

Article

# Docking Analysis of Some Bioactive Compounds from Traditional Plants against SARS-CoV-2 Target Proteins

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**Abstract:** COVID-19 is still a global pandemic that has not been stopped. Many traditional medicines have been demonstrated to be incredibly helpful for treating COVID-19 patients while fighting the disease worldwide. We introduced 10 bioactive compounds derived from traditional medicinal plants and assessed their potential for inhibiting viral spike protein (S-protein), Papain-like protease (PLpro), and RNA dependent RNA polymerase (RdRp) using molecular docking protocols where we simulate the inhibitors bound to target proteins in various poses and at different known binding sites using Autodock version 4.0 and Chimera 1.8.1 software. Results found that the chicoric acid, quinine, and withaferin A ligand strongly inhibited CoV-2 S -protein with a binding energy of  $-8.63$ ,  $-7.85$ , and  $-7.85$  kcal/mol, respectively. Our modeling work also suggested that curcumin, quinine, and demethoxycurcumin exhibited high binding affinity toward RdRp with a binding energy of  $-7.80$ ,  $-7.80$ , and  $-7.64$  kcal/mol, respectively. The other ligands, namely chicoric acid, demethoxycurcumin, and curcumin express high binding energy than the other tested ligands docked to PLpro with  $-7.62$ ,  $-6.81$ , and  $-6.70$  kcal/mol, respectively. Prediction of drug-likeness properties revealed that all tested ligands have no violations to Lipinski's Rule of Five except cepharanthine, chicoric acid, and theaflavin. Regarding the pharmacokinetic behavior, all ligand predicted to have high GI-absorption except chicoric acid and theaflavin. At the same way chicoric acid, withaferin A, and withanolide D predicted to be substrate for multidrug resistance protein (P-gp substrate). Caffeic acid, cepharanthine, chicoric acid, withaferin A, and withanolide D also have no inhibitory effect on any cytochrome P450 enzymes. Promisingly, chicoric acid, quinine, curcumin, and demethoxycurcumin exhibited high binding affinity on SARS-CoV-2 target proteins and expressed good drug-likeness and pharmacokinetic properties. Further research is required to investigate the potential uses of these compounds in the treatment of SARS-CoV-2.

**Keywords:** traditional plants; SARS-CoV-2; bioactive compounds; molecular docking; pharmacokinetic

## 1. Introduction

Coronavirus disease 2019 (COVID-19), the highly infectious viral illness caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has had a disastrous effect

on the world's demographics, resulting in more than 4.5 million deaths worldwide. SARS-CoV-2 is a positive-sense RNA (30 kb) virus. Two types of proteins characterize the human coronaviruses HCoVs, structural proteins (Spike (S), Nucleocapsid (N), Matrix (M), and Envelope (E)), and non-structural proteins including the RdRp [1,2]. The membrane and envelope proteins are associated with virus assembly, while the spike protein plays the main role in facilitating virus entry via mediating its interaction with the transmembrane surface receptor on the host cells [3].

This virus contains S- protein, PLpro, and RdRp involved in viral entry, replication, and immune response evasion. Drugs targeting these proteins therefore have great potential for inhibiting the virus. The spike protein directly interacts with the peptidase domain (PD) of the angiotensin-converting enzyme 2 (ACE2) receptor [4], which technically marks the virus entry inside the cells [5]. ACE2 interaction with spike protein could aid in producing antivirals or a vaccine that can block CoV infection by targeting ACE2 [6]. The binding to the SARSCoV-2 spike protein at the same place where the original ACE2 PD domain interacts, which shows that 23-amino acids sequence independently has the potential to inhibit the interaction of the SARS-CoV-2 S- protein and ACE2 complex. Hence, the viral entrance. Interacting residues at the interface of the SARS-CoV-2 and ACE2 PD domain are ARG403 LYS417 GLY446 TYR449 TYR453 LEU455 PHEF456 TYR473 ALA475 GLY476 GLY485 PHE486 ASN487 TYR489 GLN493 TYR495 GLY496 GLNQ498 THR500 ASN501 GLY502 VAL503 TYR505 in SARS-CoV-2 S- protein [7].

Another viral protein, PLpro, which is vital for viral replication [8], is responsible for the proteolytic processing of the product of open reading frame 1a (ORF1a) in the replicase gene of CoV2, a large viral polyprotein containing non-structural proteins which form the replicase complex [9]. The peptide bond cleavage in the active site is catalyzed by a conserved catalytic triad comprised of residues CYS111, HIS272, and ASP286 [10]. In addition, PLpro possesses deubiquitinating and deISGylating capabilities [11] which interfere with critical signaling pathways leading to the expression of type I interferons, resulting in antagonistic effect on host innate immune response [12]. Therefore, inhibition of PLpro activity can halt viral replication and disrupt its role in host immune response evasion, making it an excellent anti-viral drug target. RdRp is also a vital enzyme for the life cycle of RNA viruses. It has been targeted in various viral infections (Hepatitis C virus (HCV), Zika virus (ZIKV), and human coronaviruses (HCoV) [13]. SARS-CoV-2 polymerase residues ARG 553, ARG 555, LYS 545, and ASN 691 interaction with some bioactive compounds play an important role in virus replication inhibition [14]. Thus, many research working in computer-aided drug design in generating new small molecules that might interact with SARS-CoV-2 S -protein, RdRp, and 3CL or Mpro protease proteins to combat the COVID-19 disease.

In the U.S., the Food and Drug Administration issued an emergency use authorization (EUA) for Pfizer's Paxlovid (nirmatrelvir tablets and ritonavir tablets, co-packaged for oral use) for the treatment of mild-to-moderate coronavirus disease (COVID-19) in adults and pediatric patients (12 years of age and older weighing at least 40 kg or about 88 pounds) with positive results of direct SARS-CoV-2 testing, and who are at high risk for progression to severe COVID-19, including hospitalization or death. Paxlovid consists of nirmatrelvir, which inhibits a SARS-CoV-2 protein to stop the virus from replicating, and ritonavir, which slows down nirmatrelvir's breakdown to help it remain in the body for a longer period at higher concentrations. Paxlovid is available by prescription only and should be initiated as soon as possible after diagnosis of COVID-19 and within five days of symptom onset [15]. Another antiviral drug use for the same purpose is Lagevrio (molnupiravir). Molnupiravir is a nucleoside analogue that inhibits SARS-CoV-2 replication by viral mutagenesis. The U.S. FDA has issued an EUA to make Molnupiravir available during the COVID-19 pandemic (Molnupiravir is not FDA-approved for any uses, including use as treatment for COVID-19. This medicine is still being studied, Molnupiravir may cause serious side effects [16].

The curative effects of traditional herbal formula sometimes are not necessarily by directly inhibiting or killing the virus, but through the integration of various aspects such

as relieving cytokines storm syndrome, protecting human tissues and organs, relieving immunological injury, and enhancing the body's ability [17].

Ayurveda, known as “The Science of Life,” an ancient traditional medicinal system that originated and is practiced in India, has been utilized for reducing SARS-CoV-2 infection and treating COVID-19-associated patients [18–20]. It describes many medicinal plants and herbs possessing a broad range of therapeutic usefulness in curing various kinds of ailments, diseases, and disorders, such as *Allium sativum* (Garlic), *W. somnifera* known commonly as ashwagandha, *Zingiber officinale* Roscoe (Ginger), *Tinospora cordifolia* (Giloy), *Ocimum sanctum* (Tulsi), *Curcuma longa* (Turmeric, Haldi), *Glycyrrhiza glabra* (Licorice, mulethi), and others [20–22].

In this study we aim to investigate docking characteristics of 10 bioactive compounds identified from traditional medicines (caffeic and chicoric acids from *Echinacea purpurea*; xanthorrhizol from *Java turmeric*; curcumin and demethoxycurcumin from *Curcuma longa* L.; Quinine from *Chinchona bark*; Theaflavin from *Black tea*; withaferin A and withanolide D from *Withania somnifera* L.; cepharanthine from *Stephania cepharantha*) on S-protein, RdRp and SARS-CoV-2 PLpro to explore the potential mode of the inhibitory activity toward SARS-CoV-2. The molecules that showed better hydrogen bonding and binding energy were selected further for drug likeness and pharmacokinetics prediction.

## 2. Results and Discussion

Herbal medicines and medicinal plant-based natural compounds provide a rich resource for novel antiviral drug development. Some natural medicines have been shown to possess antiviral activities against various virus strains including coronavirus, herpes simplex virus [23–25], influenza virus [25], human immunodeficiency virus [26], hepatitis B and C viruses [27], SARS, and MERS [28,29]. These compounds antiviral action mechanisms caused by influencing of the viral life cycle, such as viral entry, replication, assembly, and release, as well as virus-host-specific interactions. In this study we analyze the binding mode of 10 ligands Figure 1 that previously identified in some traditional plants Table 1 using computational docking analysis.

**Table 1.** Traditional medicinal source for bioactive compounds docking ligands toward COVID-19.

Ligands	Traditional Medicine Source	Region	Reported Biological Activity	References
1	<i>Citrullus colocynthis</i> (fruit coat), <i>C. colocynthis</i> (Seeds), <i>Paraguariensis</i> , <i>Mesona chinensis</i> <i>Echinacea purpurea</i>	Egypt, Poland, Argentina, China	Antiviral activities Antioxidant activity	[30] [31]
2	<i>Stephania cepharantha</i> <i>Stephania rotunda</i>	Japan	Anti-inflammatory activity, Antiparasitic activities, Anti-oxidative properties, Antiviral activities and Anti-HIV activity	[32–35]
3	<i>Sickle seagrass</i> , <i>starflower</i> , <i>sweet basil</i> , <i>African basil</i> , <i>Chicory</i> and <i>Echinacea purpurea</i>	Egypt, China, India, and North America	Antivirus, anti- inflammation, glucose and lipid homeostasis, Neuroprotection, antioxidation effects. Antimicrobial activity and Antioxidant activity	[36–38]
4	<i>Curcuma longa</i> L.	Southern Asia, China, India, Indonesia, Indochina	Anti-inflammatory activity, Antioxidant activity, Anti-bacteria activity Chemopreventive and Chemotherapeutic activity Anti-HIV activity, and Nematocidal activities	[39–41]

Table 1. Cont.

