; 10: 1125–1127.
- 104. Attele AS, Zhou YP, Xie JT, et al. Antidiabetic effects of Panax ginseng. Diabetes 2002; 51: 1851–1858.
- Hou CC, Lin SJ, Cheng JT, et al. Hypoglycemic dimeric guianolides and a lignan glycoside from Lactuca indica. J Nat Prod 2003; 66: 625–629.
- 106. Krishnakumar K, Augusti KT and Vijavammal PL. Hypoglycaemic and anti-oxidant activity of Salacia oblonga wall extract in streptozotocininduced diabetic rats. Indian J Physiol Pharmacol 1999; 43: 510–514.
- 107. Ruikar AD, Khatiwora E and Ghayal NA. Studies on aerial parts of *Artemisia pallens* wall for phenol, flavonoid and evaluation of antioxidant activity. *J Pharm Bioallied Sci* 2011; 2: 302–305.
- 108. Basnet P, Kadota S and Terashima S. Two new 2-arylbenzofuran derivatives from hypoglycaemic activity-bearing fractions of *Morus insignis*. *Chem Pharm Bull (Tokyo)* 1993; 41: 1238–1243.
- 109. Silva RM, Santos FA, Rao VS, *et al.* Blood glucose and triglyceride lowering effect of trans-dehydrocrotonin, a diterpene from *Croton*

- cajucara Benth in rats. Diabetes Obes Metab 2001; 3: 452–456.
- 110. Borquez J, Loyola LA, Morales G, et al. Azorellane diterpenoids from *Laretia acaulis* inhibit nuclear factor-kappa B activity. *Phytother Res* 2007; 21: 1082–1086.
- 111. Fuentes NL, Sagua H, Morales G, et al. Experimental antihyperglycemic effect of diterpenoids of *Laretia acaulis* and *Azorella* compacta Phil (Umbelliferae) in rats. *Phytother* Res 2005; 19: 713–716.
- 112. Aniszewski T. *Alkaloids: chemistry, biology, ecology, and applications.* 2nd ed. Amsterdam: Elsevier, 2015, pp. 1–475.
- 113. Wanders RJA, van Grunsven EG and Jansen GA. Lipid metabolism in peroxisomes: enzymology, functions and dysfunction of the fatty acid α and β oxidation system in humans. *Biochem Soc Trans* 2000; 28: 141–148.
- 114. Muruganandan S, Srinivasan K, Gupta S, et al. Effect of mangiferin on hyperglycemia and atherogenicity in streptozotocin diabetic rats. *J Ethnopharmacol* 2005; 97: 497–501.
- 115. Gray AM and Flatt PR. Insulin-releasing and insulin-like activity of the traditional anti-diabetic plant *Coriandrum sativum* (coriander). *Br J Nutr* 1999; 81: 203–209.
- 116. Huang THW, Teoha AW, Lina BL, et al. The role of herbal PPAR modulators in the treatment of cardiometabolic syndrome. Pharmacol Res 2009; 60: 195–206.
- 117. Babu PS and Prince PSM. Antihyperglycaemic and antioxidant effect of hyponid, an ayurvedic herbomineral formulation in streptozotocininduced diabetic rats. *J Pharm Pharmacol* 2004; 56: 1435–1442.
- 118. Panigrahy SK, Bhatt R and Kumar A. Reactive oxygen species: sources, consequences and targeted therapy in type-II diabetes. *J Drug Target* 2017; 25: 93–101.
- 119. Kayaniyil S, Vieth R, Retnakaran R, et al. Association of vitamin D with insulin resistance and β-cell dysfunction in subjects at risk for type 2 diabetes. Diabetes Care 2010; 33: 1379–1381.
- Al-Timimi DJ and Ali AF. Serum 25(OH) D in diabetes mellitus type 2: relation to glycaemic control. 7 Clin Diagn Res 2013; 7: 2686–2688.
- 121. Talaei A, Mohamadi M and Adgi Z. The effect of vitamin D on insulin resistance in patients with type 2 diabetes. *Diabetol Metab Syndr* 2013; 5: 8.
- 122. Subramoniam A, Pushpangadan P and Rajasekharan S. Effects of *Artemisia pallens* Wall. on blood glucose levels in normal and alloxaninduced diabetic rats. *J Ethnopharmacol* 1996; 50: 13–17.

Visit SAGE journals online journals.sagepub.com/home/tae