Ligands	Traditional Medicine Source	Region	Reported Biological Activity	References
5	<i>Curcuma longa</i> L.	Southern Asia, China, India, Indonesia, Indochina,	Antimicrobial activity, Anti-inflammatory activity, Antioxidant activity, Platelet aggregation inhibitory activity, Antiallergy activity, Anticancer activity	[42–44]
6	<i>Chinchona bark</i>	Peru, Bolivia, Colombia, Ecuador, India, and Sri Lanka.	Antimalarial activity, Antioxidant activity, Anti-cancer agent, Anti-inflammatory, Antiparasitic activity and Antimicrobial property	[45]
7	Black tea	Asia and Europe	Antioxidant activity and Anti-cancer activity	[46,47]
8	<i>Withania somnifera</i> L.	Sri Lanka	Anti-cancer activity and Anti-COVID activity	[48]
9	<i>Withania somnifera</i> L. <i>Datura metel</i> L. leaves	India	Anti-inflammatory and Antioxidant activity	[49,50]
10	Java turmeric <i>Curcuma</i> species ( <i>C. zedoaria</i> , <i>C. xanthorrhiza</i> , <i>C. aeruginosa</i> and <i>C. mangga</i> )	India and Southeast Asia	Anti-inflammatory, Antioxidant, and Anti-cancer activities	[51,52]

1: Caffeic acid; 2: Cepharanthine; 3: Chicoric acid; 4: Curcumin; 5: Demethoxycurcumin; 6: Quinine; 7: Theaflavin; 8: withaferin A; 9: withanolide D; 10: Xanthorrhizol.

All tested ligands showed hydrogen interaction to papain-like protease (6W9C) except cepharanthine and theaflavin. The ligand docking on CYS111, HIS272, and ASP286 catalytic residues (Figure 2A) ranked by binding energy  $\Delta G$  in kcal/mol were as follows: chicoric acid > demethoxycurcumin > curcumin > withanolide D > xanthorrhizol > withaferin A > caffeic acid > quinine. It is known that the high binding affinity of the drug compounds depends on the type and amount of bonds occurring with the target protein [53]. The binding energies were  $-7.62$ ,  $-6.81$ ,  $-6.70$ ,  $-6.58$ ,  $-6.16$ ,  $-6.13$ ,  $-5.70$ , and  $-5.67$  kcal/mol, respectively (Table 2; Figure 3).

The first reported bioactive effect of chicoric acid is its ability to inhibit infection with human immunodeficiency virus 1 (HIV-1) [54]. Several studies reported that chicoric acid inhibits infection with HIV-1 by deactivating the HIV-1 integrase. HIV-1 integrase is a multidomain enzyme required for integration of viral DNA into the host genome, a critical step in viral replication [55]. Inhibition of HIV-1 integration by chicoric acid results in stopping virus replication, leading to increased T-lymphoblastoid cell viability [56]. Chicoric acid found in many Egyptian source as *Chicory* and *Echinacea purpurea* (Table 1). Despite that cepharanthine (CEP) showed no binding behavior toward papain-like protease, it has unique anti-inflammatory, antioxidative, immunomodulating, antiparasitic, and antiviral properties. It can suppress nuclear factor-kappa B (NF- $\kappa$ B) activation, lipid peroxidation, nitric oxide (NO) production, cytokine production, and expression of cyclooxygenase; all of which are crucial to viral replication and inflammatory response [33]. Against SARS-CoV-2 and homologous viruses, CEP predominantly inhibits viral entry and replication at low doses [57].

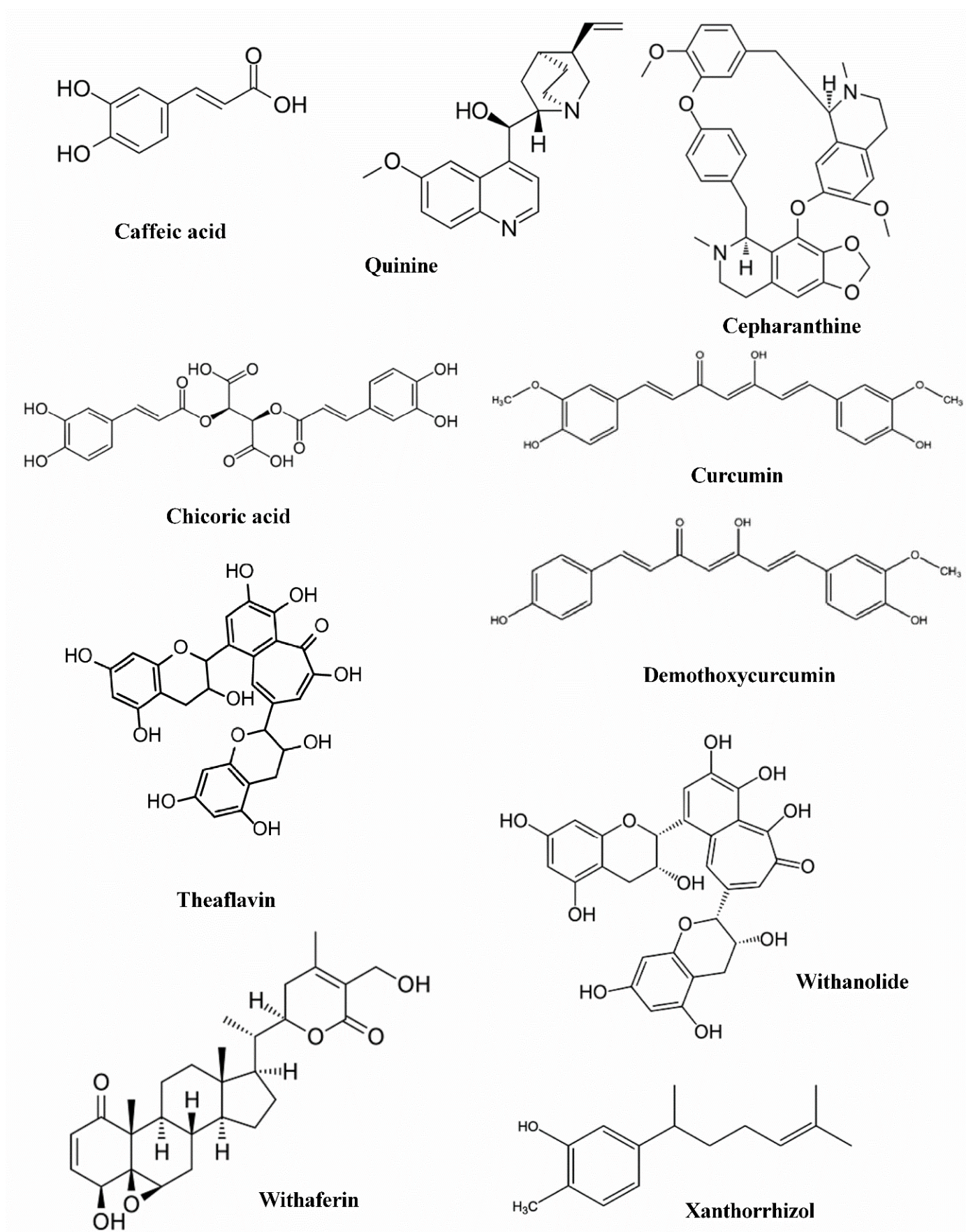
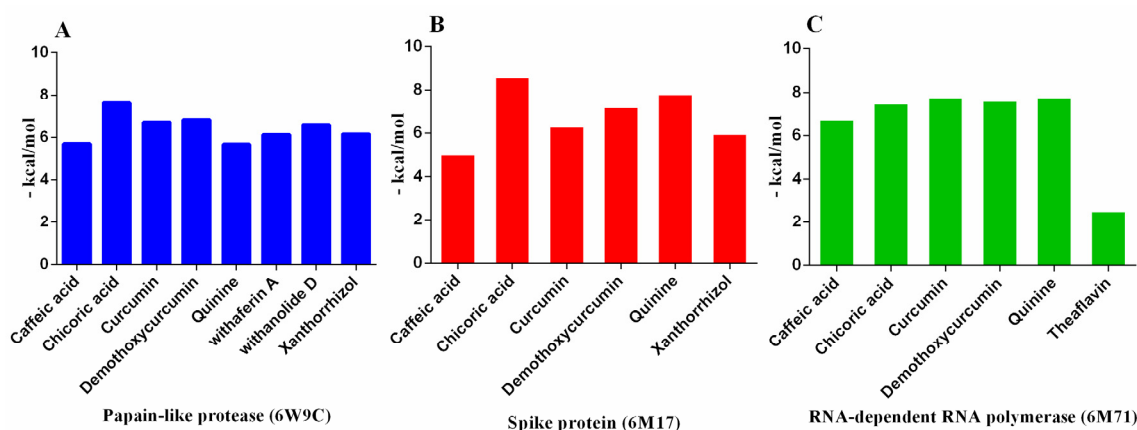


Figure 1. Structures of docking ligands from ZINC drug database.

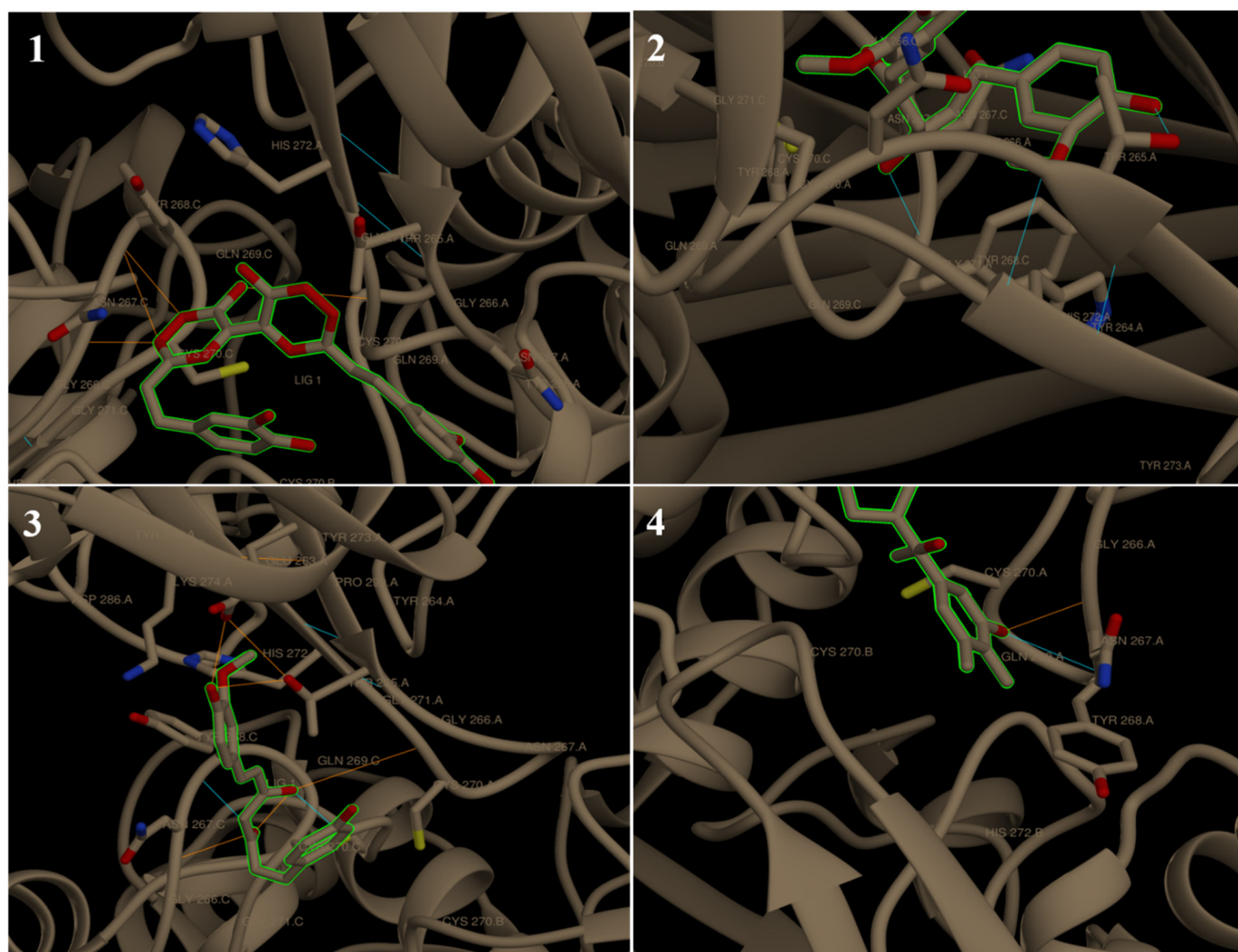


**Figure 2.** Histogram showing molecular docking results between (A) 6W9C, (B) 6M17, and (C) 6M71 and several drug candidate compounds (the binding energy value  $\Delta G$  is shown in minus kcal/mol).

**Table 2.** Molecular docking analysis of several compounds against papain-like protease (6W9C), RNA-dependent RNA polymerase (6M71), and spike protein (6M17) of SARS CoV-2.

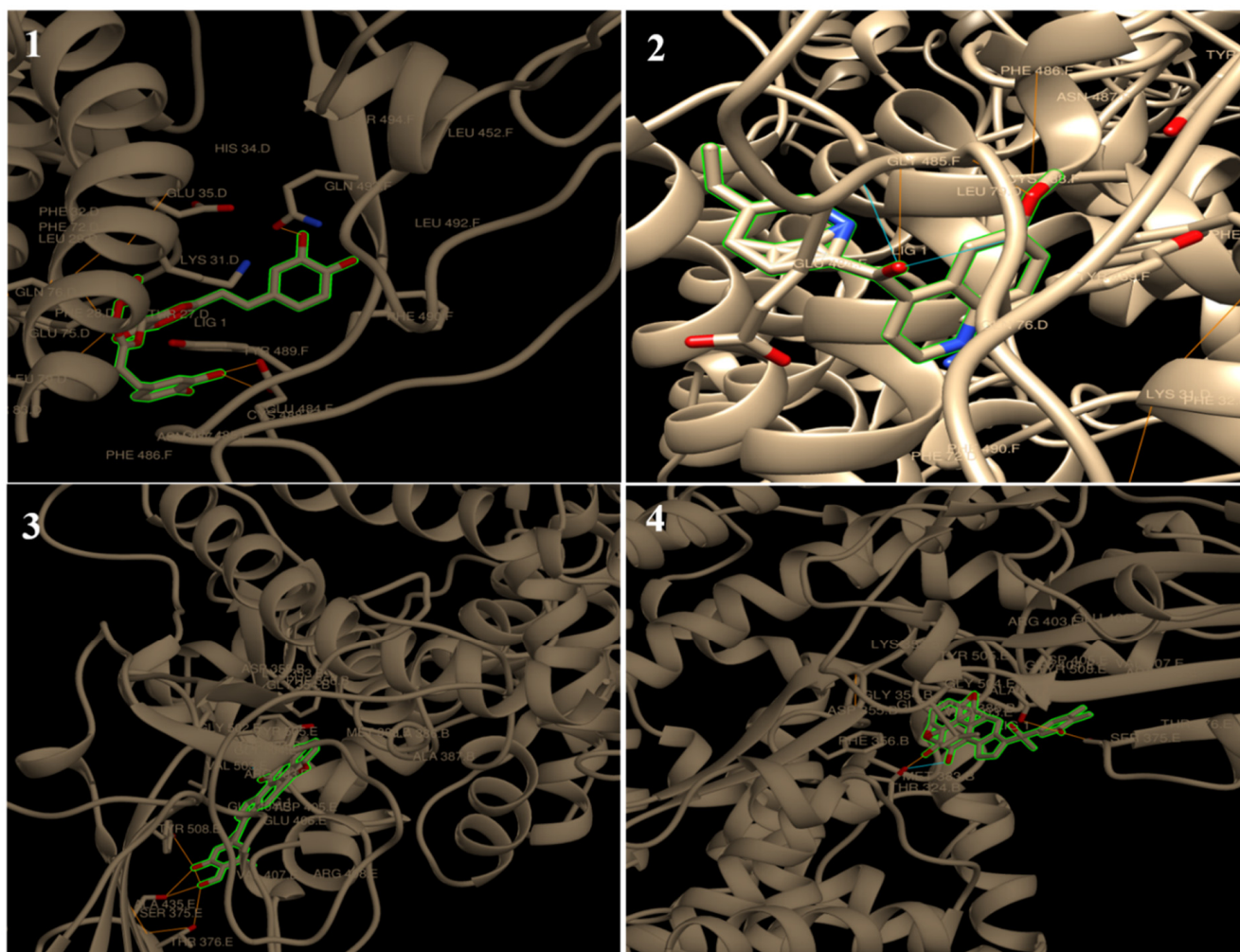
Ligand Name	1	2	3	4	5	6	7	8	9	10
Molecular formula	C <sub>9</sub> H <sub>8</sub> O <sub>4</sub>	C <sub>37</sub> H <sub>38</sub> N <sub>2</sub> O <sub>6</sub>	C <sub>22</sub> H <sub>18</sub> O <sub>12</sub>	C <sub>21</sub> H <sub>20</sub> O <sub>6</sub>	C <sub>20</sub> H <sub>18</sub> O <sub>5</sub>	C <sub>20</sub> H <sub>24</sub> N <sub>2</sub> O <sub>2</sub>	C <sub>29</sub> H <sub>24</sub> O <sub>12</sub>	C <sub>28</sub> H <sub>38</sub> O <sub>6</sub>	C <sub>28</sub> H <sub>38</sub> O <sub>6</sub>	C <sub>15</sub> H <sub>22</sub> O
Classification	Phenolics	Alkaloid	Phenylpropanoid	Phenolics	Phenolics	Alkaloid	Phenolics	Phenolics	Phenolics	Sesquiterpenoid
	6W9C									
Binding energy $\Delta G$	-5.70	-	-7.62	-6.70	-6.81	-5.67	-	-6.13	-6.58	-6.16
No. of H bonding	6	-	4	3	6	2	-	1	2	2
Binding sites	CYS 270, GLN 269, TYR 268, ASN 267 and THR 265	-	CYS 270, TYR 268, ASN 267 and ASP 286	THR 265 and TYR 268	CYS 270, TYR 268, ASN 267, THR 265 and GLU 263	ASP 286 and LEU 290	-	TYR 264	ASN 267 and TYR 268	ASN 267 and TYR 268
	6M71									
Binding energy $\Delta G$	-6.75	-	-7.50	-7.80	-7.64	-7.80	-2.55	-6.27	-6.10	-5.33
No. of H bonding	6	-	4	11	8	4	7	4	7	2
Binding sites	THR 680, ARG 555, ARG 553, THR 556, and SER 682	-	ALA 554, ARG 553 and ARG 555	ARG 555, ARG 553, THR 556, and LYS 545	ARG 555, and THR 556,	ARG 555, THR 556, SER 682 and ALA 554	ARG 553, THR 556, and SER 682	THR 556, and SER 682	THR 556, SER 682 and ASP 623	TYR 455 and ALA 554
	6M17									
Binding energy $\Delta G$	-5.06	-5.26	-8.63	-6.34	-7.23	-7.85	-7.43	-7.85	-7.78	-6.00
No. of H bonding	2	1	4	4	1	4	7	12	7	3
Binding sites	ASN 487	GLN 76	GLU 484, CYS 488, GLN 493 and GLN 76	ASP 355, THR 500 and THR 324	ASN 487	CYS 488, GLY 484 and ASN 487	THR 324, GLY 404, ARG 408 and THR 508	THR 324, SER 375, TYR 376 and TYR 508	THR 324, SER 375, and TYR 508	THR 324 and VAL 503

1: Caffeic acid; 2: Cepharanthine; 3: Chicoric acid; 4: Curcumin; 5: Demethoxycurcumin; 6: Quinine; 7: Theaflavin; 8: withaferin A; 9: withanolide D; 10: Xanthorrhizol.



**Figure 3.** Chimera visualization of 6W9C docking with chicoric acid (1), curcumin (2), demethoxycurcumin, and (3) withanolide D (4). The yellow dots show H-bonds.

Moreover, all tested ligands showed H-interaction to spike protein (6M17). The ligand docking on GLY 502, TYR 489 and TYR 505 (Figure 2B) ranked by binding energy in kcal/mol were as follows: chicoric acid > quinine = withaferin A > withanolide D > theaflavin > demethoxycurcumin > curcumin > xanthorrhizol > cepharanthine > caffeic acid. The binding energies were  $-8.63$ ,  $-7.85$ ,  $-7.85$ ,  $-7.78$ ,  $-7.43$ ,  $-7.23$ ,  $-6.34$ ,  $-6.00$ ,  $-5.26$  and  $-5.06$  kcal/mol respectively (Table 2; Figure 4). Cepharanthine (CEP) is a natural alkaloid, which has been widely used to treat many of the acute and chronic diseases. It is usually used in Japan in the form of *Stephania cepharantha* and *Stephania rotunda* (Table 1) for its anti-inflammatory, antiparasitic, anti-oxidative, antiviral, and anti-HIV activities [32–35]. In 2021 a Japanese research team reported the anti-COVID-19 activity of the combining cepharanthine/nelfinavir providing synergistic antiviral effects that CEP and nelfinavir inhibit SARS-CoV-2 entry and RNA replication, respectively [58]. In 2021, a Japanese research team reported that the combination of cepharanthine and nelfinavir inhibited SARS-CoV-2 entrance and RNA replication, offering synergistic antiviral effects [58].

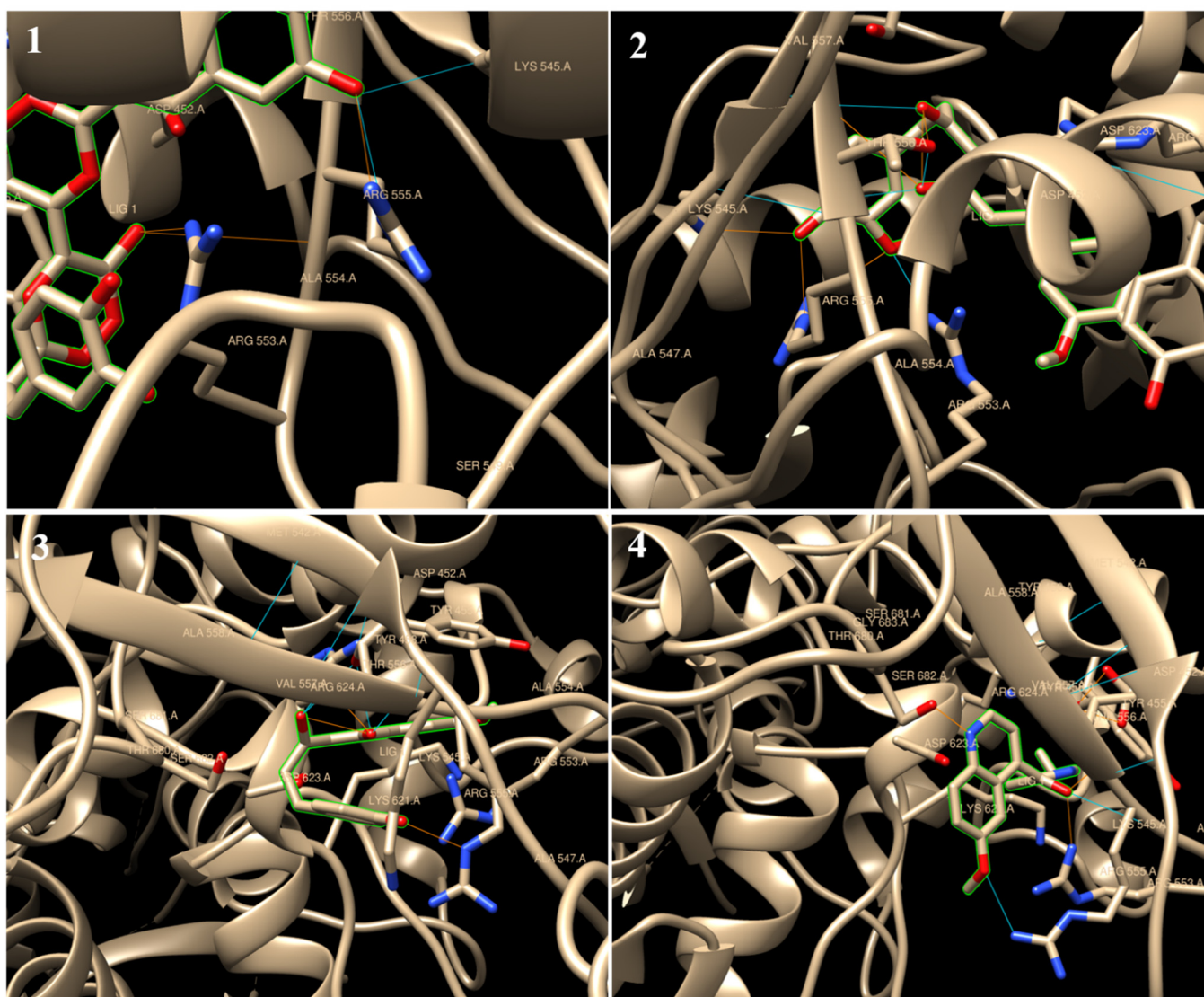


**Figure 4.** Chimera visualization of 6M17 docking with chicoric acid (1), quinine (2), withaferin A (3) and withanolide D (4). The yellow dots show H-bonds.

This previous combination based on *in silico* docking simulation that confirms that CEP molecule can bind to SARS-CoV-2 S-protein and interfere with the spike engagement to its receptor, angiotensin-converting enzyme 2 (ACE2) [59,60]. Our results strongly supported this mechanism of CEP toward SARS-CoV-2 S-protein. However, chicoric acid, quinine, withaferin A, withanolide D, theaflavin, demethoxycurcumin, curcumin, and xanthorrhizol expressed higher binding energy than cepharanthine in case SARS-CoV-2 S-protein binding. From all of this we supposed that chicoric acid and quinine may play the same role in this combination (cepharanthine/nelfinavir) beside their acting mechanism toward SARS-CoV-2, PLpro and RdRp that may be promising to limit SARS-CoV-2 proliferation than CEP.

Regarding to RdRp (6M71), all tested ligands showed H-interaction except Cepharanthine. The ligand docking on ARG 553, ARG 555, LYS 545, and ASN 691 (Figure 2C) ranked by binding energy  $\Delta G$  in kcal/mol were as follows: curcumin = quinine > demethoxycurcumin > chicoric acid > caffeic acid > withaferin A > withanolide D > xanthorrhizol > theaflavin. The binding energies were  $-7.80$ ,  $-7.80$ ,  $-7.64$ ,  $-7.50$ ,  $-6.75$ ,  $-6.27$ ,  $-6.10$ ,  $-5.33$ , and  $-2.55$  kcal/mol, respectively (Table 2; Figure 5).





**Figure 5.** Chimera visualization of 6M71 docking with chicoric acid (1), curcumin (2), demethoxycurcumin (3) and quinine (4). The yellow dots show H-bonds.

Curcumin (CC) and its analogues are the main phytonutrients of turmeric (*Curcuma longa* L.) and other *Curcuma* spp., which are widely used around the world as culinary spices, traditional medicine as well as a popular dietary supplement ingredient due to its wide range of health benefits including anti-inflammation [61], anti-cancer [62], cardiovascular regulation [63], respiratory [64], and immune system benefits [65]. In addition, the suppression of multiple cytokines by curcumin suggested that it may be a useful approach in treating Ebola patients against cytokine storm [66]. As severe cases of COVID-19 are often associated with cytokine release syndrome, the use of anti-inflammatory molecules may reduce the proinflammatory cytokines involved. Quinine, is an extract of the bark of the Chinchona tree (native to the Andes of South America), that was used to treat feverish infections, particularly malaria, for hundreds of years almost worldwide [67]. *Curcuma longa* L. and *Chinchona bark* are rich source of curcumin and quinine respectively (Table 1).

Table 3 presents the drug likeness and pharmacokinetic properties of the tested ligands. Lipinski's Rule of Five is generally used as an indicator of the drug likeness and pharmacological activities. In humans, this would make them orally active medications [68]. All the tested compounds are in the molecular weight range of 180.16 to 474.37 Da (<500 Da), except for cepharanthine and theaflavin are 606.71 and 564.49 Da. It is well known that

drug molecules typically have low molecular weight (<500 kDa) are transported, diffused, and absorbed easily compared with large molecules [69]. All the tested compounds also have fewer than 15 rotatable bonds, and all have less than 5 hydrogen bond donors (NH and OH), except for quinine and theaflavin. In addition, the numbers of hydrogen bond acceptors (O and N atoms) predicted in all compounds are less than 10, except for chicoric acid and theaflavin (Table 3). At the same time, permeability (logP) of these ligands has also been investigated, and it was found that these ligands exhibited logP values of less than 5 except cepharanthine. Moreover, the topological polar surface area (TPSA) values of all ligands are less than 140 Å, except for chicoric acid and theaflavin. TPSA is closely related to the H-bonding ability of a compound [70]. The TPSA and the logP values are the two essential characteristics in the analysis of the bioavailability of drug molecules and permeability toward bio-membranes [71]. Compounds with 10 rotatable bonds and TPSA of ≤140 Å can be predicted to have good bioavailability [72]. As a result, all of the ligands examined have good bioavailability, with the exception of chicoric acid and theaflavin, which have TPSA values of 208.12 and 217.60, respectively. Pharmacokinetic prediction provides information to evaluate the time course of drugs and their effects on the body to design an appropriate drug regimen for any patient [73].

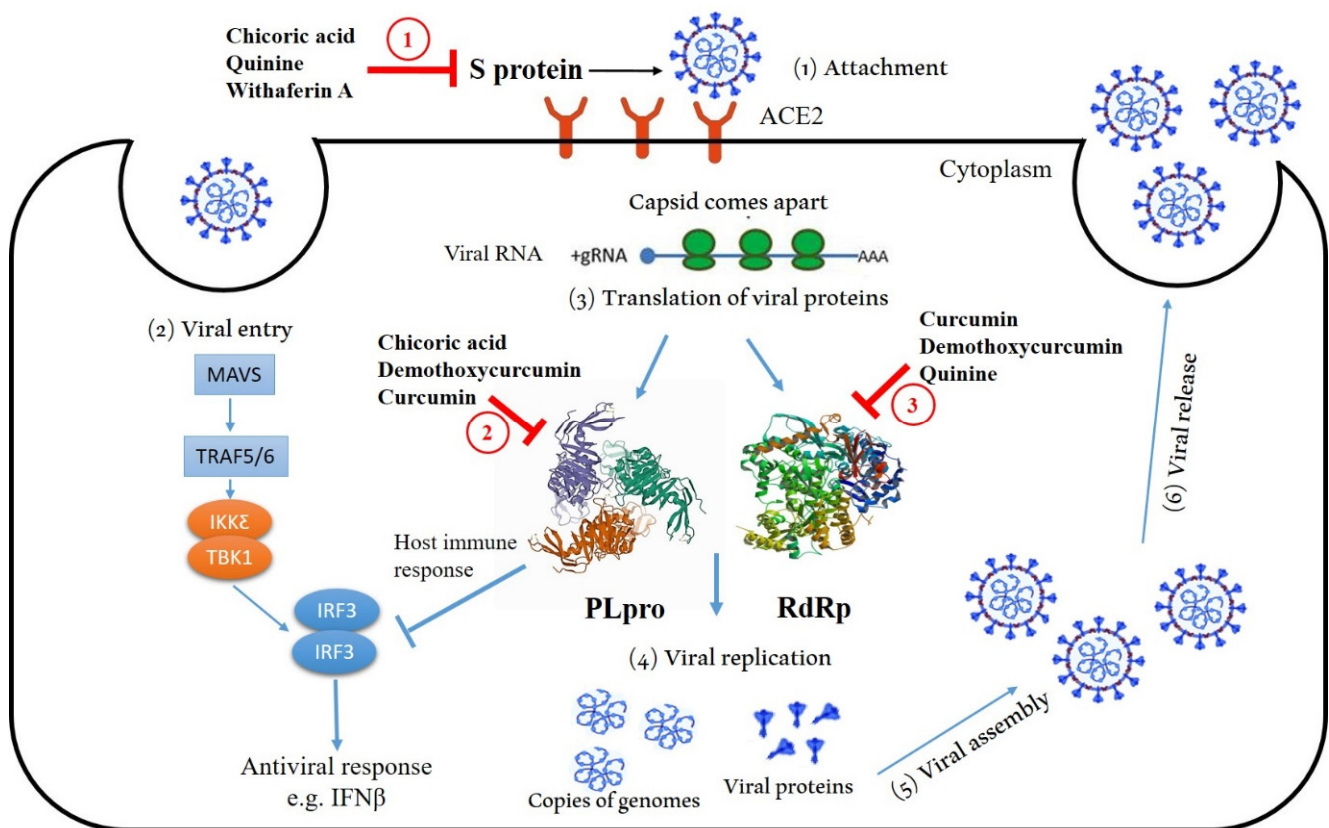
**Table 3.** Predicted drug likeness and pharmacokinetics of SARS-CoV-2 potential inhibitors.

Ligands Name	1	2	3	4	5	6	7	8	9	10
<b>Lipinski's Rule of Five</b>										
Molecular weight (<500 Da)	180.16	606.71	474.37	368.38	338.35	324.42	564.49	470.60	470.60	218.33
LogP (<5)	0.93	5.35	1.01	3.03	3.00	2.81	1.31	2.29	2.36	4.34
No. rotatable bonds (<15)	2	2	11	8	7	4	2	3	2	4
No. H-Bond donors (5)	3	0	6	2	2	1	9	4	4	1
No. H-bond acceptors (<10)	4	8	12	6	5	4	12	6	6	1
TPSA Å	77.76	61.86	208.12	93.06	83.83	45.59	217.60	115.06	115.06	20.23
Violations	0	0	0	0	0	0	3	0	0	0
<b>Pharmacokinetics</b>										
GI absorption	High	High	Low	High	High	High	Low	High	High	High
BBB	No	No	No	No	No	Yes	No	No	No	Yes
P-gp substrate	No	No	Yes	No	No	No	No	Yes	Yes	No
CYP1A2 inhibitor	No	No	No	No	Yes	No	No	No	No	No
CYP2C19 inhibitor	No	No	No	No	No	No	No	No	No	No
CYP2C9 inhibitor	No	No	No	Yes	Yes	No	Yes	No	No	Yes
CYP2D6 inhibitor	No	No	No	No	No	Yes	No	No	No	Yes
CYP3A4 inhibitor	No	No	No	Yes	Yes	No	Yes	No	No	No

1: Caffeic acid; 2: Cepharanthine; 3: Chicoric acid; 4: Curcumin; 5: Demothoxycurcumin; 6: Quinine; 7: Theaflavin; 8: withaferin A; 9: withanolide D; 10: Xanthorrhizol.

Our results of pharmacokinetic prediction (Table 3) showed that all tested ligands demonstrate a high absorption rate in the GI tract, except for chicoric acid and theaflavin. Moreover, none of the tested ligands were predicted to be able to pass through the blood–brain barrier, except for quinine and xanthorrhizol. Additionally, chicoric acid, withaferin A and withanolide D are the only compounds predicted to act as a substrate for P-gp, which decrease their further clinical application [74]. The predicted pharmacokinetic behavior indicates that only caffeic acid, cepharanthine, chicoric acid, withaferin A, and withanolide D would have no inhibitory effect on any cytochrome P450 enzymes (CYP1A2, CYP2C19, CYP2D6, and CYP3A4). Furthermore, CYP2C9 activity is only known to be affected by curcumin, demothoxycurcumin, theaflavin and xanthorrhizol. Cytochrome P450 enzymes are essential for the metabolism of many chemicals. Although this class contains more than 50 enzymes, six of them metabolize 90% of compounds. Cytochrome P450 enzyme inhibition causes unwanted adverse effects or therapeutic failures [75]. The proposed mechanism of SARS-CoV-2 inhibition by tested compounds is summarized in Figure 6; chicoric acid, quinine and withaferin A, interacting with S protein, inhibits viral ability to attach to human

ACE2. Additionally, chicoric acid, demethoxycurcumin, and curcumin interacting with PL pro, inhibits viral replication, moreover, curcumin, demethoxycurcumin, and quinine interacting with RdRp, inhibits viral replication. Furthermore, interferon (IFN) production is triggered by virus infections that cause accumulation of RNA species in the cytoplasm, initiate the oligomerization of an essential mitochondrial antiviral signaling protein, MAVS, that serves as a scaffold for the activation of TNF receptor-associated factor (TRAF) family proteins, TRAF2, TRAF3, TRAF5, and TRAF6. TRAF proteins catalyze the assembly of K63-linked ubiquitin chains that are required for the activation of serine kinases, IKKa, IKKb, IKKc, IKKe, and TBK1. Kinase activation triggers the phosphorylation and nuclear import of IRF3, driving the production of primary antiviral effectors including IFNs [76]. On the other hand, PLpro of SARS-CoV-2 interfere with innate immune response by directly cleaving IRF3, inhibits IFN $\beta$  production. In general, in silico studies are a great starting point when looking for new therapeutic targets. In the event of an unpredictable pandemic, such as the recent COVID-19 epidemic, in silico research are critical. Drug targets cannot be discovered in a short period of time using existing methods. To reach bulk targets, it is unavoidable to have extremely rapid procedures. In silico approaches can be used to speed up the search for drug targets in natural medicinal plants. This article's results warrant further evaluation of the potential anti-SARS-CoV-2 activity of these ligands in vitro.



**Figure 6.** The proposed mechanism of SARS-CoV-2 inhibition by tested compounds.

Steps from one to six describe the replication cycle of SARS-CoV-2; attachment (1), viral entry (2), translation of viral proteins (3), viral replication (4), assembly (5), and release (6). In red: chicoric acid, quinine, and withaferin A, interacting with S protein, inhibiting its viral ability to attach to human ACE2 (1). Additionally, chicoric acid, demethoxycurcumin, and curcumin interacting with PL pro inhibits viral replication (2), moreover, curcumin, demethoxycurcumin, and quinine interacting with RdRp inhibits viral replication (3). Interferon (IFN) production is triggered by virus infections that cause accumulation of RNA species in the cytoplasm, initiating the oligomerization of an essential mitochondrial

antiviral signaling protein, MAVS, which serves as a scaffold for the activation of TNF receptor-associated factor (TRAF) family proteins, TRAF2, TRAF3, TRAF5, and TRAF6. TRAF proteins catalyze the assembly of K63-linked ubiquitin chains that are required for the activation of serine kinases, IKKa, IKKb, IKKc, IKKe, and TBK1. Kinase activation triggers the phosphorylation and nuclear import of IRF3, driving the production of primary antiviral effectors including IFNs [74]. In blue: PLpro of SARS-CoV-2 interfere with innate immune response by directly cleaving IRF3, inhibits IFN $\beta$  production.

### 3. Experimental Section

#### 3.1. Docked SARS-CoV-2 Protein Structures

The structure of the SARS-CoV-2 papain-like protease (6W9C), RNA-dependent RNA polymerase (6M71), and spike protein (6M17) used as a target for ligands binding was downloaded from RCSB website [77]. PDB (Protein Data Bank) has enabled breakthroughs in research, such as this study, and education worldwide [78].

#### 3.2. Ligands and Drug Scan

The three-dimensional (3D) structures of all tested ligands were drawn in ACD/Chem-Sketch and then docked into the rigid binding pocket of 6W9C, 6M71, and 6M17 of SARS CoV-2. The compounds used in the present study were caffeic acid, cepharanthine, chicoric acid, curcumin, demothoxycurcumin, quinine, theaflavin, withaferin a, withanolide d and xanthorrhizol. The drug likeness and pharmacokinetic properties were calculated using the SWISSADME prediction website (<http://www.swissadme.ch/>) (accessed on 15 December 2021) [68,79]. Figure 1 shows the structure of the 10 docked ligands we used herein.

#### 3.3. Determination of SARS-CoV-2 Proteins Binding Hits

Table 4 contains the amino acids binding hits of the SARS-CoV-2 polymerase as described by Afonine et al. [80], papain-like protease (6W9C) as described by Baez-Santos et al., 2015 and spike protein (6M17) as described by Baig et al. [7].

**Table 4.** Protein target amino acids for molecular docking.

Amino Acids Hits	Papain-Like Protease (PLpro) (6W9C)	RNA Dependent RNA Polymerase (RdRp) (6M71)	Spike Protein (S Protein) (6M17)
	ASP 286, HIS 272, and CYS 111	ARG 553, ARG 555, and LYS 545	GLY 502, TYR 489, and TYR 505

#### 3.4. Molecular Docking

Ligand optimization was performed using Open Babel, converting ligands from mol into the PDB format. Autodock version 4.0 was used for protein optimization through the removal of water and other atoms and then addition of a polar hydrogen group. Ligand tethering of the protein was performed by regulating the genetic algorithm (GA) parameters using 10 runs of the GA criteria. Docking analyses and determination of hydrogen bonds (H-bonds) were conducted using Chimera 1.8.1 [81].

### 4. Conclusions

Antiviral treatments targeting the coronavirus disease 2019 are urgently required. We screened a panel of ligands from traditional plant sources already used in medicinal fields and some of them described for severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Chicoric acid, quinine, curcumin, and demothoxycurcumin exhibited high binding affinity on SARS-CoV-2 S- protein, PLpro, and RdRp. Binding of these proteins interfere with the viral entry, replication, and immune response evasion. Therefore, these compounds may have a great potential for inhibiting the virus. Our work provides data about the ligand mechanism toward SARS-CoV-2 target proteins and allows the comparison between them based on scientific information. Further in vitro cell-based investigations

will be needed for chicoric acid, quinine, curcumin, and demothoxycurcumin compounds to confirm their previous results and determine their antiviral mechanism at cell level.

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## References

1. Elfiky, A.A. Zika Virus: Novel Guanosine Derivatives revealed strong binding and possible inhibition of the polymerase. *Future Virol.* **2017**, *12*, 721–728. [CrossRef]
2. Hasan, A.; Paray, B.A.; Hussain, A.; Qadir, F.A.; Attar, F.; Aziz, F.M.; Sharifi, M.; Derakhshankhah, H.; Rasti, B.; Mehrabi, M.; et al. A review on the cleavage priming of the spike protein on coronavirus by angiotensin-converting enzyme-2 and furin. *J. Biomol. Struct. Dyn.* **2020**, *39*, 1–9. [CrossRef] [PubMed]
3. Li, F. Structure, function, and evolution of coronavirus spike proteins. *Annu. Rev. Virol.* **2016**, *3*, 237–261. [CrossRef] [PubMed]
4. Yan, R.; Zhang, Y.; Li, Y.; Xia, L.; Guo, Y.; Zhou, Q. Structural basis for the recognition of SARS-CoV-2 by full-length human ACE2. *Science* **2020**, *367*, 1444–1448. [CrossRef] [PubMed]
5. Hoffmann, M.; Kleine-Weber, H.; Schroeder, S.; Kruger, N.; Herrler, T.; Erichsen, S. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* **2020**, *181*, 271–280. [CrossRef]
6. Jando, J.; Camargo, S.M.R.; Herzog, B.; Verrey, F. Expression and regulation of the neutral amino acid transporter B0AT1 in rat small intestine. *PLoS ONE* **2017**, *12*, e0184845. [CrossRef]
7. Baig, M.S.; Alagumuthu, M.; Rajpoot, S.; Saqib, U. Identification of a Potential Peptide Inhibitor of SARS-CoV-2 Targeting its Entry into the Host Cells. *Drugs R&D* **2020**, *20*, 161–169. [CrossRef]
8. Harcourt, B.H.; Jukneliene, D.; Kanjanahaluethai, A.; Bechill, J.; Severson, K.M.; Smith, C.M. Identification of severe acute respiratory syndrome coronavirus replicase products and characterization of papain-like protease activity. *J. Virol.* **2004**, *8*, 13600–13612. [CrossRef]
9. Wertz, I.E.; Murray, J.M. Structurally-defined deubiquitinase inhibitors provide opportunities to investigate disease mechanisms. *Drug Discov. Today Technol.* **2019**, *31*, 109–123.
10. Baez-Santos, Y.M.; St John, S.E.; Mesecar, A.D. The SARS coronavirus papain-like protease: Structure, function and inhibition by designed antiviral compounds. *Antivir. Res.* **2015**, *115*, 21–38. [CrossRef] [PubMed]
11. Sulea, T.; Lindner, H.A.; Purisima, E.O.; Ménard, R. Deubiquitination, a new function of the severe acute respiratory syndrome coronavirus papainlike protease. *J. Virol.* **2005**, *79*, 4550–4551. [CrossRef] [PubMed]
12. Bekes, M.; van Noort, G.J.V.; Ekkebus, R.; Ovaa, H.; Huang, T.T.; Lima, C.D. Recognition of Lys48-linked di-ubiquitin and deubiquitinating activities of the SARS coronavirus papain-like protease. *Mol. Cell* **2016**, *62*, 572–585. [CrossRef] [PubMed]
13. Elfiky, A.A.; Ismail, A. Molecular dynamics and docking reveal the potency of novel GTP derivatives against RNA dependent RNA polymerase of genotype 4a HCV. *Life Sci.* **2019**, *238*, 116958. [CrossRef] [PubMed]
14. Abd El-Aziz, N.M.; Awad, O.M.; Shehata, M.S.; El-Sohaimy, S.A. Inhibition of the SARS-CoV-2 RNA-Dependent RNA Polymerase by Natural Bioactive Compounds: Molecular Docking Analysis. *Egypt. J. Chem.* **2021**, *64*, 1989–2001.
15. Fact Sheet for Healthcare Providers: Emergency Use Authorization for Paxlovid™. Available online: <https://www.fda.gov/media/155050/download> (accessed on 22 December 2021).
16. Mahase, E. COVID-19: UK becomes first country to authorise antiviral molnupiravir. *BMJ* **2021**, *375*, n2697. [CrossRef] [PubMed]
17. Liu, L. Traditional Chinese medicine contributes to the treatment of COVID-19 patients. *Chin. Herb. Med.* **2020**, *12*, 95–96. [CrossRef]
18. Gautam, K.; Adhikari, R.P.; Gupta, A.S.; Shrestha, R.K.; Koirala, P.; Suraj, K.S. Self-reported psychological distress during the COVID-19 outbreak in Nepal: Findings from an online survey. *BMC Psychol.* **2020**, *8*, 127. [CrossRef]
19. Rastogi, A.; Bhansali, A.; Khare, N.; Suri, V.; Yaddanapudi, N.; Sachdeva, N.; Puri, G.D.; Malhotra, P. Short term, high-dose vitamin D supplementation for COVID-19 disease: A randomised, placebo-controlled, study (SHADE study). *Postgrad. Med. J.* **2022**, *98*, 87–90. [CrossRef]

20. Singh, P.; Guleri, R.; Singh, V.; Kaur, G.; Kataria, H.; Singh, B.; Kaur, G.; Kaul, S.C.; Wadhwa, R.; Pati, P.K. Biotechnological interventions in *Withania somnifera* (L.) dunal. *Biotechnol. Genet. Eng. Rev.* **2015**, *31*, 1–20. [[CrossRef](#)]
21. Dhama, K.; Sharun, K.; Tiwari, R.; Dadar, M.; Malik, Y.S.; Singh, K.P.; Chaicumpa, W. COVID-19, an emerging coronavirus infection: Advances and prospects in designing and developing vaccines, immunotherapeutics, and therapeutics. *Hum. Vaccin Immunother.* **2020**, *16*, 1232–1238. [[CrossRef](#)]
22. Tiwari, R.; Latheef, S.K.; Ahmed, I.; Iqbal, H.M.N.; Bule, M.H.; Dhama, K.; Samad, H.A.; Karthik, K.; Alagawany, M.; El-Hack, M.E.A.; et al. Herbal Immunomodulators—A remedial panacea for designing and developing effective drugs and medicines: Current scenario and future prospects. *Curr. Drug Metab.* **2018**, *19*, 264–301. [[CrossRef](#)] [[PubMed](#)]
23. Calland, N.; Dubuisson, J.; Rouille, Y.; Seron, K. Hepatitis C virus and natural compounds: A new antiviral approach? *Viruses* **2012**, *4*, 2197–2217. [[CrossRef](#)]
24. Du, J.; He, Z.D.; Jiang, R.W.; Ye, W.C.; Xu, H.X.; But, P.P.H. Antiviral flavonoids from the root bark of *Morus alba* L. *Phytochemistry* **2003**, *62*, 1235–1238. [[CrossRef](#)]
25. Lugini, M.E.; Terlizzi, G.; Catucci, G.; Gilardi, M.E.; Maffei, G.G. The cranberry extract oximacro exerts in vitro virucidal activity against influenza virus by interfering with hemagglutinin. *Front. Microbiol.* **2018**, *9*, 1826.
26. Xu, H.X.; Ming, D.S.; Dong, H.; But, P.P. A new anti-HIV triterpene from *Geum japonicum*. *Chem. Pharm. Bull.* **2000**, *48*, 1367–1369. [[CrossRef](#)]
27. Sahuc, M.E.; Sahli, R.; Riviere, C.; Pene, V.; Lavie, M.; Vandeputte, A. Dehydrojuncusol, a natural phenanthrene compound extracted from *Juncus maritimus*, is a new inhibitor of hepatitis C virus RNA replication. *J. Virol.* **2019**, *93*, e02009–e02018. [[CrossRef](#)]
28. Cinatl, J.; Morgenstern, B.; Bauer, G.; Chandra, P.; Rabenau, H.; Doerr, H.W. Glycyrrhizin, an active component of liquorice roots, and replication of SARS-associated coronavirus. *Lancet* **2003**, *361*, 2045–2046. [[CrossRef](#)]
29. Lin, S.C.; Ho, C.T.; Chuo, W.H.; Li, S.; Wang, T.T.; Lin, C.C. Effective inhibition of MERS-CoV infection by resveratrol. *BMC Infect. Dis.* **2017**, *17*, 144. [[CrossRef](#)]
30. Elansary, H.O.; Szopa, A.; Kubica, P.; Ekiert, H.; Ali, H.M.; Elshikh, M.S.; Abdel-Salam, E.M.; El-Esawi, M.; El-Ansary, D.O. Bioactivities of Traditional Medicinal Plants in Alexandria. *Evid.-Based Complement. Altern. Med.* **2018**, 1463579, 13.
31. Wang, Z.; Song, Q.; Su, J.; Tang, W.; Song, J.; Huang, X.; An, J.; Li, Y.; Ye, W.; Wang, Y. Caffeic acid oligomers from *Mesona chinensis* and their in vitro antiviral activities. *Fitoterapia* **2020**, *144*, 104603. [[CrossRef](#)]
32. Liu, T.; Liu, X.; Li, W. Tetrandrine, a Chinese plant-derived alkaloid, is a potential candidate for cancer chemotherapy. *Oncotarget* **2016**, *7*, 40800–40815. [[CrossRef](#)]
33. Bailly, C. Cepharanthine: An update of its mode of action, pharmacological properties and medical applications. *Phytomedicine* **2019**, *62*, 152956. [[CrossRef](#)] [[PubMed](#)]
34. Zhang, C.H.; Wang, Y.F.; Liu, X.J.; Lu, J.H.; Qian, C.W.; Wan, Z.Y.; Yan, X.G.; Zheng, H.Y.; Zhang, M.Y.; Xiong, S.; et al. Antiviral activity of cepharanthine against severe acute respiratory syndrome coronavirus in vitro. *Chin. Med. J.* **2005**, *20*, 493–496.
35. Okamoto, M.; Ono, M.; Baba, M. Potent inhibition of HIV type 1 replication by an antiinflammatory alkaloid cepharanthine in chronically infected monocytic cells. *AIDS Res. Hum.* **1998**, *14*, 1239–1245. [[CrossRef](#)] [[PubMed](#)]
36. Mulinacci, N.; Innocenti, M.; Gallori, S.; Romani, A.; LaMarca, G.; Vincieri, F.F. Optimization of the chromatographic determination of polyphenols in the aerial parts of *Cichorium intybus* L. *Chromatographia* **2001**, *24*, 455–461. [[CrossRef](#)]
37. Ziamajidi, N.; Khagani, S.; Hassanzadeh, G.; Vardasbi, S.; Ahmadian, S.; Nowrouzi, A. Amelioration by chicory seed extract of diabetes-and oleic acid-induced non-alcoholic fatty liver disease (NAFLD)/non-alcoholic steato-hepatitis (NASH) via modulation of PPAR $\alpha$  and SREBP-1. *Food Chem. Toxicol.* **2013**, *58*, 198–209. [[CrossRef](#)]
38. Peng, Y.; Sun, Q.; Park, Y. The Bioactive Effects of Chicoric Acid As a Functional Food Ingredient. *J. Med. Food.* **2019**, *22*, 645–652. [[CrossRef](#)] [[PubMed](#)]
39. Araújo, C.C.; Leon, L.L. Biological activities of *Curcuma longa* L. *Mem Inst. Oswaldo Cruz.* **2001**, *96*, 723–728. [[CrossRef](#)] [[PubMed](#)]
40. Soni, V.K.; Mehta, A.; Ratre, Y.K.; Tiwari, A.K.; Amit, A.; Singh, R.P.; Sonkar, S.C.; Chaturvedi, N.; Shukla, D.; Vishvakarma, N.K. Curcumin, a traditional spice component, can hold the promise against COVID-19? *Eur. J. Pharmacol.* **2020**, *886*, 173551. [[CrossRef](#)]
41. Siviero, A.; Gallo, E.; Maggini, V.; Gori, L.; Mugelli, A.; Firenzuoli, F.; Vannacci, A. Curcumin, a golden spice with a low bioavailability. *J. Herb. Med.* **2015**, *5*, 57–70. [[CrossRef](#)]
42. Prakash, D.; Suri, S.; Upadhyay, G.; Singh, B.N. Total phenol, antioxidant and free radical scavenging activities of some medicinal plants. *Int. J. Food Sci. Nutr.* **2007**, *58*, 18–28. [[CrossRef](#)]
43. Sivaprabha, J.; Dharani, B.; Padma, P.R.; Sumathi, S. Induction of DNA damage by the leaves and rhizomes of *Curcuma amada* Roxb in breast cancer cell lines. *J. Acute Dis.* **2015**, *4*, 12–17. [[CrossRef](#)]
44. Mahadevi, R.; Kavitha, R. Phytochemical and pharmacological properties of *Curcuma amada*: A Review. *Int. J. Res. Pharm. Sci.* **2020**, *11*, 3546–3555. [[CrossRef](#)]
45. Gurung, P.; De, P. Spectrum of biological properties of cinchona alkaloids: A brief review. *J. Pharmacogn. Phytochem.* **2017**, *6*, 162–166.
46. O'Neill, E.J.; Termini, D.; Albano, A.; Tsiani, E. Anti-Cancer Properties of Theaflavins. *Molecules* **2021**, *26*, 987. [[CrossRef](#)]
47. Tong, T.; Liu, Y.; Kang, J.; Zhang, C.; Kang, S. Antioxidant Activity and Main Chemical Components of a Novel Fermented Tea. *Molecules* **2019**, *24*, 2917. [[CrossRef](#)] [[PubMed](#)]

48. Behl, T.; Sharma, A.; Sharma, L.; Sehgal, A.; Zengin, G.; Brata, R.; Fratila, O.; Bungau, S. Exploring the Multifaceted Therapeutic Potential of Withaferin A and Its Derivatives. *Biomedicines* **2020**, *8*, 571. [[CrossRef](#)]
49. Wang, G.; Xu, L.; Liu, W.; Xu, W.; Mu, Y.; Wang, Z.; Huang, X.; Li, L. New anti-inflammatory withanolides from *Physalis pubescens* fruit. *Fitoterapia* **2020**, *146*, 104692. [[CrossRef](#)] [[PubMed](#)]
50. Tan, J.; Liu, Y.; Cheng, Y.; Sun, Y.; Pan, J.; Guan, W.; Li, X.; Huang, J.; Jiang, P.; Guo, S.; et al. New withanolides with anti-inflammatory activity from the leaves of *Datura metel* L. *Bioorganic Chem.* **2020**, *95*, 103541. [[CrossRef](#)] [[PubMed](#)]
51. Jung, Y.; Lee, J.; Kim, H.K.; Moon, B.C.; Ji, Y.; Ryu, D.H.; Hwang, G.S. Metabolite profiling of *Curcuma* species grown in different regions using <sup>1</sup>HNMR spectroscopy and multivariate analysis. *Analyst* **2012**, *137*, 5597–5606. [[CrossRef](#)]
52. Kamazeri, T.S.A.T.; Samah, O.A.; Taher, M.; Susanti, D.; Qaralleh, H. Antimicrobial activity and essential oils of *Curcuma aeruginosa*, *Curcuma mangga*, and *Zingiber cassumunar* from Malaysia. *Asian Pac. J. Trop. Med.* **2012**, *5*, 202–209. [[CrossRef](#)]
53. Khaerunnisa, S.; Kurniawan, H.; Awaluddin, R.; Suhartati, S.; Soetjpto, S. Potential Inhibitor of COVID-19 Main Protease (M<sup>Pro</sup>) From Several Medicinal Plant Compounds by Molecular Docking Study. *Preprints* **2020**. [[CrossRef](#)]
54. Crosby, D.C.; Lei, X.Y.; Gibbs, C.G.; McDougall, B.R.; Robinson, W.E.; Reinecke, M.G. Design, synthesis, and biological evaluation of novel hybrid dicaffeoyltartaric/diketo acid and tetrazole-substituted lchicoric acid analogue inhibitors of human immunodeficiency virus type 1 integrase. *J. Med. Chem.* **2010**, *53*, 8161–8175. [[CrossRef](#)] [[PubMed](#)]
55. Hazuda, D.J.; Felock, P.; Witmer, M.; Wolfe, A.; Stillmock, K.; Grobler, J.A.; Espeseth, A.; Gabryelski, L.; Schleif, W.; Blau, C. Inhibitors of strand transfer that prevent integration and inhibit HIV-1 replication in cells. *Science* **2000**, *287*, 646–650. [[CrossRef](#)] [[PubMed](#)]
56. Reinke, R.A.; Lee, D.J.; McDougall, B.R. l-Chicoric acid inhibits human immunodeficiency virus type 1 integration in vivo and is a noncompetitive but reversible inhibitor of HIV-1 integrase in vitro. *Virology* **2004**, *326*, 203–219. [[CrossRef](#)] [[PubMed](#)]
57. Moshe, R.; Paul, O.; Igor, K. Cepharanthine: A review of the antiviral potential of a Japanese-approved alopecia drug in COVID-19. *Pharmacol. Rep.* **2020**, *72*, 1509–1516.
58. Ohashi, H.; Watashi, K.; Saso, W.; Shionoya, K.; Iwanami, S.; Hirokawa, T.; Shirai, T.; Kanaya, S.; Ito, Y.; Kim, K.S.; et al. Potential anti-COVID-19 agents, cepharanthine and nelfinavir, and their usage for combination treatment. *iScience* **2021**, *23*, 102367. [[CrossRef](#)] [[PubMed](#)]
59. Lan, J.; Ge, J.; Yu, J.; Shan, S.; Zhou, H.; Fan, S.; Zhang, Q.; Shi, X.; Wang, Q.; Zhang, L. Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor. *Nature* **2020**, *581*, 215–220. [[CrossRef](#)]
60. Walls, A.C.; Park, Y.J.; Tortorici, M.A.; Wall, A.; McGuire, A.T.; Veesler, D. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell* **2020**, *181*, 281–292. [[CrossRef](#)]
61. Deguchi, A. Curcumin targets in inflammation and cancer. *Endocr. Metab. Immune Disord. Drug Targets* **2015**, *15*, 88–96. [[CrossRef](#)]
62. Pulido-Moran, M.; Moreno-Fernandez, J.; Ramirez-Tortosa, C.; Ramirez-Tortosa, M. Curcumin and Health. *Molecules* **2016**, *21*, 264. [[CrossRef](#)] [[PubMed](#)]
63. Kim, Y.; Clifton, P. Curcumin, Cardiometabolic Health and Dementia. *Int. J. Environ. Res. Public Health* **2018**, *15*, 2093. [[CrossRef](#)] [[PubMed](#)]
64. Lelli, D.; Sahebkar, A.; Johnston, T.P.; Pedone, C. Curcumin use in pulmonary diseases: State of the art and future perspectives. *Pharmacol. Res.* **2017**, *115*, 133–148. [[CrossRef](#)] [[PubMed](#)]
65. Bright, J.J. Curcumin and autoimmune disease. *Adv. Exp. Med. Biol.* **2007**, *595*, 425–451.
66. Sordillo, P.P.; Helson, L. Curcumin Suppression of Cytokine Release and Cytokine Storm. A Potential Therapy for Patients with Ebola and Other Severe Viral Infections. *In Vivo* **2015**, *29*, 1–4.
67. Achan, J.; Talisuna, A.O.; Erhart, A.; Yeka, A.; Tibenderana, J.K.; Baliraine, F.N.; Rosenthal, P.J.; D'Alessandro, U. Quinine, an old anti-malarial drug in a modern world: Role in the treatment of malaria. *Malar. J.* **2011**, *10*, 144. [[CrossRef](#)]
68. Lipinski, C.A.; Lombardo, F.; Dominy, B.W.; Feeney, P.J. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv. Drug Deliv. Rev.* **1997**, *23*, 3–25, reprinted in *Adv. Drug Deliv. Rev.* **2001**, *46*, 3–26. [[CrossRef](#)]
69. Lipinski, C.A. Lead- and drug-like compounds: The rule-of-five revolution. *Drug Discov. Today Technol.* **2004**, *1*, 337–341.
70. Clark, D.E. Rapid calculation of polar molecular surface area and its application to the prediction of transport phenomena 1. Prediction of intestinal absorption. *J. Pharm. Sci.* **1999**, *88*, 807–814.
71. Chang, L.C.W.; Spanjersberg, R.F.; von Frijtag Drabbe, K.; Unzel, J.K.; Mulder-Krieger, T.; van den Hout, G.; Beukers, M.W.; Brussee, J.; Ijzerman, A.P. 2,4,6-Trisubstituted pyrimidines as a new class of selective adenosine A<sub>1</sub> receptor antagonists. *J. Med. Chem.* **2004**, *47*, 6529–6540. [[CrossRef](#)]
72. Veber, D.F.; Johnson, S.R.; Cheng, H.Y.; Smith, B.R.; Ward, K.W.; Kopple, K.D. Molecular properties that influence the oral bioavailability of drug candidates. *J. Med. Chem.* **2002**, *45*, 2615–2623. [[CrossRef](#)]
73. Andersson, S.; Antonsson, M.; Elebring, M.; Jansson-Lofmark, R.; Weidolf, L. Drug metabolism and pharmacokinetic strategies for oligonucleotide- and mRNA-based drug development. *Drug Discov. Today* **2018**, *23*, 1733–1745. [[CrossRef](#)] [[PubMed](#)]
74. Yu, J.; Zhou, Z.; Tay-Sontheime, J.; Levy, R.H.; Ragueneau-Majlessi, I. Intestinal Drug Interactions Mediated by OATPs: A Systematic Review of Preclinical and Clinical Findings. *J. Pharm. Sci.* **2017**, *106*, 2312–2325. [[CrossRef](#)]
75. Lynch, T.; Price, A. The effect of cytochrome P450 metabolism on drug response, interactions, and adverse effects. *Am. Fam. Physician* **2007**, *1*, 391–396.

76. Paz, S.; Vilasco, M.; Werden, S.J.; Arguello, M.; Joseph-Pillai, D.; Zhao, T.; Nguyen, T.L.; Sun, Q.; Meurs, E.F.; Lin, R. A functional C-terminal TRAF3-binding site in MAVS participates in positive and negative regulation of the IFN antiviral response. *Cell Res.* **2011**, *21*, 895–910. [[CrossRef](#)] [[PubMed](#)]
77. Gao, Y.; Yan, L.; Huang, Y.; Liu, F.; Zhao, Y.; Cao, L.; Wang, T.; Sun, Q.; Ming, Z.; Zhang, L.; et al. Structure of the RNA-dependent RNA polymerase from COVID-19 virus. *Science* **2020**, *368*, 779–782. [[CrossRef](#)]
78. Berman, H.M.; Battistuz, T.; Bhat, T. The protein data bank. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **2002**, *58*, 899–907. [[CrossRef](#)]
79. Giménez, B.G.; Santos, M.S.; Ferrarin, M.; Dos Santos Fernandes, J.P. Evaluation of blockbuster drugs under the rule-of-five. *Pharmazie* **2010**, *65*, 148–152.
80. Afonine, P.V.; Klaholz, B.P.; Moriarty, N.W.; Poon, B.K.; Sobolev, O.V.; Terwilliger, T.C.; Adams, P.D.; Urzhumtsev, A. New tools for the analysis and validation of cryo-EM maps and atomic models. *Acta Crystallogr. D Struct. Bio.* **2018**, *74*, 814–840. [[CrossRef](#)]
81. Pettersen, E.F.; Goddard, T.D.; Huang, C.C.; Couch, G.S.; Greenblatt, D.M.; Meng, E.C.; Ferrin, T.E. UCSF Chimera—A visualization system for exploratory research and analysis. *J. Comput. Chem.* **2004**, *25*, 1605–1612. [[CrossRef](#)]